






RESEARCH ARTICLE

Resistance of subarctic soil fungal and invertebrate communities to disruption of below-ground carbon supply

Thomas C. Parker¹  | Mathilde Chomel²  | Karina E. Clemmensen³  |
 Nina L. Friggens⁴  | Iain P. Hartley⁴  | David Johnson²  | Ilona Kater⁵  |
 Eveline J. Krab⁶  | Björn D. Lindahl⁶  | Lorna E. Street⁷  | Jens-Arne Subke⁸  |
 Philip A. Wookey⁸ 

¹Ecological Sciences, The James Hutton Institute, Aberdeen, UK; ²Department of Earth and Environmental Sciences, University of Manchester, Manchester, UK; ³Swedish University of Agricultural Sciences, Department of Forest Mycology and Plant Pathology, Uppsala, Sweden; ⁴Geography, College of Life and Environmental Sciences, University of Exeter, Exeter, UK; ⁵Department of Geography and Scott Polar Research Institute, University of Cambridge, Cambridge, UK; ⁶Swedish University of Agricultural Sciences, Department of Soil and Environment, Uppsala, Sweden; ⁷School of Geosciences, University of Edinburgh, Edinburgh, UK and ⁸Biological and Environmental Sciences, School of Natural Sciences, University of Stirling, Stirling, UK

Correspondence

Thomas C. Parker

Email: thomas.parker@hutton.ac.uk

Funding information

Natural Environment Research Council,
Grant/Award Number: NE/P002722/1
and NE/P002722/2

Handling Editor: Franciska de Vries

Abstract

1. The supply of recent photosynthate from plants to soils is thought to be a critical mechanism regulating the activity and diversity of soil biota. In the Arctic, large-scale vegetation transitions are underway in response to warming, and there is an urgent need to understand how these changes affect soil biodiversity and function.
2. We investigated how abundance and diversity of soil fungi and invertebrates responded to a reduction in fresh below-ground photosynthate supply in treeline birch and willow, achieved using stem girdling. We hypothesised that birch forest would support greater abundance of ectomycorrhizal (ECM) fungal species and fauna than willow shrubs, and that girdling would result in a rapid switch from ECM fungi to saprotrophs as canopy supply of C was cut, with a concomitant decline in soil fauna.
3. Birch forest had greater fungal and faunal abundance with a large contribution of root-associated ascomycetes (ericoid mycorrhizal fungi and root endophytes) compared to willow shrub plots, which had a higher proportion of saprotrophs and, contrary to our expectations, ECM fungi. Broad-scale soil fungal and faunal functional group composition was not significantly changed by girdling, even in the third year of treatment. Within the ECM community, there were some changes, with genera that are believed to be particularly C-demanding declining in girdled plots. However, it was notable how most ECM fungi remained present after 3 years of isolation of the below-ground compartment from contemporary photosynthate supply.
4. *Synthesis.* In a treeline/tundra ecosystem, distinct soil communities existed in contrasting vegetation patches within the landscape, but the structure of these communities was resistant to canopy disturbance and concomitant reduction of autotrophic C inputs.

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. *Journal of Ecology* published by John Wiley & Sons Ltd on behalf of British Ecological Society.

KEYWORDS

canopy disturbance, fungi, metabarcoding, mycorrhizal fungi, Plant–soil (below-ground) interactions, soil fauna, subarctic

1 | INTRODUCTION

The climate is changing in the Arctic faster than anywhere else on earth (Mudryk et al., 2019), causing complex changes and feedbacks within terrestrial ecosystems (Post et al., 2009; Wookey et al., 2009). One such prominent change in parts of the Arctic is the increase in productivity and associated compositional shifts in vegetation cover (Elmendorf, Henry, Hollister, Bjork, Boulanger-Lapointe, et al., 2012). Deciduous shrubs are documented to have increased in cover and range in many areas, and treelines are shifting northward and up-slope, in line with shifts in climate and other factors (Myers-Smith et al., 2011; Rees et al., 2020). Above-ground, shrub or treeline expansion is reflected in important changes in primary productivity, reflectance and microclimate (Myers-Smith et al., 2011). Below-ground, the implications of vegetation change are less well understood, but there is potential for major biogeochemical feedbacks (Parker, Thurston, et al., 2021). Greening and productivity trends across the Arctic are being punctuated and even counteracted by increasingly frequent 'browning' events and trends in some places (Myers-Smith et al., 2020; Phoenix & Bjerke, 2016). Parts of tundra and treeline forests face increased incidents of tundra fires (Bret-Harte et al., 2013), caterpillar outbreaks (Dahl et al., 2017; Jepsen et al., 2008), extreme winter warming events (Treharne et al., 2020) and range shifts of major canopy herbivores (Tape et al., 2016), all of which can contribute to reductions in ecosystem productivity.

'Greening' and 'browning' of the Arctic could represent fundamental changes to how ecosystems function in this biome. Greening is characterised by a shift from plants that form ericoid mycorrhiza (Hobbie & Hobbie, 2006; Read et al., 2004) to more productive trees and deciduous shrubs that associate with ectomycorrhizal (ECM) fungi, whereas browning events may reduce the C supply to fungal symbionts (Parker et al., 2017, 2020; Saravesi et al., 2015). The resulting change in below-ground productivity (Sloan et al., 2013) and mycorrhizal association (Hobbie et al., 2009) will alter the supply of C to soil food webs (e.g. fungi and fauna), the community composition of which is vital for regulating the turnover of C in soils (Handa et al., 2014). Mycorrhizal fungi are key components of food webs because they have a fundamental role in regulating soil organic matter (SOM) storage and turnover (Frey, 2019), and are phylogenetically and functionally diverse (Hibbett et al., 2000). These fungi represent a critical physical and energetic link between plants and other components of soil food webs and soil processes, because they are recipients of plant-derived C and sometimes also decomposers of SOM. Some species are known 'as potent decomposers', 'mining' organic N by producing oxidative enzymes (Bödeker et al., 2014; Lindahl et al., 2021), while others are better adapted for 'scavenging' of mineral N with a less direct effect on SOM (Zak et al., 2019). Hence, it

is important to determine the factors driving food web community assembly to better understand the implications of plant community change for ecosystem functioning (Clemmensen et al., 2021).

Browning events could restructure ECM fungal communities by reducing the amount of C that can be allocated below-ground, thereby adjusting the competitive balance of 'C-demanding' versus less demanding fungi (Saikkonen et al., 1999). Trees at the relatively productive subarctic birch treeline are associated with ECM fungi that specialise in rapid mobilisation, or 'mining', of organic N from the soil (Bödeker et al., 2014; Clemmensen et al., 2021). In particular, the *Cortinarius* genus, which consists of cord-forming Agaricomycetes, has been linked to oxidation of organic matter by production of manganese peroxidases in low-fertility boreal forests (Bödeker et al., 2014; Lindahl et al., 2021; Pérez-Izquierdo et al., 2021). Conversely, other fungi that form 'short distance' or 'contact' mycorrhizal morphotypes (with hyphae concentrated close to the root tip [Agerer, 2001]) may be favoured under more restrictive C supply from their hosts, especially after a browning event (Saikkonen et al., 1999). Indeed, defoliation of subarctic canopies by geometrid moth caterpillars ('autumnal' and 'winter' moths of the genera *Epirrita* and *Operophtera*, respectively) has been found to shift mycorrhizal communities from medium- and long-distance exploration types (ETs) to contact types (Parker et al., 2017; Saravesi et al., 2015). In addition, we have already demonstrated that stem girdling (disconnection of the phloem) results in a large reduction in the production of extra-radical mycelium in treeline mountain birch forest, but not in tall willow plots, which had lower baseline rates of hyphal production (Parker et al., 2020). Potentially, these results are related to a higher allocation of C to ECM fungi in the birch forest. Therefore, we predicted that girdling would result in selective reduction of ECM species in the fungal community, in particular of more C-demanding species, and most markedly in the birch forest.

The traditional view represented in food web models is that C enters soil fauna predominantly from plant litter inputs, either via bacterial-based or fungal-based energy channels (Hunt et al., 1987). However, there is an increasing evidence that C from recent photosynthate is a major source of energy for soil fauna (Chomel et al., 2019; Eissfeller et al., 2013; Gilbert et al., 2014; Goncharov et al., 2016; Pollierer et al., 2007; Ruf et al., 2006). Mycorrhizal fungi are likely to be a key pathway for C delivery to microbivore fauna, such as microarthropods (e.g. Collembola and oribatid mites), because the substantial flux of recent photosynthate through extra-radical mycelium in forests (Heinemeyer et al., 2007; Högberg et al., 2001, 2008). Indeed, experiments in laboratory model systems have demonstrated transfer of recent photosynthate to Collembola (Kanters et al., 2015) and that the presence of ECM fungi may

modify the composition of faunal communities (Setälä, 2000; Setälä et al., 1999). Despite these findings from laboratory experiments, it is unclear whether changes in the belowground supply of recent photosynthate and associated alterations of fungal communities lead to changes in microarthropod communities in the field.

Soil biodiversity under shrub willows (*Salix* spp.), which are particularly prevalent in wetter areas of the tundra (Elmendorf, Henry, Hollister, Bjork, Bjorkman, et al., 2012; Myers-Smith et al., 2011; Tape et al., 2006), is also poorly characterised. The fungal and faunal communities associated with *Salix* shrubs may be distinct from those of *Betula* shrubs or forest (Clemmensen et al., 2021) because of a tendency of *Salix* to grow in moister areas prone to seasonal flooding, potential anoxia, and a higher influx of aeolian and snow-borne mineral nutrients (Nadelhoffer et al., 1991; Sturm et al., 2005). Relatively, high nutrient availability may reduce the need for *Salix* shrubs to invest in ECM fungi, or at least in C-demanding ECM fungi that specialise in 'N-mining' (Clemmensen et al., 2021). Reduced competition from ECM fungi may favour proliferation of saprotrophic fungi when N is more available (Kyaschenko et al., 2017). Lower ECM hyphal production was measured in willow soils compared to drier mountain birch soils (Parker et al., 2020), and we therefore expect a smaller proportion of ECM fungal species in willow plots. Consequently, microarthropods, who may depend on recent photosynthate delivery by ECM fungi and/or on their hyphal (necro)mass as a food source (Setälä, 2000; Setälä et al., 1999), are expected to be less abundant under willow than in birch soils, particularly in deeper soils where mycorrhizal fungi dominate in this system (Clemmensen et al., 2021).

The objective of our study is to characterise soil fungal and microarthropod communities in mountain birch forest (*Betula pubescens*) and tall willow stands (*Salix* spp.); two important subarctic and arctic vegetation types that are documented to be expanding in range and cover at high latitudes (Myers-Smith et al., 2011; Rees et al., 2020). We also documented the response of soil communities to a stem girdling treatment, to test their dependence on the delivery of recently fixed autotrophic C. We hypothesised that (H1) willow and birch would support distinct soil communities, reflecting differences in host and soil conditions. Specifically, we predict greater abundance of ECM fungi and microarthropods in birch plots, reflecting greater overall mycelium production and drier conditions. In a preceding study on the same experiment (Parker et al., 2020), we observed a large reduction in soil CO₂ efflux in both treeline mountain birch forest and willow shrub stands in response to girdling; therefore, we here further hypothesised that (H2) disruption of the supply of recently fixed photosynthate to the rhizosphere would cause a shift in soil fungal communities with saprotrophs replacing ECM fungi and a reduction in microarthropods in deeper organic soil layers, where ECM fungi are dominant. Based on previous findings of a large decrease in hyphal production in birch plots, but not in willow plots (Parker et al., 2020), we further hypothesised (H3a) that girdling would result in a larger decline in ECM species (H3b) particularly of medium- and long-distance ETs in birch forest compared with willow shrubs.

2 | MATERIALS AND METHODS

2.1 | Site selection and experimental design

A girdling experiment, outlined in detail by Parker et al. (2020), was carried out in a permafrost-free forest-tundra ecotone 4–5 km south of the Abisko Scientific Research Station, Sweden (68°18'N 18°49'E, ~600m a.s.l.). Briefly, six pairs of plots in mountain birch forest (*Betula pubescens* with a dense ericaceous understorey of primarily *Empetrum nigrum* and *Vaccinium* spp.) and five pairs of willow thickets (*Salix lapponum*) were located across a 0.88 km² area. Average soil and canopy characteristics did not vary significantly between control and girdled plots prior to girdling (Parker et al., 2020), and one of each pair was girdled in June 2017. During girdling, the bark and phloem were removed around the circumference of all birch or willow stems, resulting in a disruption of the transport of photosynthate from canopy to roots. Birch plots had a circular area with a radius of 10 m and willow plots had a radius of 2 m (with a trenched perimeter to prevent root ingrowth from adjacent plants). Re-sprouting shoots from below the girdle-line were removed whenever observed during the experiment. Birch and willow plants retained leaves until natural senescence in 2017, and all birch trees produced leaves above the girdle-line in spring 2018. However, leaf production in girdled birch canopies mostly failed in 2019 and in girdled willow shrub canopies in both 2018 and 2019.

2.2 | Soil sampling and DNA amplification for analyses of fungal communities

The organic soil horizon (O horizon) was sampled on 3 August 2017, 1 August 2018 and 5 August 2019. For each plot (22 in total), nine 3.8 cm diameter soil cores were collected; in the birch plots, cores were taken in a grid across a 3×3 m central area (corresponding to the central area in which soil CO₂ efflux, root and mycelium production measurements were taken; Parker et al., 2020). In the willow plots, the nine cores were distributed evenly across the plot area within the trenched perimeter. The uppermost litter layer, as well as the mineral horizons underlying the organic horizon (clearly identifiable at these sites), was removed from cores immediately after coring. The nine cores from each plot were homogenised and pooled within 6 h of sampling, coarse roots (>2 mm diameter) were removed and samples were frozen at -20°C until further analysis. Some birch forest samples from 2019 were lost, resulting in fewer replicates for that year, and 59 samples (out of 66 plots) went forward for analysis. A further homogenisation of the pooled soil samples was carried out in the laboratory using a custom-built large grinder that breaks up soil cores within 1 s, using rotating blades, while maintaining them in a frozen state.

Soil sub-samples (approximately 10 g) were freeze-dried and ball milled to a fine powder. DNA was extracted from a 50 mg sub-sample using the NucleoSpin Soil Kit (Macherey-Nagel). ITS2 markers were amplified using the fungal gITS7 forward (Ihrmark et al., 2012) and

reverse primer mix of 3/4 of ITS4 (for general eukaryotes [White et al., 1990]) and 1/4 of ITS4arc (adapted for Archaeorhizomycetes [Sterkenburg et al., 2018]) with minimal cycle numbers (51 samples were amplified with 23 cycles, 5 samples at 21 cycles and 3 at 25 cycles) to minimise biases in the community data (Castaño et al., 2020). PCRs were run in duplicates per sample, with 50 µl in each reaction containing the following reactants: Approximately 25 ng of DNA template, 0.2 mM dNTPs, 0.75 mM MgCl₂, 1.25 units of DreamTaq polymerase in its buffer (ThermoFisher) and primer concentrations of 0.5 µM of gITS7, 0.3 µM of ITS4 and 0.1 µM of ITS4arc. The PCR cycling conditions were as follows: 95°C for 5 minutes, then 21–30 cycles (depending on the sample) of 95°C for 30s, 56°C for 30s, 72°C for 30s and 7 min at 72°C. The duplicate PCR products from each sample were pooled and purified using AMPure magnetic beads (Beckman Coulter), concentrations measured with Qubit (Invitrogen) and equal DNA amounts mixed into a single pool. The pool was further cleaned using the E.Z.N.A CyclePure kit (Omega) and sequenced with the Pacific Biosciences SMRT Sequel technology after adaptor ligation (Castaño et al., 2020) by SciLifeLab NGI.

2.3 | Bioinformatic processing and fungal taxonomic identification

We used a community metabarcoding approach on the pooled DNA samples to identify distinct taxonomic groups at species level, which we now term 'species hypotheses' (SHs). Raw sequences were quality filtered and clustered into sequence clusters at an approximate species level, using the SCATA pipeline (scata.mykopat.slu.se; Ihrmark et al., 2012). Sequences were first quality filtered (requiring a mean quality score of 20 or higher) and screened for primer sequences (90% similarity required) and identification tags, which were removed. Sequences were then pairwise compared using USEARCH (Edgar, 2013) and clustered into SHs using a single linkage algorithm with a 98.5% similarity cut-off (Lindahl et al., 2013). Plant sequences were removed. Sequencing produced 181,560 reads with 100,282 passing quality control. Only SHs that contributed >1% of the fungal sequences in at least one sample were retained. In the final analysis, 38,659 reads were analysed in a matrix of 521 SHs in 59 samples. Three reference datasets (Swedish Soil Inventory: [Lindahl et al., 2021], Swedish Boreal forest: [Clemmensen et al., 2015] and nearby Abisko database: [Clemmensen et al., 2021]) were included in the clustering process to aid taxonomic classifications. These were verified against the UNITE database using 98% similarity for species-level identification. SHs were assigned to functional guilds (ECM fungi, moulds, litter saprotrophs and root-associated ascomycetes) based on the FungalTraits database (Pöhlme et al., 2020). The root-associated ascomycetes included ericoid mycorrhizal fungi and root endophytes but was deliberately left as an unspecified group due to the often broad or undefined ecology of many species (Kohout, 2017). ECM SHs were assigned to one of five well-defined ETs (Agerer, 2001) based on established datasets (Tedersoo & Smith, 2013). Fungal community composition and ITS

copy numbers (see next section) data were stored at the NERC EIDC (Parker et al., 2022a).

2.4 | ITS region quantitative PCR

Copy numbers of the fungal ITS2 region were estimated by quantitative PCR (qPCR) using the IQ SYBR green supermix on an iQ5 real-time PCR system (Bio-Rad). The 20 µl reactions contained approximately 5 ng of DNA template, 0.1% bovine serum albumin and the ITS4/ITS4arc and gITS7 primers (as above). The thermal cycling conditions were 95°C for 5 min, then 40 cycles of 95°C for 15 s, 56°C for 30s, 72°C for 40s and 78°C for 5 s, with the fluorescent signal acquired at the last step of each cycle. PCR inhibition tests where known plasmid copy numbers were amplified with M13 primers (pGEM-T plasmid) showed no significant inhibition by the sample extracts. Standard curves for quantification were obtained by serial dilutions of linearised plasmids containing the ITS2 marker. The relative abundance of each SH and functional guild (which is well preserved during PCR cycles and PacBio sequencing; Castaño et al., 2020) was multiplied by total fungal ITS copy numbers (after correcting for amplification of non-fungal markers based on sequencing results) to estimate copy numbers for each SH and guild in each sample.

2.5 | Soil mesofauna sampling and extraction

Soils were sampled across all plots for microarthropods (Collembola and mites) on July 29th 2019. Within each plot, three 4.5 cm diameter soil cores were taken to the depth of the soil horizon (until rock or last clasts) within the central 3×3 m area of the birch plots, and across the willow plots. The mineral horizon of each soil core was removed and discarded, while the litter horizon was retained and pooled separately for each plot. Cores were inserted into tight fitting plastic rings to maintain soil structure for optimal extraction efficiency. The soil and litter samples were extracted for soil fauna using a Tullgren funnel fauna extractor (Van Straalen & Rijninks, 1982). In all, 22 pooled soil and 22 pooled litter samples were placed at random under individual heat lamps (to avoid spatial biases in extraction efficiency) and left for 10 days to extract fauna into ethanol. At the end of the extraction, the soil and litter samples were oven-dried for 24 h at 70°C and weighed to determine soil dry weights. The fauna samples were preserved in 70% ethanol prior to identification and quantification. Extracted fauna were counted and identified under a dissecting microscope to species level for Collembola using the key of Hopkin (2007) and to order level for Acari (Oribatid, Mesostigmatid, Prostigmatid). Other invertebrates were separated at higher taxonomic levels (e.g. Diptera, Coleoptera, Araneae). Soil mesofauna data were stored at the NERC EIDC (Parker et al., 2022b).

To facilitate trait analyses needed for testing H2, Collembola species were assigned trait values from 1 to 6 for soil vertical life-form

and moisture preference according to Kuznetsova (2003) and maximum body length according to Fjellberg (2007). Community-weighted mean (CWM) values were calculated for each trait according to

$$\text{CWM}_j = \sum_{k=1}^{n_j} A_{kj} \times \text{FT}_{kj},$$

where in the community in sample j , n_j is the number of species samples, A_{kj} is the relative abundance of species k and FT_{kj} is the functional trait of species k (Krab, Van Schrojenstein Lantman, et al., 2013). CWMs were calculated for organic soil and litter horizons separately.

2.6 | Statistical analyses

Total fungal ITS copy numbers were compared between birch and willow plots using a linear mixed effects model (the `lme` function from the `nlme` package in R; Pinheiro et al., 2017) with vegetation type, sample year and girdling treatment as fixed effects, and plot nested within plot pair as a random intercept term. Linear mixed effects models were used to test the effect of vegetation type, girdling treatment and soil horizon depth on mite and Collembola abundances and CWM vertical preference, moisture preference and body length of collembola. Plot nested within plot pair was used as random intercept term for soil fauna models. Models were simplified by removing non-significant three-way interaction terms to increase statistical power to test fixed factors. The effect of each factor in the final model was assessed relative to the null model (intercept only) by likelihood ratio tests (Crawley, 2007).

Differences and trends in fauna and fungal community composition were visualised by non-metric multidimensional scaling based on a Bray–Curtis dissimilarity matrix using the `metaMDS` function of the `VEGAN` package in R (Oksanen, 2013). The effect of vegetation type, girdling treatment and sampling year, and their interactions,

on fungal community composition (functional guild or SH) was assessed by multivariate analysis of copy numbers assigned to each fungal group. The `manyglm` function of the `MVABUND` package in R V 4.0.0 (Wang et al., 2012) was used to test the effect of the above factors on the multivariate copy number data of fungal SHs, guilds, ECM genera and ECM ETs using a Poisson distribution (which was found to fit the mean–variance assumption of the analysis). The role of different factors was compared using the ANOVA function, which applied Wald statistics following 999 PIT-trap resamplings (Warton et al., 2017). Models were simplified by removing interaction terms if they were not significant ($p < 0.05$). Furthermore, to assess the effects on individual functional groups within each test, additional univariate tests were carried out within the `manyglm` function.

3 | RESULTS

Soil in birch forest had a greater fungal abundance (ITS copies mg OM^{-1}) than willow plots ($F_{1,9} = 5.7$, $p = 0.043$), with no significant variation between years ($F_{1,9} = 0.9$, $p = 0.33$). The differences in fungal abundance between birch and willow plots were primarily accounted for by a greater abundance of root-associated ascomycetes in the birch plots, particularly ericoid mycorrhizal genera such as *Hyaloscypha*. Willow plots contained slightly more ECM fungi (Table S1, $\text{Wald}_{1,51} = 2.4$, $p = 0.02$), but a similar abundance of litter saprotrophs and moulds. The differences in abundance of root-associated ascomycetes and ECM fungi between birch and willow plots were linked to a statistically significant difference in fungal community composition at the functional guild level (Table S1, $\text{Wald}_{1,51} = 8.3$, $p < 0.001$). The fungal community of birch plots contained 60% root-associated ascomycetes (Figure 1) with litter saprotrophs as the second most abundant guild (15%), followed by ECM fungi (9.5%) and moulds (4.5%). Willow plots had a more even abundance of the four guilds (Figure 1), with root-associated ascomycetes contributing, on average, 29% of the fungal community, and litter

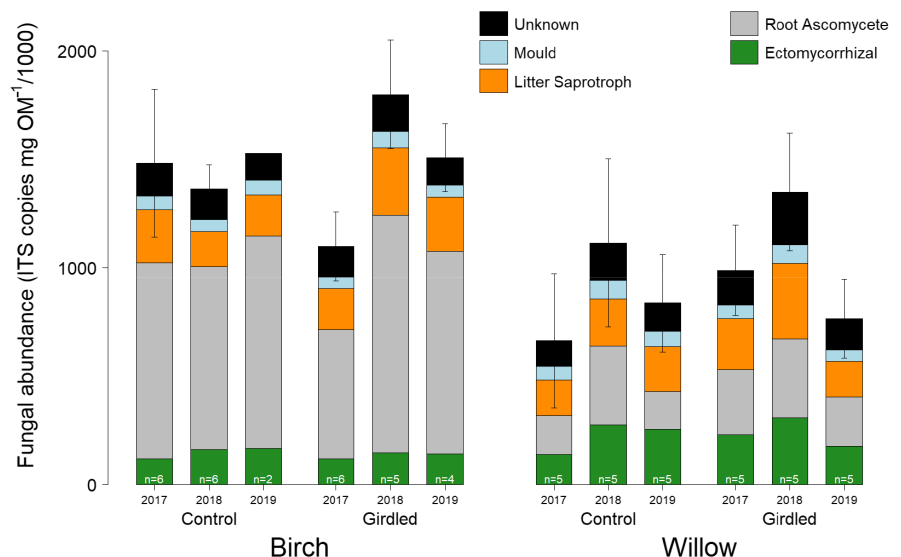


FIGURE 1 Fungal abundance partitioned into four functional guilds over the years 2017–2019 in soil from girdled and control plots of birch and willow at a subarctic treeline. Error bars represent ± 1 standard error of the mean total copy numbers. Birch control plots in 2019 do not have an error estimate because $n = 2$; replicate numbers for each bar are indicated at the base of the figure.

saprotrophs, ECM fungi and moulds contributing 23%, 22% and 6.8%, respectively (Figure 1).

At the level of individual species, community composition of both fungi (Figure 2a, Table S2, $Wald_{1,51} = 48.5$, $p < 0.001$) and soil fauna (Figure 2b, Table S3, $Wald_{1,20} = 6.608$, $p < 0.001$) contrasted significantly between the drier and mycelium-rich birch soils and the wetter, more nitrogen (N)-rich willow soils. Fungal communities in birch forest soils were also less variable than fungal communities in willow soils.

Across both birch and willow plots, girdling induced no significant change in total fungal abundance (Figure 1, $F_{1,39} = 1.7$, $p = 0.21$), abundance of different fungal guilds (Figure 1, Table S1, $Wald_{1,46} = 2.3$, $p = 0.37$) or fungal community composition at the species level (Figure 2a, Table S2, $Wald_{1,46} = 22.3$, $p = 0.31$). The abundance of litter saprotrophs and moulds did not increase relative to ECM fungi in any of the sampling years up to 26 months after girdling. Furthermore, there was no significant interaction between vegetation type and girdling treatment on total fungal abundance ($F_{1,39} = 1.3$, $p = 0.27$), abundance of different fungal guilds ($Wald_{1,47} = 1.6$, $p = 0.69$) or fungal community composition ($Wald_{1,47} = 16.0$, $p = 0.48$). Thus, the community composition and guild abundances did not change significantly more in birch plots than willow plots in response to girdling.

Birch and willow plots had significantly different communities of ECM fungi at the genus level (Figure 3, Table S4, $Wald_{1,51} = 11.9$, $p = 0.001$), with higher abundance of *Cenococcum*, *Entoloma*, *Inocybe*, *Russula* and *Tomentella* (Figure 5, Table S5) as well as more medium distance-smooth ETs (Figure 6, Table S6) in the willow plots. Overall, there was a small but statistically significant effect of girdling

on the genus composition among ECM fungi (Figure 3, Table S4, $Wald_{1,50} = 6.2$, $p = 0.034$) with reductions in *Piloderma*, *Entoloma* and *Cortinarius* and an increase in *Pseudotomentella* ($p < 0.1$, Figure 3, Table S4). Despite the reductions in *Piloderma* and *Cortinarius* in response to girdling, medium distance-fringe types showed a less clear reduction, whereas medium distance-smooth types responded negatively ($p = 0.02$, Figure 4, Table S5). There were no detectable differences in ECM community composition between years and no significant interaction between girdling and vegetation type (Figure 3, Figure 4, Tables S4 and S5, $Wald_{1,47} = 3.7$, $p = 0.42$), meaning that the response of ECM communities to girdling was similar in birch and willow plots.

There was greater abundance of mites and Collembola in birch plots compared with willow plots, which was primarily the result of a disproportionately higher density of these faunal groups in the organic horizon. Collembola in birch plots were more stratified according to vertical preference, with typically deeper dwelling species found in high abundances in the organic horizon and shallow dwelling species mainly found in the litter, whereas communities of the organic and litter horizons in willow were more similar. Collembola communities in birch and willow had similar moisture preference, but, as with vertical preference, there was greater stratification with depth in the birch plots, with drought-sensitive species dwelling preferably in the organic horizon and drought-tolerant species in the litter horizon. Willow plots typically contained collembola with larger body size in both the litter and organic horizons (Figure 6, Table S6). There was no statistically significant girdling effect on the abundance of mites or Collembola, or on any collembolan traits (Figures 5 and 6, Table S6).

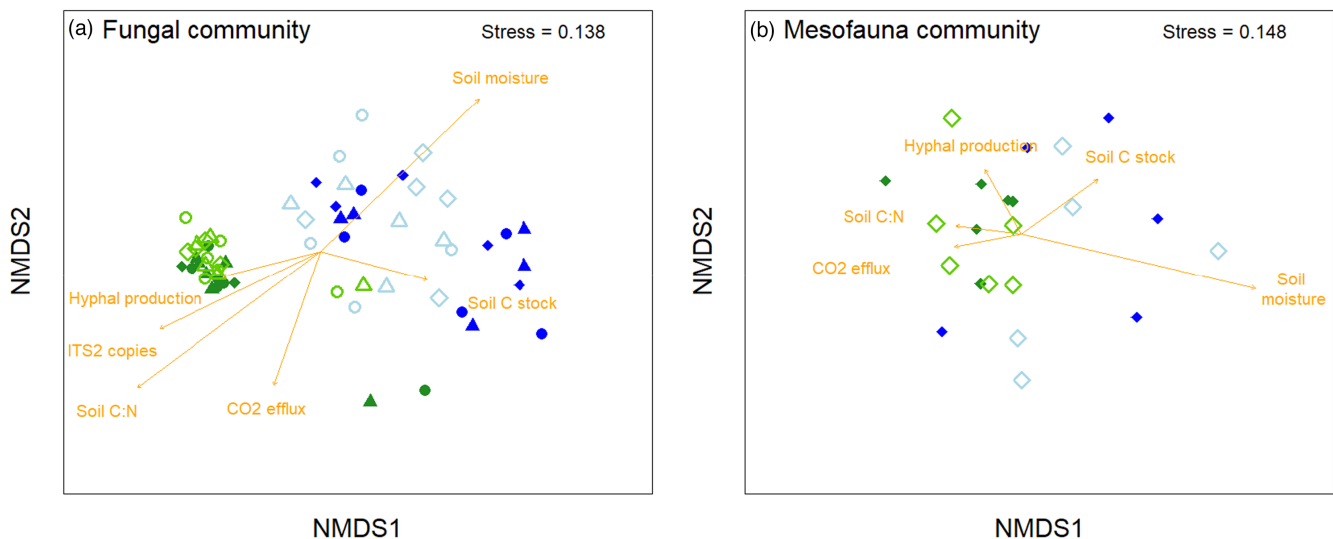


FIGURE 2 Nonmetric multidimensional scaling (NMDS) ordination of (a) fungal community composition in the 59 organic horizon samples across vegetation type, treatment and year and (b) soil fauna community composition in the organic horizon from 2019 at a subarctic treeline. Environmental vectors (orange) in (a) are based on data from Parker et al. (2020) and show the degree and direction of correlation of measured variables with fungal community composition; vectors in (b) are based on new data collected in 2019 using the same protocols as Parker et al. (2020).

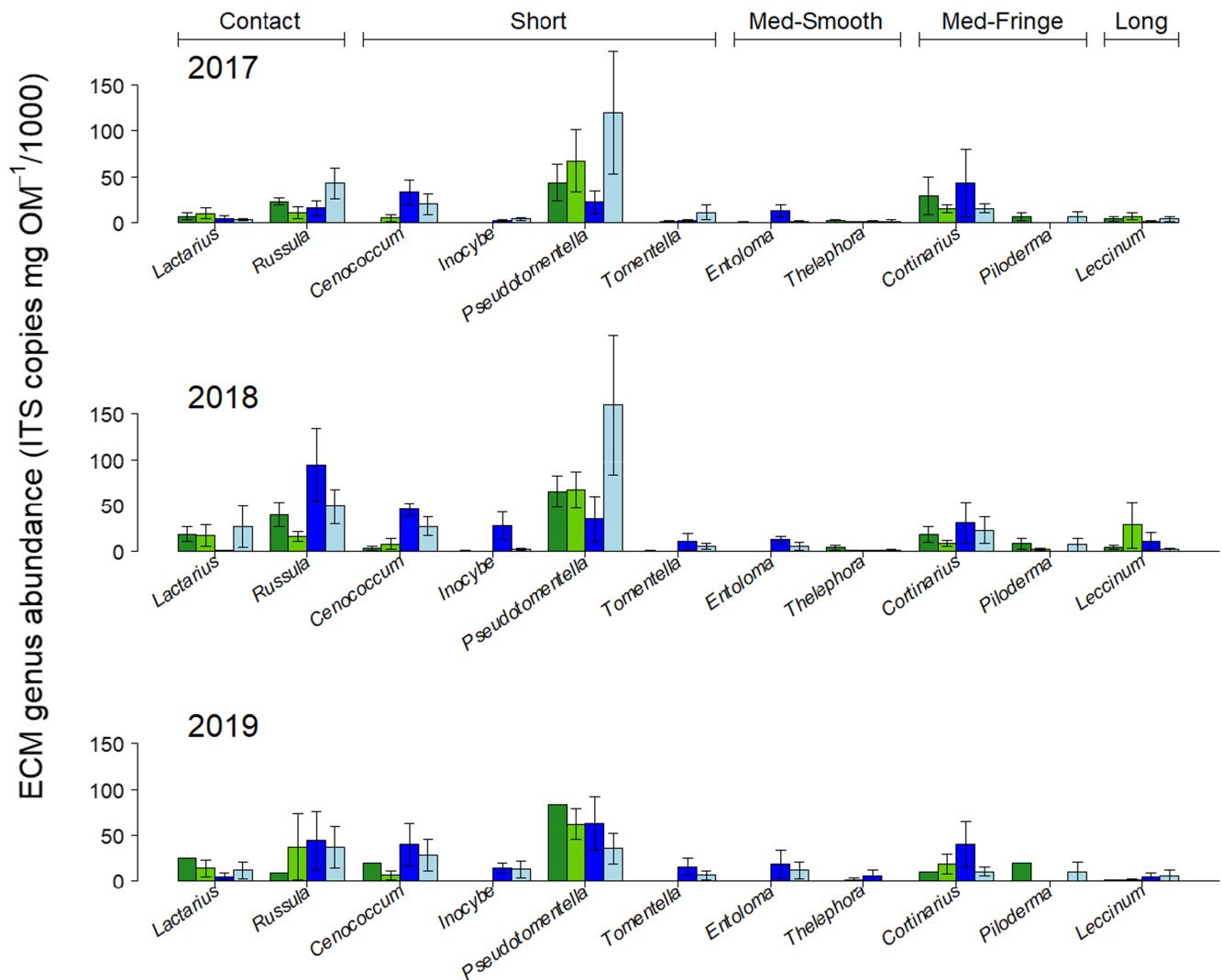


FIGURE 3 Abundance of ectomycorrhizal (ECM) genera in non-girdled (dark shade) and girdled (light shade) plots of birch (green) and willow (blue) at a subarctic treeline. Different panels show the three sampling years. Genera are grouped according to their exploration types (ETs), with short-to-long ranging strategies ordered from left to right. Sample sizes for each group per year are indicated in Figures 1 and 4. Error bars signify ± 1 SE of the mean; birch control plots in 2019 do not have an error estimate because $n = 2$.

4 | DISCUSSION

4.1 | The abundance and diversity of soil fungi and fauna in treeline forest and shrubs are resistant to declines in below-ground allocation of photosynthates

We found that fungal and faunal soil communities in two distinct treeline and tundra ecosystems were resistant to cessation of below-ground transfer of C from the phloem. Girdling completely severed photosynthate transport between canopy and roots, but even after 2 years, there was no significant restructuring of the soil communities, neither among fungi or collembolan species, nor among fungal or microarthropod functional groups. Informed by a nearby trenching and root ingrowth experiment (Clemmensen et al., 2021), as well as trenching and clearcutting experiments from further south (Kohout et al., 2018;

Sterkenburg et al., 2018, 2019), we hypothesised (H2) that there would be a replacement of ECM fungi by saprotrophic fungi and a reduction in soil fauna abundance in the organic soil. However, we observed that the proportion of ECM and free-living saprotrophic fungi in the community was maintained after girdling. Furthermore, faunal communities did not restructure or decline in abundance. Our results are even more surprising given that we previously observed large reductions in response to girdling both in soil CO₂ efflux in the same experiment and in hyphal production in the birch plots (Parker et al., 2020).

4.2 | Resistance of fungal and faunal community structure to girdling

The apparent stability of the soil communities in treeline and tundra shrub ecosystems after a disruption to supply of C belowground

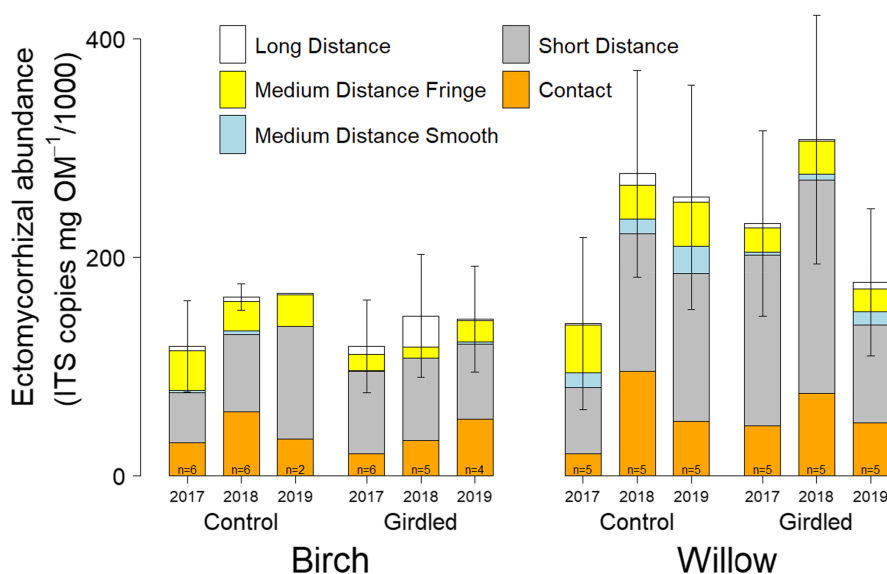


FIGURE 4 Abundance of ectomycorrhizal (ECM) fungi partitioned into five exploration types (ETs) over the years 2017–2019 in girdled and control plots of birch and willow at the subarctic treeline. Error bars represent ± 1 standard error of the mean total copy numbers. Birch control plots in 2019 do not have an error estimate because $n = 2$.

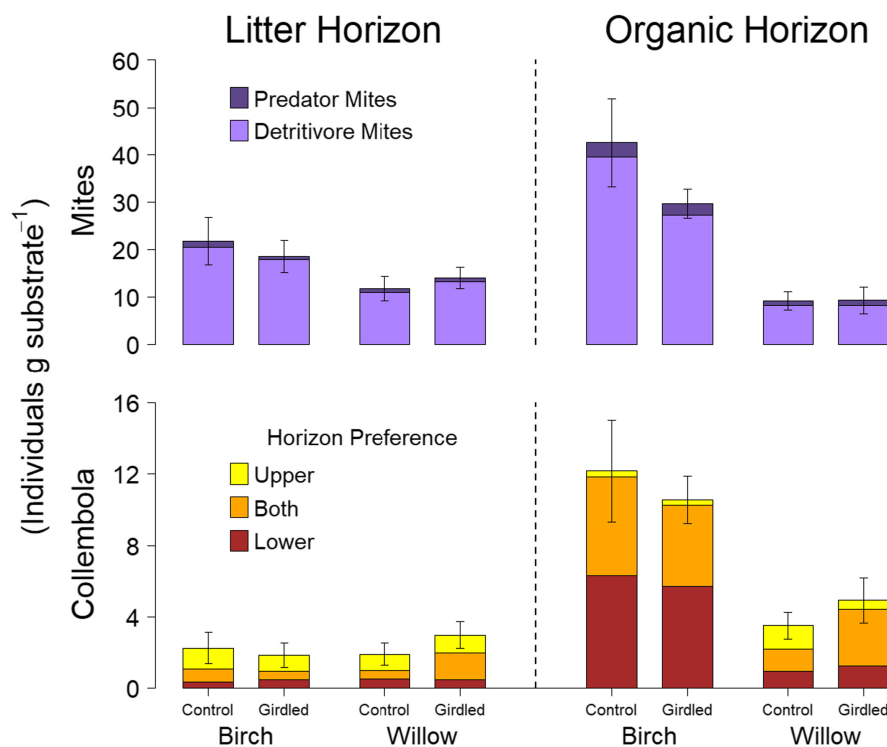


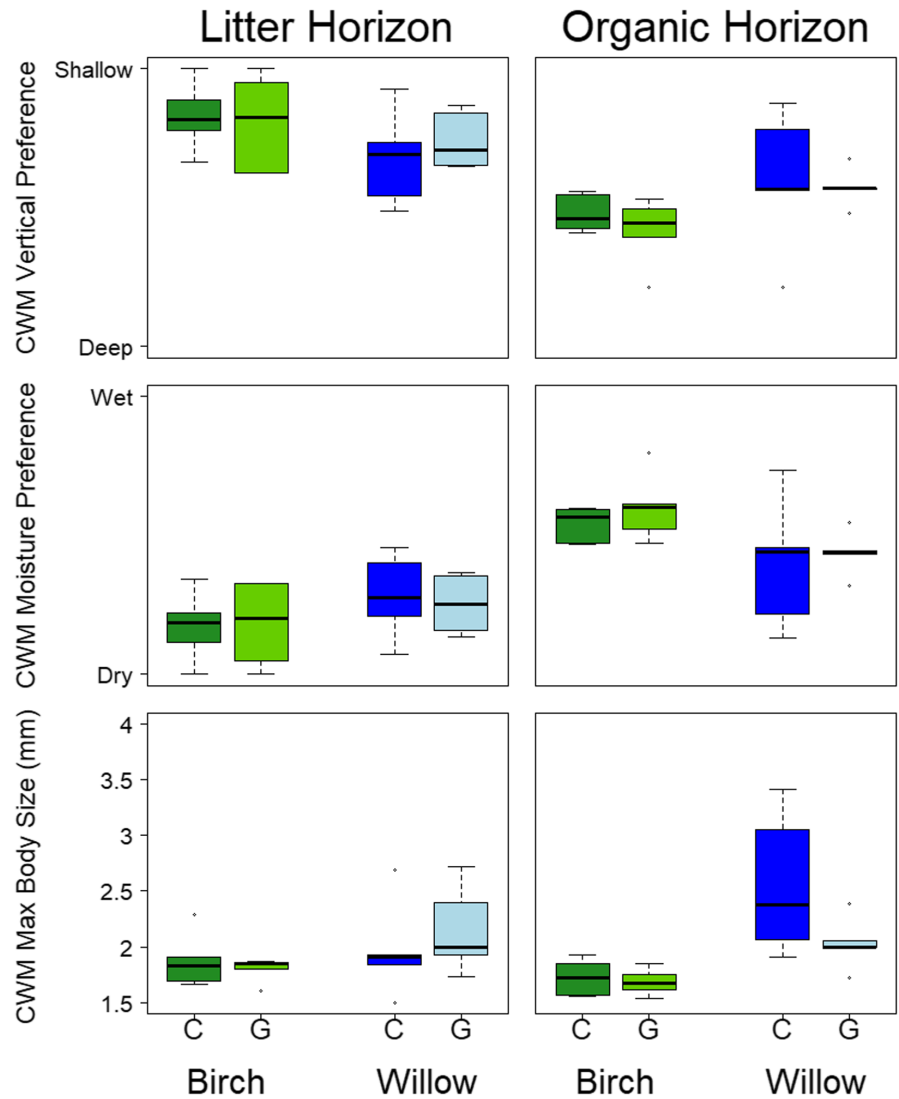
FIGURE 5 Soil mite and collembola abundances in the litter horizon and organic soil horizons of control and girdled plots of birch and willow-dominated plant communities at a subarctic treeline. Bars represent six samples per treatment in birch plots and five samples in willow plots. Error bars represent ± 1 standard error of the mean of total mites or collembola.

contrasts with results from other experiments, which observed significant community shifts over similar time frames, particularly for fungi. Here we discuss the potential mechanisms and ecological drivers for our observed results in fungal and faunal communities.

With regard to soil fungal community composition, our result contrasts with trenching and clear-cutting experiments, but these differences may provide explanations as to why we observed no change in fungal community composition. In long-lived plants, a proportion of recently fixed C is allocated to storage as non-structural carbohydrates that can be retained for a number of years in parenchyma of various organs, including in the hardwood and coarse roots of trees (Hartmann & Trumbore, 2016). In fact, a

large proportion of contemporary root activity in a boreal forest was found to be driven by metabolism of year-old non-structural carbohydrates (Schoor & Trumbore, 2006). From a methodological perspective, we conducted a girdling experiment with phloem disruption located at the bottom of the stem, potentially leaving a number of metres of roots (Friggens et al., 2019) between the point of girdling and the root tips where mycorrhizal symbiosis is formed and activity may be highest. In trenching experiments, where root-shoot connection is severed less than 50 cm from root tips (typically in a 1 × 1 m plot [Sterkenburg et al., 2018]), the root tips are disconnected, not only from aboveground photosynthesis, but also from a significant amount non-structural carbohydrates

FIGURE 6 Community-weighted mean (CWM) values of three traits (soil vertical preference, moisture preference and maximum body size) for abundant collembola species in the litter horizon and organic soil horizon of control (C) and girdled (G) birch and willow plots at the subarctic treeline. Boxplots show the median, interquartile range and extreme values of each treatment.



stored elsewhere in the plant. A lack of non-structural carbohydrate supply might explain why the fungal community response is rapid in trenching experiments (within a year [Lindahl et al., 2010; Sterkenburg et al., 2018]). It is possible that ECMs were kept alive in the birch plots by roots from outside the plot (less likely in willow plots which where the perimeter was trenched). However, this is unlikely given that lateral spread of boreal forest trees was found to be at maximum 5 m (Göttlicher et al., 2008) and the minimum distance between the sampling points and the perimeter of our plots was 8.5 m. We speculate that, even though the large flux of photosynthesis product from the shoot was eliminated by the girdling treatment, and hyphal production declined (Parker et al., 2020), root tips were able to survive and maintain their fungal partners through the mobilisation and allocation of a small supply of C from non-structural carbohydrates.

Clearcutting of temperate spruce forest (Kohout et al., 2018) and subarctic pine forest (Sterkenburg et al., 2019) leads to a substantial shift away from ECM fungi within 12 months. Clearcutting differs from girdling as the total removal of the xylem connection to the canopy results in a complete loss of the tree water and nutrient

sink, and light conditions are also altered. In the present study, birch trees and willow shrubs maintained leaves for up to two seasons after girdling (Parker et al., 2020). This could mean that the continued demand for nutrients may have maintained a reliance on, and continued C delivery to, mycorrhizal symbionts, for longer than if connections to the canopy had been completely severed. Of course, the present experimental system contrasts from previous similar experiments in many other ways, not least in the environmental stress that forest and tundra are adapted to, with extreme cold and disturbance being a regular feature of the landscape (Bjerke et al., 2014).

Abundance and community composition of soil fauna were also resistant to stem girdling. Mites and collembola were extracted 2 years after girdling treatment in birch and willow plots but their community structure remained relatively unchanged, suggesting that they are not sensitive to a change in C resource availability. Hyphal production decreased in the birch plots in this experiment (Parker et al., 2020), but there was no proportional decline in the fauna (although there were some reductions in mite abundance). Fungal hyphae form a significant part of mite and Collembola diets (Anslan et al., 2018; Renker et al., 2005), but in the treeline systems

studied here, root-associated ascomycetes and saprotrophs were the dominant functional guilds and represent a viable food source (Anslan et al., 2018). In boreal forests, girdling negatively affected specialised fungivores, such as fungivorous nematodes (Kudrin et al., 2021) or proturans (Malmström & Persson, 2011), and specific species like the Oribatid mite *Oppiella nova* (Remén et al., 2008), but had no effects on Collembola communities (Malmström & Persson, 2011) or other Oribatid species (Remén et al., 2008). Mites and Collembola are flexible heterotrophs that are able to feed on a variety of food resources, including fungi, algae and detritus (Anslan et al., 2018; Chahartaghi et al., 2005; Ferlian et al., 2015; Krab, Berg, et al., 2013; Schneider et al., 2004). Although it has been shown that Collembola can feed on ECM fungi (Pollierer et al., 2007; Schneider et al., 2005), they could probably easily shift to a heavier reliance on saprotrophic fungi if ECM hyphae become rare (Pollierer & Scheu, 2021). Furthermore, girdling could induce an important input of dead roots and fungal hyphae that could compensate for the decrease in fresh ECM-derived carbon. It seems important to further investigate the complex ecological controls over soil fauna communities in the Arctic, as climate and vegetation cover are changing around them.

The fungal community, as assessed by our DNA-based approach, was largely resistant to the dramatic disturbance above-ground. An immediate explanation for this observation could be that DNA in dead fungal mycelium was still present in the soil, indeed, across biomes, it is known that extracellular 'relic DNA' makes up, on average, 40% of extracted and amplified DNA in soils (Carini et al., 2016). Our results are likely to be influenced by a larger fraction of relic DNA in girdled plots. However, several experiments, in which the C delivery to the rhizosphere was disrupted, found very different results from our own. Using DNA amplification of Swedish boreal forest soils, Lindahl et al. (2010) found that ECM species decreased in abundance 2 weeks after root trenching, and Sterkenburg et al. (2018) observed a large reduction in amplifiable DNA from ECM fungi after a year. Low temperatures at our subarctic site may have preserved DNA from dead fungi (Strickler et al., 2015) for a longer time compared to these warmer boreal sites; however, clear-cutting at a cold subarctic *Pinus sylvestris* forest (67°N) resulted in a 70% reduction in ECM fungal abundance in the O horizon (Sterkenburg et al., 2019). Furthermore, canopy defoliation by caterpillars in subarctic birch forests resulted in large changes in fungal community structure as measured using DNA techniques (Saravesi et al., 2015). Therefore, despite an important, yet unquantified contribution of relic DNA to assessed fungal communities, we do not believe that the apparent resistance of the fungal community to girdling was only an artefact of the method.

An ecological factor that may be related to the observed resistance of treeline soil communities to girdling is that both mountain birches and willow shrubs are well adapted to regular, intense canopy disturbance. Mountain birch forests across Northern Fennoscandia endure cyclical outbreaks of geometrid moth caterpillars, which cause widespread and often complete, defoliation of the canopy (Jepsen et al., 2008). However, mountain birch trees often survive these outbreaks by resprouting stems from their base (Karlsson

& Weih, 2003; Tenow et al., 2005), again pointing to a significant storage of non-structural carbohydrates. Arctic willows are equally subject to intense disturbance by herbivores (Dahl et al., 2017; Tape et al., 2016; te Beest et al., 2016) and change their morphology in response to herbivory, for example, by resprouting shoots (Christie et al., 2014). We speculate that there is a below-ground facet to birch and willow survival after disturbance: the ability to maintain the mycorrhizosphere until above-ground productivity can rebound to normal levels, which can take a number of years (Karlsson & Weih, 2003; Vindstad et al., 2019). As much as some areas of the tundra are increasing in productivity and undergoing shrub expansion, others are experiencing more 'browning events' in the form of disturbance (Myers-Smith et al., 2020; Phoenix & Bjerke, 2016). Therefore, it is important to know how short- and long-term cessation in C-fixation influence root-driven processes (Parker, Thurston, et al., 2021). Our data suggest that maintenance of a soil fungal community in the face of disturbance may aid willow survival through intermittent 'browning events' as they continue on their overall 'greening' trend (Myers-Smith et al., 2020).

4.3 | ECM community dynamics in response to changes to photosynthate delivery

We hypothesised (H3) that girdling would result in a large compositional change in the ECM fungal community within the treeline birch forest (more so than in the willow plots) with a reduction in supposed C-demanding genera. Although we did observe some systematic shifts within the ECM community after girdling, changes were as large in willow plots as in birch plots. In support of our hypothesis, species of 'medium distance-fringe' genera, like *Piloderma* and *Cortinarius*, declined in abundance across both plant communities in response to girdling, probably because the reduction in photosynthate delivery reduced the competitiveness of these supposedly C-demanding species (Saikkonen et al., 1999) that turn over biomass quickly (Clemmensen et al., 2021) and/or express costly extracellular enzymes at high levels (Bödeker et al., 2014). Saikkonen et al. (1999) argued that, as plant C delivery to ECM root tips is reduced by disturbance in subarctic ecosystems, less 'C-demanding' species hold an advantage. This hypothesis has empirical support from studies on defoliation (Parker et al., 2017; Saravesi et al., 2015) and now also in the context of this girdling experiment. It is now important to understand whether the large-scale changes in productivity occurring in the Arctic and boreal biomes will influence ECM community composition and function. More specifically, will major disruptions in C delivery to ECM fungi alter the rate of mycorrhizal-driven turnover of organic matter (Clemmensen et al., 2021)?

4.4 | Soil community composition in important tundra vegetation types

Characterising the soil community in willow soils is important, as willows play a key role in Arctic 'greening' and 'shrubification'

(Myers-Smith et al., 2011; Tape et al., 2006). We hypothesised that ECM fungi would be less prolific under willow hosts as a result of high soil moisture and relatively high nutrient availability (Chen et al., 2020; Nadelhoffer et al., 1991), as suggested by the observation that overall hyphal production was less in willow plots (Parker et al., 2020). Soil in willow plots was wetter and had lower C:N ratio but the concentration of ECM fungal gene copies in the organic horizon was greater than in the birch plots. Conversely, there were significantly fewer mites and Collembola in willow plots (as we hypothesised), which may be linked to lower overall fungal biomass (Figure 1) and soil respiration (Parker et al., 2020). Willows in the tundra tend to grow in depressions in the landscape where snow accumulates and water saturation is a regular feature over the annual cycle (Parker et al., 2020; Tape et al., 2006). The regularly high water table may restrict proliferation of soil fauna as the 'habitable' volume of soil is small. This hypothesis is supported by our observation that the Collembola community in willow plots tended to include more species with a preference for surface habitats. The ECM fungal community in willow plots differed from that of birch plots, with more short distance and medium distance-smooth morphotypes. The higher abundance of *Tomentella* and *Cenococcum*, which tend to respond positively to N deposition (van der Linde et al., 2018), accords with a more nutrient-rich environment, whereas *Piloderma*, which typically responds negatively to N deposition (van der Linde et al., 2018), was less abundant. The molecular data presented here show that there is an important ECM community in willow plots but this assemblage may produce either fewer hyphae, or hyphae that are less likely to grow into sandbags (the standard method for measuring hyphal production; Wallander et al., 2013).

In birch plots, ECM fungal species had a relative abundance of only 9.5% of the total fungal community, but considering that fast mycelial growth rates have been consistently observed in mountain birch forests (Clemmensen et al., 2021; Friggens et al., 2019; Parker et al., 2015, 2020), it is likely that this small constituent of the fungal community has a disproportionately large effect on turnover of C. Indeed, Clemmensen et al. (2021) linked ECM fungi in treeline birch forests to a high potential for SOM degradation and below-ground C turnover. As with our study, Clemmensen et al. (2021) showed that root-associated ascomycetes are the dominant fungal guild in mountain birch forest; their DNA is present in abundance but they may have slower metabolism and turnover rate than ECM fungi (Clemmensen et al., 2015). A high proportion of a collembolan fungal diet comprises ascomycetes (Anslan et al., 2018), and the large stock of ascomycete biomass could be a primary driver of high soil fauna abundance which, in turn, controls ascomycete biomass turnover. As Clemmensen et al. (2021) suggested, the mycelium of mycorrhizal fungi associated with ericaceous plants may contribute to SOM formation, while the less abundant, but more active, ECM fungi may act to reduce SOM stocks. The high numbers of soil fauna in birch forests could therefore be driving the turnover of mycelial biomass that would otherwise be difficult to access (Clemmensen et al., 2015). Due to the large-scale vegetation change occurring at Arctic treelines and in the tundra, further characterisation of soil and fungal communities in relation to plant communities from around the Arctic should be a

research priority, if we are to understand the C cycle feedbacks associated with this change (Parker, Thurston, et al., 2021).

5 | CONCLUSIONS

We show that fungal and microarthropod communities of mountain birch forest and willow shrub are distinct, but once established, they are resistant to complete cessation of below-ground C supply from the canopy phloem. This finding has implications for how Arctic systems may respond to disturbance events that have similar impacts on below ground C supply, such as canopy disturbance events. Our results contrast similar experiments from more southerly biomes, raising questions about the mechanisms driving the ecology and adaptations of Arctic ecosystems to stress and disturbance. Given the progressive greening trend around the Arctic and the growing importance of intermittent browning events, unpicking the belowground-mediated feedbacks will be key to understanding Arctic biome-wide responses to climate change.

AUTHOR CONTRIBUTIONS

Thomas C. Parker, Ilona Kater, Philip A. Wookey, Iain P. Hartley, David Johnson, Björn D. Lindahl, Karina E. Clemmensen, Nina L. Friggens, Lorna E. Street and Jens-Arne Subke designed, implemented and collected the plot-level data from the experiment; Mathilde Chomel processed and identified fauna extractions; Thomas C. Parker analysed the data and wrote the manuscript. All authors significantly contributed to multiple drafts of the paper.

ACKNOWLEDGEMENTS

This work was funded by the Natural Environment Research Council (NERC) grant nos. NE/P002722/1 and NE/P002722/2 to PAW, DJ, JA-S and IPH. We warmly thank Gwen Lancashire for assistance in collecting field data. We thank staff of the Abisko Naturvetenskapliga Station for their assistance and logistical support.

CONFLICT OF INTEREST

We declare no conflicts of interest.

PEER REVIEW

The peer review history for this article is available at <https://publons.com/publon/10.1111/1365-2745.13994>.

DATA AVAILABILITY STATEMENT

All sequence data are stored at NCBI-SRA (accession PRJNA846260): <https://www.ncbi.nlm.nih.gov/bioproject/PRJNA846260/>. ITS copy numbers and species hypothesis relative abundance data are available at: <https://doi.org/10.5285/d6c787ec-146c-461b-b8a2-e0251259036c> (Parker et al., 2022a). Flux, hyphae and root data are available at <https://doi.org/10.5285/4418c631-c39c-467c-b3b8-c75142fcae0a> (Parker, Clemmensen, et al., 2021). Soil fauna data are available at <https://doi.org/10.5285/d3c98f24-7a4d-40b8-989a-6cc340e91cac> (Parker et al., 2022b).

ORCID

- Thomas C. Parker  <https://orcid.org/0000-0002-3648-5316>
 Mathilde Chomel  <https://orcid.org/0000-0001-5110-2355>
 Karina E. Clemmensen  <https://orcid.org/0000-0002-9627-6428>
 Nina L. Friggens  <https://orcid.org/0000-0002-5536-6312>
 Iain P. Hartley  <https://orcid.org/0000-0002-9183-6617>
 David Johnson  <https://orcid.org/0000-0003-2299-2525>
 Ilona Kater  <https://orcid.org/0000-0002-2910-1337>
 Eveline J. Krab  <https://orcid.org/0000-0001-8262-0198>
 Björn D. Lindahl  <https://orcid.org/0000-0002-3384-4547>
 Lorna E. Street  <https://orcid.org/0000-0001-9570-7479>
 Jens-Arne Subke  <https://orcid.org/0000-0001-9244-639X>
 Philip A. Wookey  <https://orcid.org/0000-0001-5957-6424>

REFERENCES

- Agerer, R. (2001). Exploration types of ectomycorrhizae—A proposal to classify ectomycorrhizal mycelial systems according to their patterns of differentiation and putative ecological importance. *Mycorrhiza*, *11*, 107–114.
- Anslan, S., Bahram, M., & Tedersoo, L. (2018). Seasonal and annual variation in fungal communities associated with epigeic springtails (*Collembola* spp.) in boreal forests. *Soil Biology and Biochemistry*, *116*, 245–252.
- Bjerke, J. W., Karlsen, S. R., Hogda, K. A., Malnes, E., Jepsen, J. U., Lovibond, S., Vikhamar-Schuler, D., & Tommervik, H. (2014). Record-low primary productivity and high plant damage in the Nordic Arctic region in 2012 caused by multiple weather events and pest outbreaks. *Environmental Research Letters*, *9*, 084006.
- Bödeker, I. T. M., Clemmensen, K. E., de Boer, W., Martin, F., Olson, Å., & Lindahl, B. D. (2014). Ectomycorrhizal Cortinari species participate in enzymatic oxidation of humus in northern forest ecosystems. *New Phytologist*, *203*, 245–256.
- Bret-Harte, M. S., Mack, M. C., Shaver, G. R., Huebner, D. C., Johnston, M., Mojica, C. A., Pizano, C., & Reiskind, J. A. (2013). The response of Arctic vegetation and soils following an unusually severe tundra fire. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, *368*, 20120490.
- Carini, P., Marsden, P. J., Leff, J. W., Morgan, E. E., Strickland, M. S., & Fierer, N. (2016). Relic DNA is abundant in soil and obscures estimates of soil microbial diversity. *Nature Microbiology*, *2*, 16242.
- Castaño, C., Berlin, A., Brandström Durling, M., Ihrmark, K., Lindahl, B. D., Stenlid, J., Clemmensen, K. E., & Olson, Å. (2020). Optimized metabarcoding with Pacific biosciences enables semi-quantitative analysis of fungal communities. *New Phytologist*, *228*, 1149–1158.
- Chahartaghi, M., Langel, R., Scheu, S., & Ruess, L. (2005). Feeding guilds in collembola based on nitrogen stable isotope ratios. *Soil Biology and Biochemistry*, *37*, 1718–1725.
- Chen, W., Tape, K. D., Euskirchen, E. S., Liang, S., Matos, A., Greenberg, J., & Fraterrigo, J. M. (2020). Impacts of Arctic shrubs on root traits and belowground nutrient cycles across a northern Alaskan climate gradient. *Frontiers in Plant Science*, *11*, 1943.
- Chomel, M., Lavallee, J. M., Alvarez-Segura, N., de Castro, F., Rhymes, J. M., Caruso, T., de Vries, F. T., Baggs, E. M., Emmerson, M. C., Bardgett, R. D., & Johnson, D. (2019). Drought decreases incorporation of recent plant photosynthate into soil food webs regardless of their trophic complexity. *Global Change Biology*, *25*, 3549–3561.
- Christie, K. S., Ruess, R. W., Lindberg, M. S., & Mulder, C. P. (2014). Herbivores influence the growth, reproduction, and morphology of a widespread Arctic willow. *PLoS ONE*, *9*, e101716.
- Clemmensen, K. E., Durling, M. B., Michelsen, A., Hallin, S., Finlay, R. D., & Lindahl, B. D. (2021). A tipping point in carbon storage when forest expands into tundra is related to mycorrhizal recycling of nitrogen. *Ecology Letters*, *24*, 1193–1204.
- Clemmensen, K. E., Finlay, R. D., Dahlberg, A., Stenlid, J., Wardle, D. A., & Lindahl, B. D. (2015). Carbon sequestration is related to mycorrhizal fungal community shifts during long-term succession in boreal forests. *New Phytologist*, *205*, 1525–1526.
- Crawley, M. J. (2007). *The R book*. Wiley.
- Dahl, M. B., Priemé, A., Brejnrod, A., Brusvang, P., Lund, M., Nymand, J., Kramshøj, M., Ro-Poulsen, H., & Haugwitz, M. S. (2017). Warming, shading and a moth outbreak reduce tundra carbon sink strength dramatically by changing plant cover and soil microbial activity. *Scientific Reports*, *7*, 16035.
- Edgar, R. C. (2013). UPARSE: Highly accurate OTU sequences from microbial amplicon reads. *Nature Methods*, *10*, 996–998.
- Eissfeller, V., Beyer, F., Valtanen, K., Hertel, D., Maraun, M., Polle, A., & Scheu, S. (2013). Incorporation of plant carbon and microbial nitrogen into the rhizosphere food web of beech and ash. *Soil Biology and Biochemistry*, *62*, 76–81.
- Elmendorf, S. C., Henry, G. H. R., Hollister, R. D., Bjork, R. G., Bjorkman, A. D., Callaghan, T. V., Collier, L. S., Cooper, E. J., Cornelissen, J. H. C., Day, T. A., Fosaa, A. M., Gould, W. A., Grétarsdóttir, J., Harte, J., Hermanutz, L., Hik, D. S., Hofgaard, A., Jarrad, F., Jónsdóttir, I. S., ... Wookey, P. A. (2012). Global assessment of experimental climate warming on tundra vegetation: Heterogeneity over space and time. *Ecology Letters*, *15*, 164–175.
- Elmendorf, S. C., Henry, G. H. R., Hollister, R. D., Bjork, R. G., Boulanger-Lapointe, N., Cooper, E. J., Cornelissen, J. H. C., Day, T. A., Dorrepaal, E., Elumeeva, T. G., Gill, M., Gould, W. A., Harte, J., Hik, D. S., Hofgaard, A., Johnson, D. R., Johnstone, J. F., Jónsdóttir, I. S., Jorgenson, J. C., ... Wipf, S. (2012). Plot-scale evidence of tundra vegetation change and links to recent summer warming. *Nature Climate Change*, *2*, 453–457.
- Ferlian, O., Klarner, B., Langeneckert, A. E., & Scheu, S. (2015). Trophic niche differentiation and utilisation of food resources in collembolans based on complementary analyses of fatty acids and stable isotopes. *Soil Biology and Biochemistry*, *82*, 28–35.
- Fjellberg, A. (2007). *The collembola of Fennoscandia and Denmark, part II: Entomobryomorpha and Symphypleona*. Brill.
- Frey, S. D. (2019). Mycorrhizal fungi as mediators of soil organic matter dynamics. *Annual Review of Ecology, Evolution, and Systematics*, *50*, 237–259.
- Friggens, N. L., Aspray, T. J., Parker, T. C., Subke, J.-A., & Wookey, P. A. (2019). Spatial patterns in soil organic matter dynamics are shaped by mycorrhizosphere interactions in a treeline forest. *Plant and Soil*, *447*, 521–535.
- Gilbert, K. J., Fahey, T. J., Maerz, J. C., Sherman, R. E., Bohlen, P., Dombroskie, J. J., Groffman, P. M., & Yavitt, J. B. (2014). Exploring carbon flow through the root channel in a temperate forest soil food web. *Soil Biology and Biochemistry*, *76*, 45–52.
- Goncharov, A. A., Tsurikov, S. M., Potapov, A. M., & Tiunov, A. V. (2016). Short-term incorporation of freshly fixed plant carbon into the soil animal food web: Field study in a spruce forest. *Ecological Research*, *31*, 923–933.
- Göttlicher, S. G., Taylor, A. F. S., Grip, H., Betson, N. R., Valinger, E., Högberg, M. N., & Högberg, P. (2008). The lateral spread of tree root systems in boreal forests: Estimates based on ¹⁵N uptake and distribution of sporocarps of ectomycorrhizal fungi. *Forest Ecology and Management*, *255*, 75–81.
- Handa, I. T., Aerts, R., Berendse, F., Berg, M. P., Bruder, A., Butenschoen, O., Chauvet, E., Gessner, M. O., Jabiol, J., Makkonen, M., McKie, B. G., Malmqvist, B., Peeters, E. T. H. M., Scheu, S., Schmid, B., van Ruijven, J., Vos, V. C. A., & Hättenschwiler, S. (2014). Consequences of biodiversity loss for litter decomposition across biomes. *Nature*, *509*, 218–221.
- Hartmann, H., & Trumbore, S. (2016). Understanding the roles of non-structural carbohydrates in forest trees—From what we can measure to what we want to know. *New Phytologist*, *211*, 386–403.

- Heinemeyer, A., Hartley, I. P., Evans, S. P., De la Fuente, J. A. C., & Ineson, P. (2007). Forest soil CO₂ flux: Uncovering the contribution and environmental responses of ectomycorrhizas. *Global Change Biology*, 13, 1786–1797.
- Hibbett, D. S., Gilbert, L. B., & Donoghue, M. J. (2000). Evolutionary instability of ectomycorrhizal symbioses in basidiomycetes. *Nature*, 407, 506–508.
- Hobbie, J. E., & Hobbie, E. A. (2006). ¹⁵N in symbiotic fungi and plants estimates nitrogen and carbon flux rates in Arctic tundra. *Ecology*, 87, 816–822.
- Hobbie, J. E., Hobbie, E. A., Drossman, H., Conte, M., Weber, J. C., Shamhart, J., & Weinrobe, M. (2009). Mycorrhizal fungi supply nitrogen to host plants in Arctic tundra and boreal forests: ¹⁵N is the key signal. This article is one of a selection of papers in the special issue on polar and Arctic microbiology. *Canadian Journal of Microbiology*, 55, 84–94.
- Högberg, P., Högberg, M. N., Gottlicher, S. G., Betson, N. R., Keel, S. G., Metcalfe, D. B., Campbell, C., Schindlbacher, A., Hurry, V., Lundmark, T., Linder, S., & Näsholm, T. (2008). High temporal resolution tracing of photosynthate carbon from the tree canopy to forest soil microorganisms. *New Phytologist*, 177, 220–228.
- Högberg, P., Nordgren, A., Buchmann, N., Taylor, A. F. S., Ekblad, A., Högberg, M. N., Nyberg, G., Ottosson-Lofvenius, M., & Read, D. J. (2001). Large-scale forest girdling shows that current photosynthesis drives soil respiration. *Nature*, 411, 789–792.
- Hopkin, S. P. (2007). *A key to the collembola (springtails) of Britain and Ireland*. FSC Publications.
- Hunt, H. W., Coleman, D. C., Ingham, E. R., Ingham, R. E., Elliott, E. T., Moore, J. C., Rose, S. L., Reid, C. P. P., & Morley, C. R. (1987). The detrital food web in a shortgrass prairie. *Biology and Fertility of Soils*, 3, 57–68.
- Ihrmark, K., Bödeker, I. T. M., Cruz-Martinez, K., Friberg, H., Kubartova, A., Schenck, J., Strid, Y., Stenlid, J., Brandström-Durling, M., Clemmensen, K. E., & Lindahl, B. D. (2012). New primers to amplify the fungal ITS2 region—Evaluation by 454-sequencing of artificial and natural communities. *FEMS Microbiology Ecology*, 82, 666–677.
- Jepsen, J. U., Hagen, S. B., Ims, R. A., & Yoccoz, N. G. (2008). Climate change and outbreaks of the geometrids *Operophtera brumata* and *Epirrita autumnata* in subarctic birch forest: Evidence of a recent outbreak range expansion. *Journal of Animal Ecology*, 77, 257–264.
- Kanters, C., Anderson, I. C., & Johnson, D. (2015). Chewing up the wood-wide web: Selective grazing on ectomycorrhizal fungi by collembola. *Forests*, 6, 2560–2570.
- Karlsson, P. S., & Weih, M. (2003). Long-term patterns of leaf, shoot and wood production after insect herbivory in the mountain birch. *Functional Ecology*, 17, 841–850.
- Kohout, P. (2017). Biogeography of ericoid mycorrhiza. In L. Tedersoo (Ed.), *Biogeography of mycorrhizal symbiosis* (pp. 179–193). Springer.
- Kohout, P., Charvátová, M., Štursová, M., Mašíňová, T., Tomšovský, M., & Baldrian, P. (2018). Clearcutting alters decomposition processes and initiates complex restructuring of fungal communities in soil and tree roots. *The ISME Journal*, 12, 692–703.
- Krab, E. J., Berg, M. P., Aerts, R., van Logtestijn, R. S. P., & Cornelissen, J. H. C. (2013). Vascular plant litter input in subarctic peat bogs changes collembola diets and decomposition patterns. *Soil Biology and Biochemistry*, 63, 106–115.
- Krab, E. J., Van Schroyen Lantman, I. M., Cornelissen, J. H. C., & Berg, M. P. (2013). How extreme is an extreme climatic event to a subarctic peatland springtail community? *Soil Biology and Biochemistry*, 59, 16–24.
- Kudrin, A. A., Zuev, A. G., Taskaeva, A. A., Konakova, T. N., Kolesnikova, A. A., Gruzdev, I. V., Gabov, D. N., Yakovleva, E. V., & Tiunov, A. V. (2021). Spruce girdling decreases abundance of fungivorous soil nematodes in a boreal forest. *Soil Biology and Biochemistry*, 155, 108184.
- Kuznetsova, N. A. (2003). Humidity and distribution of springtails. *Entomological Review*, 83, 230–238.
- Kyaschenko, J., Clemmensen, K. E., Karlton, E., & Lindahl, B. D. (2017). Below-ground organic matter accumulation along a boreal forest fertility gradient relates to guild interaction within fungal communities. *Ecology Letters*, 20, 1546–1555.
- Lindahl, B. D., de Boer, W., & Finlay, R. D. (2010). Disruption of root carbon transport into forest humus stimulates fungal opportunists at the expense of mycorrhizal fungi. *ISME Journal*, 4, 872–881.
- Lindahl, B. D., Kyaschenko, J., Varenius, K., Clemmensen, K. E., Dahlberg, A., Karlton, E., & Stendahl, J. (2021). A group of ectomycorrhizal fungi restricts organic matter accumulation in boreal forest. *Ecology Letters*, 24, 1341–1351.
- Lindahl, B. D., Nilsson, R. H., Tedersoo, L., Abarenkov, K., Carlsen, T., Kjoller, R., Koljalg, U., Pennanen, T., Rosendahl, S., Stenlid, J., & Kausarud, H. (2013). Fungal community analysis by high-throughput sequencing of amplified markers—A user's guide. *New Phytologist*, 199, 288–299.
- Malmström, A., & Persson, T. (2011). Responses of collembola and *Protura* to tree girdling—some support for ectomycorrhizal feeding. *Soil Org*, 83, 279–285.
- Mudryk, L., Brown, R., Luojus, K., Decharme, B., & Helfrich, S. (2019). *Arctic Report Card 2019*.
- Myers-Smith, I. H., Forbes, B. C., Wilmking, M., Hallinger, M., Lantz, T., Blok, D., Tape, K. D., Macias-Fauria, M., Sass-Klaassen, U., Levesque, E., Boudreau, S., Ropars, P., Hermanutz, L., Trant, A., Collier, L. S., Weijers, S., Rozema, J., Rayback, S. A., Schmidt, N. M., ... Hik, D. S. (2011). Shrub expansion in tundra ecosystems: Dynamics, impacts and research priorities. *Environmental Research Letters*, 6, 045509.
- Myers-Smith, I. H., Kerby, J. T., Phoenix, G. K., Bjerke, J. W., Epstein, H. E., Assmann, J. J., John, C., Andreu-Hayles, L., Angers-Blondin, S., Beck, P. S. A., Berner, L. T., Bhatt, U. S., Bjorkman, A. D., Blok, D., Bryn, A., Christiansen, C. T., Cornelissen, J. H. C., Cunliffe, A. M., Elmendorf, S. C., ... Wipf, S. (2020). Complexity revealed in the greening of the Arctic. *Nature Climate Change*, 10, 106–117.
- Nadelhoffer, K. J., Giblin, A. E., Shaver, G. R., & Laundre, J. A. (1991). Effects of temperature and substrate quality on element mineralization in 6 arctic soils. *Ecology*, 72, 242–253.
- Oksanen, J. (2013). *Community ecology package*.
- Parker, T. C., Chomel, M., Clemmensen, K. E., Friggens, N. L., Hartley, I. P., Johnson, D., Krab, E. J., Lindahl, B. D., Street, L. E., Subke, J.-A., & Wookey, P. A. (2022a). *Soil fungi gene copy numbers and community composition under birch and willow vegetation in girdled and non-girdled plots, subarctic Sweden, 2017–2019* [Data set]. NERC EDS Environmental Information Data Centre. <https://doi.org/10.5285/D6C787EC-146C-461B-B8A2-E0251259036C>
- Parker, T. C., Chomel, M., Clemmensen, K. E., Friggens, N. L., Hartley, I. P., Johnson, D., Krab, E. J., Lindahl, B. D., Street, L. E., Subke, J. A., & Wookey, P. A. (2022b). *Soil fauna abundance under birch and willow vegetation in girdled and non-girdled plots, subarctic Sweden, 2019* [Data set]. NERC EDS Environmental Information Data Centre. <https://doi.org/10.5285/D3C98F24-7A4D-40B8-989A-6CC340E91CAC>
- Parker, T. C., Clemmensen, K. E., Friggens, N. L., Hartley, I. P., Johnson, D., Lindahl, B. D., Olofsson, J., Siewert, M. B., Street, L. E., Subke, J.-A., & Wookey, P. A. (2020). Rhizosphere allocation by canopy-forming species dominates soil CO₂ efflux in a subarctic landscape. *New Phytologist*, 227, 1818–1830.
- Parker, T. C., Clemmensen, K. E., Friggens, N. L., Hartley, I. P., Johnson, D., Lindahl, B. D., Olofsson, J., Siewert, M. B., Street, L. E., Subke, J.-A., & Wookey, P. A. (2021). *CO₂ efflux, root and mycorrhizal hyphae production in subarctic willow shrub and treeline birch forest, Abisko 2017–2018* [Data set]. NERC Environmental Information Data Centre. <https://doi.org/10.5285/4418C631-C39C-467C-B3B8-C75142FCAEOA>

- Parker, T. C., Sadowsky, J., Dunleavy, H., Subke, J.-A., Frey, S. D., & Wookey, P. A. (2017). Slowed biogeochemical cycling in sub-arctic birch Forest linked to reduced mycorrhizal growth and community change after a defoliation event. *Ecosystems*, *20*, 316–330.
- Parker, T. C., Subke, J.-A., & Wookey, P. A. (2015). Rapid carbon turnover beneath shrub and tree vegetation is associated with low soil carbon stocks at a subarctic treeline. *Global Change Biology*, *21*, 2070–2081.
- Parker, T. C., Thurston, A. M., Raundrup, K., Subke, J.-A., Wookey, P. A., & Hartley, I. P. (2021). Shrub expansion in the Arctic may induce large-scale carbon losses due to changes in plant-soil interactions. *Plant and Soil*, *463*, 643–651.
- Pérez-Izquierdo, L., Clemmensen, K. E., Strengbom, J., Granath, G., Wardle, D. A., Nilsson, M.-C., & Lindahl, B. D. (2021). Crown-fire severity is more important than ground-fire severity in determining soil fungal community development in the boreal forest. *Journal of Ecology*, *109*, 504–518.
- Phoenix, G. K., & Bjerke, J. W. (2016). Arctic browning: Extreme events and trends reversing arctic greening. *Global Change Biology*, *22*, 2960–2962.
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., & R Core Team. (2017). *nlme: Linear and nonlinear mixed effects models*.
- Pollierer, M. M., Langel, R., Körner, C., Maraun, M., & Scheu, S. (2007). The underestimated importance of belowground carbon input for forest soil animal food webs. *Ecology Letters*, *10*, 729–736.
- Pollierer, M. M., & Scheu, S. (2021). Stable isotopes of amino acids indicate that soil decomposer microarthropods predominantly feed on saprotrophic fungi. *Ecosphere*, *12*, e03425.
- Pölme, S., Abarenkov, K., Henrik Nilsson, R., Lindahl, B. D., Clemmensen, K. E., Kauterud, H., Nguyen, N., Kjäller, R., Bates, S. T., Baldrian, P., Frøsløv, T. G., Adojaan, K., Vizzini, A., Suija, A., Pfister, D., Baral, H. O., Järv, H., Madrid, H., Nordén, J., ... Tedersoo, L. (2020). FungalTraits: A user-friendly traits database of fungi and fungus-like stramenopiles. *Fungal Diversity*, *105*, 1–16.
- Post, E., Forchhammer, M. C., Bret-Harte, M. S., Callaghan, T. V., Christensen, T. R., Elberling, B., Fox, A. D., Gilg, O., Hik, D. S., Høye, T. T., Ims, R. A., Jeppesen, E., Klein, D. R., Madsen, J., McGuire, A. D., Rysgaard, S., Schindler, D. E., Stirling, I., Tamstorf, M. P., ... Aastrup, P. (2009). Ecological dynamics across the Arctic associated with recent climate change. *Science*, *325*, 1355–1358.
- Read, D. J., Leake, J. R., & Perez-Moreno, J. (2004). Mycorrhizal fungi as drivers of ecosystem processes in heathland and boreal forest biomes. *Canadian Journal of Botany-Revue Canadienne De Botanique*, *83*, 1243–1263.
- Rees, G., Hofgaard, A., Boudreau, S., Cairns, D., Harper, K., Mamet, S., Mathisen, I., Swirad, Z., & Tutubalina, O. (2020). Is subarctic forest advance able to keep pace with climate change? *Global Change Biology*, *26*, 3965–3977.
- Remén, C., Persson, T., Finlay, R., & Ahlström, K. (2008). Responses of oribatid mites to tree girdling and nutrient addition in boreal coniferous forests. *Soil Biology and Biochemistry*, *40*, 2881–2890.
- Renker, C., Otto, P., Schneider, K., Zimdars, B., Maraun, M., & Buscot, F. (2005). Oribatid mites as potential vectors for soil microfungi: Study of mite-associated fungal species. *Microbial Ecology*, *50*, 518–528.
- Ruf, A., Kuz'yakov, Y., & Lopatovskaya, O. (2006). Carbon fluxes in soil food webs of increasing complexity revealed by ¹⁴C labelling and ¹³C natural abundance. *Soil Biology and Biochemistry*, *38*, 2390–2400.
- Saikkonen, K., Ahonen-Jonnarth, U., Markkola, A. M., Helander, M., Tuomi, J., Roitto, M., & Ranta, H. (1999). Defoliation and mycorrhizal symbiosis: A functional balance between carbon sources and below-ground sinks. *Ecology Letters*, *2*, 19–26.
- Saravesi, K., Aikio, S., Wäli, P. R., Ruotsalainen, A. L., Kaukonen, M., Huusko, K., Suokas, M., Brown, S. P., Jumpponen, A., Tuomi, J., & Markkola, A. (2015). Moth outbreaks Alter root-associated fungal communities in Subarctic Mountain birch forests. *Microbial Ecology*, *69*, 788–797.
- Schneider, K., Migge, S., Norton, R. A., Scheu, S., Langel, R., Reineking, A., & Maraun, M. (2004). Trophic niche differentiation in soil microarthropods (Oribatida, Acari): Evidence from stable isotope ratios (¹⁵N/¹⁴N). *Soil Biology and Biochemistry*, *36*, 1769–1774.
- Schneider, K., Renker, C., & Maraun, M. (2005). Oribatid mite (Acari, Oribatida) feeding on ectomycorrhizal fungi. *Mycorrhiza*, *16*, 67–72.
- Schuur, E. A. G., & Trumbore, S. E. (2006). Partitioning sources of soil respiration in boreal black spruce forest using radiocarbon. *Global Change Biology*, *12*, 165–176.
- Setälä, H. (2000). Reciprocal interactions between scots pine and soil food web structure in the presence and absence of ectomycorrhiza. *Oecologia*, *125*, 109–118.
- Setälä, H., Kulmala, P., Mikola, J., & Markkola, A. M. (1999). Influence of ectomycorrhiza on the structure of detrital food webs in pine rhizosphere. *Oikos*, *87*, 113–122.
- Sloan, V. L., Fletcher, B. J., Press, M. C., Williams, M., & Phoenix, G. K. (2013). Leaf and fine root carbon stocks and turnover are coupled across Arctic ecosystems. *Global Change Biology*, *19*, 3668–3676.
- Sterkenburg, E., Clemmensen, K. E., Ekblad, A., Finlay, R. D., & Lindahl, B. D. (2018). Contrasting effects of ectomycorrhizal fungi on early and late stage decomposition in a boreal forest. *The ISME Journal*, *12*, 2187–2197.
- Sterkenburg, E., Clemmensen, K. E., Lindahl, B. D., & Dahlberg, A. (2019). The significance of retention trees for survival of ectomycorrhizal fungi in clear-cut scots pine forests. *Journal of Applied Ecology*, *56*, 1367–1378.
- Strickler, K. M., Fremier, A. K., & Goldberg, C. S. (2015). Quantifying effects of UV-B, temperature, and pH on eDNA degradation in aquatic microcosms. *Biological Conservation*, *183*, 85–92.
- Sturm, M., Schimel, J., Michaelson, G., Welker, J. M., Oberbauer, S. F., Liston, G. E., Fahnestock, J., & Romanovsky, V. E. (2005). Winter biological processes could help convert arctic tundra to shrubland. *Bioscience*, *55*, 17–26.
- Tape, K., Sturm, M., & Racine, C. (2006). The evidence for shrub expansion in northern Alaska and the pan-Arctic. *Global Change Biology*, *12*, 686–702.
- Tape, K. D., Gustine, D. D., Ruess, R. W., Adams, L. G., & Clark, J. A. (2016). Range expansion of moose in Arctic Alaska linked to warming and increased shrub habitat. *PLoS ONE*, *11*, e0152636.
- te Beest, M., Sitters, J., Ménard, C. B., & Olofsson, J. (2016). Reindeer grazing increases summer albedo by reducing shrub abundance in Arctic tundra. *Environmental Research Letters*, *11*, 125013.
- Tedersoo, L., & Smith, M. E. (2013). Lineages of ectomycorrhizal fungi revisited: Foraging strategies and novel lineages revealed by sequences from belowground. *Fungal Biology Reviews*, *27*, 83–99.
- Tenow, O., Bylund, H., Nilssen, A. C., & Karlsson, P. S. (2005). Long-term influence of herbivores on northern birch forests. In M. M. Caldwell (Ed.), *Plant ecology, herbivory, and human impact in Nordic Mountain birch forests* (pp. 165–181). Springer Berlin Heidelberg.
- Trehanne, R., Bjerke, J. W., Tømmervik, H., & Phoenix, G. K. (2020). Extreme event impacts on pCO₂ fluxes across a range of high latitude, shrub-dominated ecosystems. *Environmental Research Letters*, *15*, 104084.
- van der Linde, S., Suz, L. M., Orme, C. D. L., Cox, F., Andreae, H., Asi, E., Atkinson, B., Benham, S., Carroll, C., Cools, N., de Vos, B., Dietrich, H. P., Eichhorn, J., Gehrman, J., Grebenc, T., Gweon, H. S., Hansen, K., Jacob, F., Kristöfel, F., ... Bidartondo, M. I. (2018). Environment and host as large-scale controls of ectomycorrhizal fungi. *Nature*, *558*, 243–248.
- Van Straalen, N. M., & Rijninks, P. C. (1982). Efficiency of Tullgren apparatus with respect to interpreting seasonal changes in age structure of soil arthropod populations. *Pedobiologia*, *24*, 197–209.

- Vindstad, O. P. L., Jepsen, J. U., Ek, M., Pepi, A., & Ims, R. A. (2019). Can novel pest outbreaks drive ecosystem transitions in northern-boreal birch forest? *Journal of Ecology*, *107*, 1141–1153.
- Wallander, H., Ekblad, A., Godbold, D. L., Johnson, D., Bahr, A., Baldrian, P., Bjork, R. G., Kieliszewska-Rokicka, B., Kjoller, R., Kraigher, H., Plassard, C., & Rudawska, M. (2013). Evaluation of methods to estimate production, biomass and turnover of ectomycorrhizal mycelium in forests soils - a review. *Soil Biology & Biochemistry*, *57*, 1034–1047.
- Wang, Y., Naumann, U., Wright, S. T., & Warton, D. I. (2012). Mvabund—An R package for model-based analysis of multivariate abundance data. *Methods in Ecology and Evolution*, *3*, 471–474.
- Warton, D. I., Thibaut, L., & Wang, Y. A. (2017). The PIT-trap—A “model-free” bootstrap procedure for inference about regression models with discrete, multivariate responses. *PLoS ONE*, *12*, e0181790.
- White, T. J., Bruns, T. D., Lee, S. B., & Taylor, J. W. (1990). *Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics*. Academic Press, Inc.
- Wookey, P. A., Aerts, R., Bardgett, R. D., Baptist, F., Brathen, K. A., Cornelissen, J. H. C., Gough, L., Hartley, I. P., Hopkins, D. W., Lavorel, S., & Shaver, G. R. (2009). Ecosystem feedbacks and cascade processes: Understanding their role in the responses of Arctic and alpine ecosystems to environmental change. *Global Change Biology*, *15*, 1153–1172.
- Zak, D. R., Pellitier, P. T., Argiroff, W. A., Castillo, B., James, T. Y., Nave, L. E., Averill, C., Beidler, K., Bhatnagar, J., Blesh, J., Classen, A. T., Craig, M., Fernandez, C. W., Gundersen, P., Johansen, R., Koide, R. T., Lilleskov, E. A., Lindahl, B. D., Nadelhoffer, K. J., ... Tunlid, A. (2019). Exploring the role of ectomycorrhizal fungi in soil carbon dynamics. *New Phytologist*, *223*, 33–39.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Parker, T. C., Chomel, M., Clemmensen, K. E., Friggens, N. L., Hartley, I. P., Johnson, D., Kater, I., Krab, E. J., Lindahl, B. D., Street, L. E., Subke, J.-A., & Wookey, P. A. (2022). Resistance of subarctic soil fungal and invertebrate communities to disruption of below-ground carbon supply. *Journal of Ecology*, *110*, 2883–2897. <https://doi.org/10.1111/1365-2745.13994>