

Accepted refereed manuscript of:

Van Tien N, Chinh DTM, Huong TTM, Phuong TH, Irvin S & Glencross B (2016) Development of a nutritional model to define the energy and protein requirements of cobia, *Rachycentron canadum*, *Aquaculture*, 463, pp. 193-200.

DOI: [10.1016/j.aquaculture.2016.05.037](https://doi.org/10.1016/j.aquaculture.2016.05.037)

© 2016, Elsevier. Licensed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International  
<http://creativecommons.org/licenses/by-nc-nd/4.0/>

1                   **Development of a nutritional model to define the energy and protein**  
2                   **requirements of cobia, *Rachycentron canadum***

3  
4  
5   Nguyen Van Tien<sup>1\*</sup>, Dam Thi My Chinh<sup>1</sup>, Tran Thi Mai Huong<sup>1</sup>, Truong Ha Phuong<sup>2</sup>, Simon Irvin<sup>3</sup>  
6   and Brett Glencross<sup>3,4</sup>

7  
8       1        Research Institute for Aquaculture No1, Dinh Bang, Tu Son, Bac Ninh Province, Vietnam

9       2        Research Institute for Aquaculture No3, Nha Trang, Khanh Hoa Province, Vietnam

10      3        CSIRO Aquaculture Program, PO Box 2583, Brisbane, QLD 4001, Australia

11      4        Present address; Institute of Aquaculture, University of Stirling, Stirling FK9 4LA,  
12              Scotland, United Kingdom

13  
14   \* Author for correspondence

15                   (e) [biotien@yahoo.com](mailto:biotien@yahoo.com)

16                   (p) +84-3827-3069

17                   (f) +84-3827-3070

18  
19  
20   Keywords : Cobia, nutritional model, energetics, protein requirements, specifications, bioenergetic

1 **Abstract**

2

3 This study assessed the protein and energy requirements of Cobia (*Rachycentron canadum*) using a  
4 bio-energetic factorial approach. Using a series of inter-related studies, several parameters were  
5 defined to enable the construction of a bio-energetic factorial model for this species. The studies  
6 included two controlled laboratory experiments and also extensive field-data collection from  
7 commercial and research farms in Vietnam. The devised model includes parameters for both  
8 maintenance and protein demands; the effect of fish live-weight on maintenance protein ( $LW^{0.697}$ ),  
9 lipid ( $LW^{0.972}$ ), and energy demands ( $LW^{0.815}$ ); the efficiencies of protein, lipid and energy utilisation  
10 at various protein, lipid and energy intake levels; and the variability in whole body composition with  
11 varying live-weight. The protein utilisation efficiencies ( $0.456 \cdot [\text{protein intake}] - 0.445$ ), lipid  
12 utilisation efficiencies ( $1.292 \cdot [\text{lipid intake}] - 1.120$ ) and energy utilisation efficiencies ( $0.651 \cdot$   
13  $[\text{energy intake}] - 48.41$ ) were similar to other carnivorous fish species. However, the maintenance  
14 requirements for both energy ( $74.3 \text{ kJ/ kgBW}^{0.8}/ \text{d}$  at  $28^\circ\text{C}$ ) and protein ( $0.99 \text{ g/ kgBW}^{0.7}/ \text{d}$  at  $27.9^\circ\text{C}$ )  
15 were about double to other species. Using this modelling approach it was possible to iteratively derive  
16 optimal dietary protein and energy specifications for this species.

17

## 1    **1.    Introduction**

2  
3            *Cobia* (*Rachycentron canadum*) is the only species in the family Rachycentridae. The species  
4 is distributed worldwide in warm marine waters, except for the central and eastern Pacific. The  
5 species is generally regarded as a fast growing, tropical pelagic animal. In offshore net cage systems,  
6 cobia can grow from 0.5 kg fingerling to 6.0 to 8.0 kg marketable size within 6 to 8 months with a  
7 feed conversion ratio of 1.5 (Liao *et al.*, 2004) or 6 kg after 1 year at 28°C (Benetti *et al.*, 2010). Due  
8 to their high quality white flesh, cobia is suitable for sashimi or fillet production (Chou *et al.*, 2001).  
9 The global aquaculture production of *Cobia* has increasing rapidly from 2002, reaching to 41,774 MT  
10 in 2012 (FAO, 2014). The three main producers of cobia in 2012 were China, Taiwan and Vietnam,  
11 where annual production was approximately 38,014 metric tons (MT), 1,384 MT and 2,000 MT,  
12 respectively (FAO, 2014). While *Cobia* cultured in offshore net cage systems is generally reared using  
13 formulated feeds (Liao *et al.*, 2004), most cobia production in traditional inshore sea cages is still  
14 based on trash fish (Petersen *et al.*, 2015). Currently, the limited supply of trash fish as the main feed  
15 source for cobia grow-out has become a major constraint for cobia culture in Viet Nam and other  
16 countries.

17            *Cobia* culture has been rapidly gaining in popularity since the early 1990s, but formulated  
18 feed development for aquaculture of this species is still lagging behind compared with other fish  
19 species such as salmon or barramundi (Zhou *et al.*, 2007; Xiao *et al.*, 2009; Liu *et al.*, 2010). Despite,  
20 many studies have been undertaken to identify a range of nutritional requirements of this species, the  
21 energy and protein requirements are still undefined and pelleted feed are still not well established  
22 (Salze *et al.*, 2010). Earlier studies have suggested that the optimum dietary protein and lipid levels in  
23 juvenile cobia were 45% and 5–15% dry weight, respectively (Chou *et al.*, 2001; Craig *et al.*, 2006).  
24 Maximum growth and the best feed conversion ratios have been recorded at 27–29°C in juvenile  
25 cobia with an optimum feed ration level determined at 9% initial body weight per day for fish of 10-  
26 200g live-weight (Sun *et al.*, 2006; Webb, 2009; Sun and Chen, 2014).

27            The requirements for protein and energy for most aquaculture species have traditionally been  
28 determined using empirical dose-response studies (Mercer, 1982). More recently, the use of bio-  
29 energetic factorial modelling has proven to be a useful alternative method in estimating these  
30 requirements (Shearer, 1995; Glencross, 2008; Trung *et al.*, 2011). The benefits of bio-energetic  
31 factorial modelling are that it provides a method for estimating nutritional requirements independent  
32 of animal size and it results in a series of nutrient specifications that are indexed against energy  
33 demand and as such it underpins the potential for a wide range of diet specifications to be developed  
34 subject to different formulation strategies (Lupatsch *et al.*, 2003; Booth *et al.*, 2010; Glencross *et al.*,  
35 2011). Additionally, this modelling approach also has an advantage over an empirical approach in that  
36 it can also be used to define the optimal feed rations as well as specifications. This has further merits  
37 in that total nutrient and energy budgets, including losses through wastage and excretion, and also raw

1 material demands can be determined and strategies examined by which to improve fish production  
2 (Glencross, 2010).

3         This paper describes a series of studies designed to determine the energy and protein  
4 requirements of cobia (*R. canadum*). Using farm-collected data, samples and experiments from both  
5 Vietnam and Australia, and a series of studies undertaken to determine key parameters of the model.  
6 These parameters include; the estimation of growth potential of fish with varying size, changes in  
7 body protein and energy composition with fish size; determination of the energy and protein  
8 requirements for maintenance, determination of the protein and energy digestibility of a reference  
9 diet, and determination of the partial efficiencies of both protein and energy utilisation. From this  
10 series of studies the results are then integrated to present an iterative approach to the determination of  
11 the protein and energy requirements for this species over a range of fish sizes.

12

## 2. Methods

### 2.1 Study 1 – Endogenous losses of protein, lipid and energy

This experiment was conducted at the Cat Ba National Broodstock Center of Marine Aquaculture of the Research Institute for Aquaculture - 1 (RIA-1), in Vietnam. Twelve 1,000 L tanks were each stocked with ten cobia (*R. canadum*). Fish sizes within each tank were in one of four general size classes (100 g, 200 g, 500 g and 1,000 g fish<sup>-1</sup>), with three replicates being used for each size class. Additional fish (n=5 for each size class) of similar approximate weights to those four size classes were euthanized at the beginning of the study to determine the dry matter, ash, protein, lipid and energy composition of the fish at the beginning of the study. The experimental tanks were supplied with aeration, flow-through marine water (salinity 32PSU) at 28.4 ± 1.58°C. The transferred fish were kept in the tanks for 21 days, without feeding. After this period the fish were re-weighed and all fish from each tank were used as a replicate to determine weight, energy, lipid and protein loss. Following weighing five of the fish from each size class were euthanized, pooled and assessed for composition change in dry matter, ash, lipid, protein and energy concentrations.

### 2.2 Study 2 – Energy and protein digestibility

This study was conducted at the Cleveland Laboratory of the CSIRO Aquaculture Program in Australia. A single basal diet was formulated to provide protein and lipid at 489 g/kg and 138 g/kg diet at a gross energy level of 22.2 MJ kg<sup>-1</sup> (estimated digestible protein and energy of 406 g kg<sup>-1</sup> and 19.7 MJ kg<sup>-1</sup>, respectively) (Table 1). The dry ingredients were first blended in a series of batches using a 60 L upright Hobart mixer (HL600, Hobart, Pinkenba, QLD, Australia), to produce a single batch of basal mash which was extruded using a laboratory-scale, twin-screw extruder with intermeshing, co-rotating screws (MPF19:25, Baker Perkins, Peterborough, United Kingdom). The resultant pellets produced through a 3 mm Ø die were cut into 4 to 5 mm lengths using a four-bladed variable speed cutter and collected and dried at 60°C for 12 h in a fan-forced drying oven. The remaining oil allocation was vacuum infused post-drying according to the methods reported by Diu et al (2015).

#### TABLE 1 HERE

Three 100 L tanks of flow through seawater (27.9 ± 0.32°C) were each stocked with 10 juvenile (~200 g) fish. The transferred fish were allowed to acclimate to the tanks and were fed the reference diet for 23 days before faecal collection was initiated. Faeces were collected using stripping techniques similar to that used for barramundi (Blyth *et al.*, 2014).

Diet and faecal samples were analysed for dry matter, yttrium, protein, total lipid, gross energy and ash content (AOAC, 2005). Differences in the concentrations of the protein, lipid, energy and yttrium in the feed and faeces on a dry matter basis in each treatment were calculated to

1 determine the apparent digestibility ( $AD_{\text{diet}}$ ) of each nutritional parameter. Those digestibilities  
2 examined were based on the following equation:

$$ADC = \left( 1 - \frac{Y_{\text{feed}} \times \text{Parameter}_{\text{faeces}}}{Y_{\text{faeces}} \times \text{Parameter}_{\text{feed}}} \right)$$

5  
6 Where  $Y_{\text{diet}}$  and  $Y_{\text{faeces}}$  represent the yttrium content of the diet and faeces respectively, and  
7  $\text{Parameter}_{\text{diet}}$  and  $\text{Parameter}_{\text{faeces}}$  represent the nutritional parameter of concern (protein, lipid or  
8 energy) content of the diet and faeces respectively.

### 10 2.3 Study 3 – Energy and protein utilisation efficiency

11 This study was conducted at the Cleveland Laboratory of the CSIRO Aquaculture Program in  
12 Australia. Twenty four 100 L tanks were each stocked with 10 cobia juveniles ( mean weight  $136.2 \pm$   
13  $0.71$  g) . A series of six feed ration treatments were assigned in quadruplicate to the array. The same  
14 diet as used in the digestibility study was used in this study (Table 1). Each ration level was  
15 determined based on satiety, 80%, 60%, 40%, 20% of satiety and starved. The sub-satiety levels were  
16 estimated based on feed intake measured in the three days preceding the initiation of the experiment  
17 when fish were being acclimated to the tanks. Water temperature was maintained at  $27.9 \pm 0.32^\circ\text{C}$  for  
18 the duration of the study. The trial was run for 23 days to minimize the time that fish were unfed  
19 before a result could be obtained. The apparent satiety ration level was determined based on the loss  
20 of feeding activity after the fish being offered food on three or more independent feeding episodes  
21 within a one-hour period. Any uneaten food was collected by siphoning and accounted for. After 23  
22 days the weight gain was assessed by weighing all fish within each tank to determine tank mean  
23 weight gain. At this point three fish from each tank were also euthanized and whole fish samples were  
24 collected for the analysis of dry matter, protein, lipid and energy content.

### 26 2.4 Study 4 – Fish composition variation

27 A range of sizes of cobia from 25 g to 2013 g were collected (n=18) from both laboratory  
28 stocks and commercial grow-out producers in northern Vietnam, with further fish also sourced from  
29 the Bribie Island Research Centre in Woorim, QLD, Australia. Whole fish were minced and then  
30 analysed for dry matter, protein, lipid and energy content. These fish were obviously fed a range of  
31 diets, therefore representing the average genetic response of the species to variations in dietary protein  
32 and energy balance provision in diets. All analyses were conducted according to the methods  
33 specified by the AOAC (2005).

### 35 2.5 Study 5 – Assessment of fish growth rates

1 Growth rates were assessed from a combination of both farm and laboratory data sources.  
2 Eight commercial sea cage production facilities in Khanh Hoa, Cat Ba and Nghe An provinces in  
3 Vietnam were each assessed at monthly intervals (from April 2010 to April 2011) by weighing around  
4 15 fish from each cage to determine mean weight gain and daily growth rates ( $\text{g day}^{-1}$ ). Growth rates  
5 (range 0.23 to 17.77  $\text{g day}^{-1}$ ) were expressed relative to the geometric mean weight (range 4 to 5,040 g  
6  $\text{fish}^{-1}$ ) of the fish from each measurement. Water temperature (range 18.0 to 29.5 °C, mean  $\pm$  SD =  
7  $25.6 \pm 2.8^\circ\text{C}$ ) was also measured at each sampling time.

## 8 9 2.6 *Chemical analyses*

10 Fish and samples of the reference diet were analysed for dry matter, protein, total lipids and  
11 energy content. Diet and faecal samples were also analysed for yttrium. Dry matter was calculated by  
12 gravimetric analysis following oven drying at 105°C for 24 h. Gross energy content was determined  
13 using ballistic bomb calorimetry or in some cases calculated using protein = 23.6  $\text{kJ g}^{-1}$ , lipid = 38.5  
14  $\text{kJ g}^{-1}$  and carbohydrate = 17.3  $\text{kJ g}^{-1}$ . Protein levels were calculated from the determination of total  
15 nitrogen by combustion analysis using a CHNOS autoanalyser in Australia and Kjeldhal in Vietnam  
16 and multiplying N by 6.25. Total lipid contents were determined gravimetrically following extraction  
17 by chloroform and methanol (2:1 v/v) solution according to the method of Folch (1957). Total  
18 yttrium concentrations were determined after digestion with concentrated nitric acid (60%) using  
19 inductively coupled plasma atomic emission spectrophotometry (ICP-AES). Carbohydrate was  
20 determined as the difference in dry matter content minus protein, ash and total lipids. All of these  
21 determinations were conducted according to the methods specified by the AOAC (2005).

## 22 23 2.7 *Statistical analysis*

24 All values are mean  $\pm$  SEM unless otherwise specified. Fish weights were converted to the  
25 geometric mean (GMW) of initial and final weights prior to plotting on the figures. Graphical  
26 presentation was done using Microsoft Excel. Regression analysis on linear, power and polynomial  
27 functions (including derivation of error terms) was done using Statistica version 9.0 (StatSoft, Tulsa,  
28 OK, USA). Error terms for exponents were determined based on natural logarithmic transformations  
29 of the data prior to linear regression analysis.

### 3. Results

#### 3.1 Study 1 – Starvation energy and protein losses

Protein, lipid and energy losses by cobia over a 21 d period of starvation were shown to vary with the size of the fish (Figures 1, 2 and 3). Overall endogenous protein loss was greater with increasing fish size, and is expressed as an exponential relationship (Equation 1). Concomitant with that protein loss, was also the loss of lipid during this starvation period (Equation 2). The loss of these two nutrients due to starvation is expressed as the energy loss (Equation 3). A greater protein and lipid losses were recorded in larger fish (Fig. 1, 2). Similar to protein losses, the relationship with size and energy losses was also described by an exponential relationship with body weight (BW) (Equation 3). The determined exponents of protein ( $BW^{0.697}$ ), lipid ( $BW^{0.989}$ ), and energy ( $BW^{0.822}$ ), loss were so similar to standard exponents ( $BW^{0.70}$ ,  $BW^{1.00}$  and  $BW^{0.80}$  respectively) for other fish species that it was decided to standardise their use to these common exponents for further calculations.

(Equation 1) Protein loss (g/fish) =  $0.235 \cdot (GMW)^{0.697}$ , ( $R^2 = 0.844$ )

(Equation 2) Lipid loss (g/fish) =  $0.021 \cdot (GMW)^{0.989}$ , ( $R^2 = 0.870$ )

(Equation 3) Energy loss (kJ/fish) =  $4.902 \cdot (GMW)^{0.822}$ , ( $R^2 = 0.874$ )

FIGURES 1, 2 AND 3 HERE

#### 3.2 Study 2 – Energy, lipid and protein digestibility

Protein, lipid and energy digestibility values of the reference diet were determined. Protein digestibility was measured at  $83.0 \pm 0.2\%$ . Lipid digestibility was measured at  $94.6 \pm 0.8\%$  and energy digestibility was measured at  $89.0 \pm 0.1\%$ . This equated to a digestible protein content of the reference diet of 40.6%, a digestible lipid content of 13.0% and a digestible energy content of 19.7 MJ/kg, each on a dry matter basis.

#### 3.3 Study 3 – Energy and protein utilisation efficiency

The measured utilisation of protein, lipid and energy by cobia, was based on the assessment of net gain in each parameter relative to the varying intake of dietary digestible protein (Figure 4; Equation 4), digestible lipid (Figure 5; Equation 5) and digestible energy (Figure 6; Equation 6). Each relationship followed a linear function.

(Equation 4) Protein gain (g/  $kg^{0.70}$ / d) =  $0.456 \cdot (\text{digestible protein intake}) - 0.450$ , ( $R^2 = 0.938$ )

(Equation 5) Lipid gain (g/  $kg^{1.00}$ / d) =  $1.292 \cdot (\text{digestible energy intake}) - 1.120$ , ( $R^2 = 0.973$ ).

(Equation 6) Energy gain (kJ/  $kg^{0.80}$ / d) =  $0.651 \cdot (\text{digestible energy intake}) - 48.411$ , ( $R^2 = 0.996$ )

FIGURES 4, 5 AND 6 HERE

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37

The intercept of each linear function with the X-axis was used to determine the maintenance requirements for each of protein, lipid and energy.

### 3.4 Study 4 – Fish composition

The composition of the fish varied over the live-weight range of fish examined (Figure 7). Protein content was relatively constant and was described by a linear function (Equation 7). Typically, the live-weight total lipid composition was also observed to increase with increasing live-weight (Figure 7; Equation 8). The increase in total lipid content with increasing fish size was also consistent with an increase in energy density of the fish (Figure 7; Equation 9).

$$\text{(Equation 7) Live-weight protein (\%)} = 0.001 \cdot (\text{live-weight}) + 0.168, (R^2 = 0.007)$$

$$\text{(Equation 8) Live-weight lipid (\%)} = 0.0054 \cdot (\text{live-weight})^{0.451}, (R^2 = 0.822)$$

$$\text{(Equation 9) Live-weight energy (MJ/ kg)} = 2.669 \cdot (\text{live-weight})^{0.1715}, (R^2 = 0.791)$$

FIGURE 7 HERE

### 3.5 Study 5 – Assessment of fish potential growth rate

The growth rates of juvenile cobia showed that larger fish typically had the potential to gain greater total biomass per day than smaller fish (Figure 8). The temperature independent function for this growth can be expressed as Equation 10. This equation derived from growth data of cobia cultured at an average water temperature  $25.6 \pm 2.8^\circ\text{C}$ . Where the geometric mean weight (GMW) is in gram per fish.

$$\text{(Equation 10) Fish growth rate (g/d)} = 0.159 \cdot (\text{GMW})^{0.574} (R^2 = 0.847)$$

FIGURE 8 HERE

### 3.6 Study 6 – Iterative design of dietary protein and energy specifications

From the starvation data and the calculated maintenance protein, lipid and energy demands a function describing the relationship between fish live-weight and those maintenance demands was derived. Because insufficient data was collected on temperature effects on maintenance demands the determination of a temperature response was not attempted.

$$\text{(Equation 11) Maintenance Protein Demand (g/ d/ fish)} = 0.99 \cdot (\text{live-weight})^{0.70}$$

$$\text{(Equation 12) Maintenance Lipid Demand (g/ d/ fish)} = 0.87 \cdot (\text{live-weight})^{1.00}$$

$$\text{(Equation 13) Maintenance Energy Demand (kJ/ d/ fish)} = 74.3 \cdot (\text{live-weight})^{0.80}$$

1

2 TABLE 2 HERE

3

## 4. Discussion

Factorial models have proven to be useful in defining both protein and energy demands and total feed ration management for a range of fish species (Lupatsch *et al.*, 2003; Glencross, 2008; Pirozzi *et al.*, 2010). This study reports on the development of a model for a new carnivorous fish species and as such adds to the volume of data on such fish species.

### 4.1 Comparisons to the growth model

There are actually only a few studies examining growth by cobia in the literature. A study by Benetti *et al.*, (2010) reported the growth of cobia cultured in open ocean submerged cages in the Caribbean. Growth rates in that study were consistently 94% (at 27.8°C) and 82% (at 25.5°C) of those reported in the present model at water temperature of 27.8°C. A common issue with the comparison of the growth model against much of the published literature is that most of the published literature appears to be with very small fish and often under limiting conditions of feed quality situations and with fish growing much slower than that encountered from our farm-based data collection. Such vagaries in the growth rates throughout the literature highlight the need to develop a benchmark standard against which laboratory studies should be compared.

### 4.2 Protein requirements

The metabolic weight exponent for protein metabolism in cobia is 0.697. This is similar to the generic protein exponent for most fish species is 0.70. The efficiency of protein use by cobia, based on the regression of the protein gain against the digestible protein intake, was linear over the protein intake range examined and had a coefficient of 0.456. This coefficient value for the partial efficiency of protein gain for this species is also similar to that observed for most other fish species – barramundi: 0.48, gilthead seabream: 0.53, rainbow trout: 0.40 - 0.47, yellowtail kingfish: 0.51, (Lupatsch *et al.*, 2003; Glencross, 2008; 2009; Glencross *et al.*, 2008; Booth *et al.*, 2010). Although in most other studies this relationship between protein gain and protein intake has usually been observed to be curvilinear, in the present study this response was linear over the feed intake ranges studied (Lupatsch *et al.*, 2003; Glencross, 2008; Dumas *et al.*, 2010; Glencross, 2010; Glencross *et al.*, 2011). Such linear responses have been observed before (Lupatsch *et al.*, 2001). Though it has been argued that such linear responses are indicative of underfeeding as even the curvilinear responses reported are close to linear at the lower levels of feed intake (Glencross and Bermudes, 2012).

A notable feature of this study was the higher maintenance protein requirements ( $DP_{\text{maint}}$ ) observed of this species. Based on the point of zero net protein gain a  $DP_{\text{maint}}$  intake of 0.99 g/ kg<sup>0.70</sup>/d was calculated (Figure 4). This is about 50% higher than the value of 0.66 g /kg<sup>0.70</sup>/d determined for *D. labrax* (Lupatsch *et al.*, 2001), and double the 0.45 g /kg<sup>0.70</sup>/d determined for barramundi

1 (Glencross, 2008). However, it is only about half that reported for yellowtail kingfish (1.70 g  
2 /kg<sup>0.70</sup>/d), another highly active pelagic carnivorous species (Booth *et al.*, 2010).

### 3 4 4.3 Energy requirements

5 The relationship between this specie's energy metabolism and its body weight also conform  
6 to the allometric equation:  $a \cdot BW(\text{kg})^b$  as is the case for virtually every other fish species studied  
7 (Withers, 1998; Dumas *et al.*, 2010). Similarly, the exponent value of body weight ( $BW^{\text{exponent value}}$ ) for  
8 energy metabolism in cobia was observed to be 0.822 which is similar to the result determined by  
9 Watson and Holt (2010) using indirect calorimetry with this species (0.809). It is also similar to other  
10 fish species including barramundi (0.80), grouper (0.79), gilthead seabream (0.82), European seabass  
11 (0.80), Pangasius catfish (0.84) and tilapia (0.85) (Lupatsch *et al.*, 2003; Glencross, 2008; Glencross  
12 and Bermudes, 2011; Glencross *et al.*, 2011; Trung *et al.*, 2011).

13 The maintenance energy requirements ( $DE_{\text{maint}} = 74.3 \text{ kJ/kg}^{0.80}/\text{d}$ ), as defined by the point of  
14 zero net energy gain, in this study was substantially higher from that seen for other species like  
15 rainbow trout (40.1 kJ / kg<sup>0.80</sup> /d), barramundi (42.6 kJ / kg<sup>0.80</sup> /d) and mulloway (26.3 kJ / kg<sup>0.80</sup> /d)  
16 (Glencross *et al.*, 2008; Glencross, 2008; Pirozzi *et al.* 2010). However, the  $DE_{\text{maint}}$  was similar to the  
17 87.4 kJ/kg<sup>0.80</sup>/d reported by Booth *et al.* (2010) for another pelagic carnivorous fish species the  
18 yellowtail kingfish (*Seriola lalandi*). This observation poses a question whether it is this active  
19 pelagic nature of these animals that results in such a higher or some other feature like the partial  
20 endothermy observed in some Scombrid species (Glencross *et al.*, 2001).

21 The partial efficiency of energy use is determined as the slope of the regression of the energy  
22 intake against energy retention, on a metabolic body weight basis (Lupatsch *et al.*, 2001). In the  
23 present study for cobia species, the response of full energy intake range was recorded to be linear.  
24 This contrasts with the curvilinear response observed with other species (Lupatsch *et al.*, 2003;  
25 Bureau *et al.* 2006; Glencross, 2008; Glencross *et al.*, 2008; Trung *et al.*, 2011), but is consistent with  
26 the linear response reported in other studies (Cho & Bureau, 1998; Lupatsch *et al.*, 2001).

27 In the present study, the partial efficiency of energy gain was observed to be 0.651. This value  
28 is consistent with other carnivorous fish species e.g. Gilthead Seabream (0.65), white grouper,  
29 *Epinephelus aeneus* (0.69), barramundi (0.68), rainbow trout (0.62), yellowtail kingfish (0.65) and  
30 mulloway (0.60) (Lupatsch *et al.*, 2003; Glencross, 2008; 2009; Booth *et al.*, 2010; Pirozzi *et al.*,  
31 2010).

### 32 33 4.4 Iterative diet design

34 Key dietary parameters of energy and protein specifications can be derived iteratively from  
35 this model for fish at any phase of its production cycle (Glencross, 2008; Booth *et al.*, 2010;  
36 Glencross *et al.*, 2010). This iterative approach was also used to define the energy and protein  
37 requirements for cobia from 100g to 2000g at each of three dietary energy densities (Table 2). Based

1 on a combination of the somatic and non-somatic (maintenance) energy demands a simplistic energy  
2 budget was created that dictates how much energy the fish needs to consume to achieve a prescribed  
3 growth potential. The amount of feed (g/fish) rationed to the animal then being this energy demand  
4 divided by the digestible energy density of that feed (Table 2).

5 Similarly, the needs for protein for both somatic and non-somatic demands can also be  
6 defined using this approach which defines the appropriate DP:DE ratio (Table 2). Using the  
7 empirically derived equations from studies 1 to 5 the requirements for protein and energy at a range of  
8 fish sizes was determined (Table 2). Based on a combination of the predicted growth, the protein and  
9 energetic cost of that weight gain, the efficiencies associated with those gains and the maintenance  
10 requirements, the total daily requirements for both protein and energy at a range of fish sizes were  
11 calculated (Table 2). From this both the daily energy and protein intake requirement were defined.  
12 This has subsequently allowed us to iteratively specify a series of hypothetical diets of varying energy  
13 density (12 MJ/kg, 16 MJ/kg and or 20 MJ/kg) (Table 2).

14 In applying this iterative approach, it is assumed that the fish will eat to an energetic demand  
15 and as such the energy content of each diet will define total feed consumption. This total feed  
16 consumption also influences the amount of dietary protein required to satisfy the daily protein demand  
17 (Dumas *et al.*, 2010).

18 Using this iterative approach the present study shows that there are several strategies that can  
19 be employed to define the theoretically optimal diet energy and protein specifications and that these  
20 change with fish size, consistent with what has been reported in numerous other similar studies  
21 (Lupatsch *et al.*, 2003; Glencross, 2008; Booth *et al* 2010; Trung *et al.*, 2011; Glencross and  
22 Bermudes, 2012). When the diet energy density and/or fish size varies the present model demonstrates  
23 that there is a need to vary the dietary protein supply for this species. This model also demonstrates  
24 how the choice of diet energy density has an effect on the biological feed conversion ratio (FCR).  
25 When a lower FCR is achieved with a higher energy density simply due to the energetic demands  
26 being satisfied by fewer grams of feed. Importantly though, this lower feed ration combined with the  
27 same daily protein requirement also means that the protein concentration required in that diet for it to  
28 satisfy the daily protein demands has to increase for it to be effective. Similar to other species, it was  
29 noted that the most dramatic changes in the protein demand (based on the required protein : energy  
30 ratio) of cobia occur over the first 500 g of its growth, where the optimal DP:DE changes from 36  
31 g/MJ at 50 g to 24 g/MJ at 500 g (Figure 9).

32  
33 FIGURE 9 HERE

34  
35 For cobia, the optimal DP:DE ratios at 100 g and 1000 g were 32 and 22 g/MJ, respectively  
36 (Table 2) and by comparison barramundi optimal DP:DE ratios at 100 g and 1000 g were 30.2 and  
37 19.9 g/MJ (Glencross, 2008). This contrasts those determined for yellowtail kingfish which had

1 optimal DP:DE ratios at 100 g and 1000 g of 39 and 27 g/MJ, respectively (Booth *et al.*, 2010). It can  
2 be seen that for each of the sizes of cobia, examined in the present study that the optimal DP:DE  
3 ratios were marginally higher than those of barramundi, but substantially lower than those of  
4 yellowtail kingfish (Figure 9).

#### 6 4.4 Conclusions

7 This study used a factorial method for determining the protein and energy requirements for  
8 cobia. This study adds to the volume of literature using this method to estimate these requirements for  
9 a range of fish species. Comparison of the data derived from this study with that obtained for other  
10 species indicates a high degree of homology of most energetic parameters. The primary difference, in  
11 comparison to the many other models developed for most other carnivorous fish species, is that this  
12 species has a marginally higher demand for protein, but most notably its maintenance requirements  
13 for protein and energy are substantially higher than other studied species. The only exception to this  
14 being the comparison with another pelagic marine fish, the yellowtail kingfish, which also has  
15 similarly high maintenance demands.

16 This study represents a series of estimations based on a series of inter-related studies and their  
17 derived parameters. As such the estimations deduced from this modelling exercise are only as robust  
18 as the weakest data estimates. It would be prudent to take the outputs from this model and  
19 independently validate them and also test some of the assumptions used to increase the robustness of  
20 this model.

#### 22 Acknowledgement

23 The authors would like to sincerely thank the Australian Centre for International Agricultural  
24 Research (ACIAR) for funding this project (FIS/2006/141), and all project collaborators at CSIRO,  
25 Australia and Research Institute for Aquaculture No 1, Viet Nam who have contributed to data  
26 collection, sample analysis and who have given constructive comments on this manuscript.

1 **References**

- 2 AOAC (Association of Official Analytical Chemists). 2005. Official Methods of Analysis of the  
3 Association of Official Analytical Chemists. 15<sup>th</sup> edition. Association of Official Analytical  
4 Chemists. Washington, DC, USA.
- 5 Benetti, D.D., B. O'Hanlon, J.A. Rivera, A.W. Welch, C. Maxey, Orhun M.R., 2010. Growth rates of  
6 cobia (*R. canadum*) cultured in open ocean submerged cages in the Caribbean. *Aquaculture*  
7 302, 195-201.
- 8 Bermudes, M., Glencross, B.D., Austen, K., Hawkins, W., 2010. Effect of high water temperatures on  
9 nutrient and energy retention in barramundi (*Lates calcarifer*). *Aquaculture* 306, 160-166.
- 10 Blyth, D., Tabrett, S.J., Glencross, B.D., 2014. A study of the effects of faecal collection method and  
11 acclimation time on the digestibility of diets and ingredients when fed to juvenile barramundi  
12 (*Lates calcarifer*). *Aquaculture Nutrition* 21, 248–255. doi: 10.1111/anu.12159.
- 13 Booth, M.A., Pirozzi, I., Allan, G.A., 2010. Estimation of digestible protein and energy requirements  
14 of yellowtail kingfish *Seriola lalandi* using a factorial approach. *Aquaculture* 307, 247-259.
- 15 Bureau, D.P., Hua, K., Cho, C.Y., 2006. Effect of feeding level on growth and nutrient deposition in  
16 rainbow trout (*Oncorhynchus mykiss* Walbaum) growing from 150 to 600 g. *Aquacult. Res.*, 37,  
17 1090–1098.
- 18 Cho, C.Y., Bureau, D.P., 1998. Development of bioenergetic models and the Fish-PrFEQ software to  
19 estimate production, feeding ration and waste output in aquaculture. *Aquatic Living Resources*  
20 11, 199-210.
- 21 Chou R.L, Su M.S., Chen H.Y., 2001. Optimal dietary protein and lipid levels for juvenile cobia  
22 (*Rachycentron canadum*). *Aquaculture* 193 (1-2), 81-89.
- 23 Chou, R.-L., M.-S. Su, Chen H.Y., 2001. Optimal dietary protein and lipid levels for juvenile cobia  
24 (*Rachycentron canadum*). *Aquaculture* 193, 81-89.
- 25 Craig, S.R., Schwarz, M.H., McLean, E., 2006. Juvenile cobia (*Rachycentron canadum*) can utilize a  
26 wide range of protein and lipid levels without impacts on production characteristics.  
27 *Aquaculture* 261, 384–391.
- 28 Diu, N.T., Pirozzi, I., Glencross, B.D., 2015. Digestibility of canola meals in barramundi (Asian  
29 seabass; *Lates calcarifer*). *Aquaculture* 435, 442–449.

- 1 Dumas, A., France, J., Bureau, D.P., 2010. Modelling growth and body composition in fish nutrition:  
2 Where have we been and where are we going? *Aquaculture Research* 41, 161-181.
- 3 FAO 2014. Fisheries and aquaculture software. FishStatJ - software for fishery statistical time  
4 series. In: FAO Fisheries and Aquaculture Department [online]. Rome. Updated 22 July 2014.
- 5 Folch, J., Lees, M., Sloane-Stanley, G.H., 1957. A simple method for the isolation and purification of  
6 total lipides from animal tissues. *J. Biol. Chem.* 226, 497–509.
- 7 Fraser, T.W.K., Davies S.J., 2009. Nutritional requirements of cobia, *Rachycentron canadum*  
8 (Linnaeus): a review. *Aquaculture Research* 40, 1219-1234.
- 9 Glencross, B.D., Carter, C.G., Gunn, J, van Barneveld, R.J., Rough, K.M., Clarke, S.M., 2001.  
10 Southern bluefin tuna. In: *Nutrient requirements and Feeding of Finfish for Aquaculture*  
11 (Webster and Lim, Eds). CABI Publishing, Wallingford, UK. pp 112-124.
- 12 Glencross B.D., Michael R., Austen K., Hauler R., 2008. Productivity, carcass composition, waste  
13 output and sensory characteristics of large barramundi *Lates calcarifer* fed high-nutrient  
14 density diets. *Aquaculture* 284 (1-4),167-173.
- 15 Glencross, B.D., 2008. A factorial growth and feed utilisation model for barramundi, *Lates calcarifer*  
16 based on Australian production conditions. *Aquaculture Nutrition* 14, 360-373.
- 17 Glencross, B.D., 2009. Reduced water oxygen levels affect maximal feed intake, but not protein or  
18 energy utilisation by rainbow trout (*Oncorhynchus mykiss*). *Aquaculture Nutrition* 15, 1-8.
- 19 Glencross, B.D., 2010. Using modelling approaches to understand the implications of physiological  
20 challenges and raw material demands on aquaculture feed designs. In: Cruz-Suarez, L.E.,  
21 Ricque-Marie, D., Tapia-Salazar, M., Nieto-López, M.G., Villarreal-Cavazos, D. A., Gamboa-  
22 Delgado, J. (Eds), *Avances en Nutrición Acuícola X - Memorias del Décimo Simposio*  
23 *Internacional de Nutrición Acuícola*, 8-10 de Noviembre, San Nicolás de los Garza, N. L.,  
24 México. ISBN en trámite. Universidad Autónoma de Nuevo León, Monterrey, México, pp. 341-  
25 365.
- 26 Glencross B.D. and Bermudes M., 2011. The effect of high water temperatures on the allometric  
27 scaling effects of energy and protein starvation losses in juvenile barramundi, *Lates calcarifer*.  
28 *Comp Biochem Physiol. A. Mol. Integr. Physiol.*, 159(2),167-74. doi:  
29 10.1016/j.cbpa.2011.02.013.

- 1 Glencross, B.D., Phuong, N.T., Hien, T.T.T., Cam Tu, T.T., Phu, T.M., 2011. A factorial growth and  
2 feed utilisation model for Tra catfish, *Pangasianodon hypophthalmus*. *Aquaculture Nutrition*  
3 17, e396-e405.
- 4 Glencross B.D. and Bermudes M., 2012. Using a bioenergetic modelling approach to understand the  
5 implications of heat stress on barramundi (*Lates calcarifer*) growth, feed utilisation and optimal  
6 protein and energy requirements - Options for adapting to climate change? *Aquaculture*  
7 *Nutrition* 18(4), 411-422.
- 8 Glencross, B.D., Bermudes, M., 2012. Using a bioenergetic modelling approach to understand the  
9 implications of heat stress on barramundi (*Lates calcarifer*) growth, feed utilisation and optimal  
10 protein and energy requirements – Options for adapting to climate change? *Aquaculture*  
11 *Nutrition* 18, 411-422.
- 12 Liao, I.C., Huang T.S., Tsai W.S., Hsueh C.M. , Chang S.L. and Eduardo M.L., 2004. Cobia culture  
13 in Taiwan: current status and problems. *Aquaculture* 237, 155-165.
- 14 Liu, K., X.J. Wang, Q. Ai, K. Mai, and Zhang W., 2010. Dietary selenium requirement for juvenile  
15 cobia, *Rachycentron canadum* L. *Aquaculture Research* 41, e594-e601.
- 16 Liu, K., Q.H. Ai, K.S. Mai, W.B. Zhang, L. Zhang, Zheng S.X., 2012. Dietary manganese  
17 requirement for juvenile cobia, *Rachycentron canadum* L. *Aquaculture Nutrition* 19:461-467.
- 18 Lupatsch, I., Kissil, G.W., Sklan, D., 2001. Optimization of feeding regimes for European sea bass  
19 *Dicentrarchus labrax*: a factorial approach. *Aquaculture* 202, 289-302.
- 20 Lupatsch, I., Kissil, G.W., Sklan, D., 2003. Comparison of energy and protein efficiency among three  
21 fish species *Sparus aurata*, *Dicentrarchus labrax* and *Epinephelus aeneus*: energy expenditure  
22 for protein and lipid deposition. *Aquaculture* 225, 175-189.
- 23 Mercer, L.P., 1982. The quantitative nutrient-response relationship. *J. Nutr.* 112, 560-566.
- 24 Niu, J., Y.J. Liu, L.X. Tian, K.S. Mai, H.J. Yang, C.X. Ye, Zhu Y., 2008. The effect of different  
25 levels of dietary phospholipid on growth, survival and nutrient composition of early juvenile  
26 cobia (*Rachycentron canadum*). *Aquaculture Nutrition* 14, 249-256.
- 27 Petersen E.H., Glencross B.D., Tien N.V., Tuan L.A., Tuan V.A., Phuong T.H., 2015. Recent  
28 Changes in the Bioeconomic of Finfish Mariculture in Vietnam. *J. Aquac. Res. Development* 6  
29 (3), 311. doi:10.4172/2155-9546.1000311

- 1 Pirozzi, I., Booth, M.A., Allan, G.L., 2010. A factorial approach to deriving dietary specifications and  
2 daily feed intake for mulloway, *Argyrosomus japonicus*, based on the requirements for  
3 digestible protein and energy. *Aquaculture* 302, 235-242.
- 4 Salze, G., McLean, E., Battle, P.R., Schwarz, M.H., Craig, S.R., (2010) Use of soy protein concentrate  
5 and novel ingredients in the total elimination of fish meal and fish oil in diets for juvenile cobia,  
6 *Rachycentron canadum*. *Aquaculture* 298, 294–299.
- 7 Shearer, K.D., 1995. The use of factorial modeling to determine the dietary requirements for essential  
8 elements in fishes. *Aquaculture* 133, 57–72.
- 9 Sun, L., Chen, H., Huang, L., 2006. Effect of temperature on growth and energy budget of juvenile  
10 cobia (*Rachycentron canadum*). *Aquaculture* 261, 872–878.
- 11 Sun, L., Chen, H. 2014. Effects of water temperature and fish size on growth and bioenergetics of  
12 cobia (*Rachycentron canadum*). *Aquaculture* 426–427, 172–180.
- 13 Trung D.V., Diu, N.T., Hao, N.T., Glencross, B.D., 2011. Development of a nutritional model to  
14 define the energy and protein requirements of Tilapia, *Oreochromis niloticus*. *Aquaculture* 320,  
15 69-75.
- 16 Ursin, E., 1967. A mathematical model of some aspects of fish growth, respiration and mortality.  
17 *Journal of the Fisheries Research Board of Canada* 24, 2355-2453.
- 18 Watson, A.M., Holt, G.J., 2010. Energy Budget of Early Juvenile Cobia, *Rachycentron canadum*.  
19 *Journal of the World Aquaculture Society* 41, 224-234.
- 20 Webb, K.A., L.T. Rawlinson, and Holt G.J., 2009. Effects of dietary starches and the protein to  
21 energy ratio on growth and feed efficiency of juvenile cobia, *Rachycentron canadum*.  
22 *Aquaculture Nutrition* 16, 447-456.
- 23 Withers, P.C., 1992. *Comparative Animal Physiology*. Brooks/Cole, Singapore.
- 24 Xiao, L.D., K.S. Mai, Q.H. Ai, W. Xu, X.J. Wang, W.B. Zhang, Z.G. Liufu. 2009. Dietary ascorbic  
25 acid requirement of cobia, *Rachycentron canadum* Linnaeus. *Aquaculture Nutrition* 16, 582-  
26 589.
- 27 Zhou, Q.C., Z.H. Wu, B.P. Tan, S.Y. Chi, Yang Q.H., 2006. Optimal dietary methionine requirement  
28 for Juvenile Cobia (*Rachycentron canadum*). *Aquaculture* 258, 551-557.

1 Zhou, Q.C., Z.-H. Wu, S.Y. Chi, Yang Q.H., 2007. Dietary lysine requirement of juvenile cobia  
2 (*Rachycentron canadum*). Aquaculture 273, 634-640.

3

4

1 Table 1. Reference diet formulation (% as used) and composition (% dry basis unless  
 2 otherwise indicated)

<b>Ingredients</b>	<b>(%)</b>
Brown fish meal	65.5
Wheat flour	14.5
Wheat gluten	10.0
Fish oil	9.4
Mineral and vitamin premix*	0.5
Marker (yttrium oxide)	0.1
<b>Composition</b>	
Dry matter (% as fed)	95.8
Crude protein	48.9
Digestible protein	40.6
Total lipid	13.8
Digestible lipid	13.0
Crude ash	11.0
Carbohydrate**	21.1
Gross energy (kJ/g)	22.2
Digestible Energy (kJ/g)	19.7

4 \* Vitamin and mineral premix includes (IU kg<sup>-1</sup> or g kg<sup>-1</sup> of premix): Vitamin A, 1.3MIU; Vitamin D3, 0.5 MIU; Vitamin E, 0.17  
 5 MIU; Vitamin K, 3, 3.4 g; Vitamin B1, 6.7 g; Vitamin B2, 5.8 g; Vitamin B6, 6.7 g; Vitamin B12, 0.003 g; Folic acid, 0.8 g; D-  
 6 Calpan, 20 g; Niacin, 11.7 g; Biotin, 0.17 g; Vitamin C, 33 g; Inositol, 45 g; Iron, 8.3 g; Zinc, 16.7 g; Copper, 8.3 g; Manganese, 3.0  
 7 g; Cobalt, 0.67 g; Iodine, 0.17 g; Selenium, 0.07 g.

8 \*\* Calculated only

9  
 10

1 Table 2 Calculations of dietary energy and protein requirements for growing cobia at 26°C including feed specifications based on a series (12, 16 and 20  
 2 MJ/kg) suggested dietary DE densities  
 3

Fish live-weight (g/fish)	100	500	1000	2000	100	500	1000	2000	100	500	1000	2000
Growth (g/fish/day) @ 25.6 °C <sup>a</sup>	2.23	5.62	8.37	12.47	2.23	5.62	8.37	12.47	2.23	5.62	8.37	12.47
<i>Energy</i>												
Metabolic BW (kg <sup>0.80</sup> )	0.158	0.574	1.000	1.741	0.158	0.574	1.000	1.741	0.158	0.574	1.000	1.741
DE <sub>maint</sub> (kJ/fish/day) <sup>b</sup>	11.78	42.67	74.30	129.36	11.78	42.67	74.30	129.36	11.78	42.67	74.30	129.36
Energy gain (kJ/fish/day) <sup>c</sup>	13.12	43.58	73.07	122.53	13.12	43.58	73.07	122.53	13.12	43.58	73.07	122.53
DE <sub>growth</sub> (kJ/fish/day) <sup>d</sup>	20.16	66.94	112.25	188.21	20.16	66.94	112.25	188.21	20.16	66.94	112.25	188.21
DE <sub>total</sub> (kJ/fish/day) <sup>e</sup>	31.93	109.62	186.55	317.58	31.93	109.62	186.55	317.58	31.93	109.62	186.55	317.58
<i>Protein</i>												
Metabolic Protein BW (kg <sup>0.80</sup> )	0.200	0.616	1.000	1.625	0.200	0.616	1.000	1.625	0.200	0.616	1.000	1.625
DPro <sub>maint</sub> (g/fish/day) <sup>f</sup>	0.20	0.61	0.99	1.61	0.20	0.61	0.99	1.61	0.20	0.61	0.99	1.61
Protein gain (g/fish/day) <sup>g</sup>	0.37	0.94	1.41	2.09	0.37	0.94	1.41	2.09	0.37	0.94	1.41	2.09
DPro <sub>growth</sub> (g/fish/day) <sup>h</sup>	0.82	2.07	3.08	4.59	0.82	2.07	3.08	4.59	0.82	2.07	3.08	4.59
DPro <sub>total</sub> (g/fish/day) <sup>i</sup>	1.02	2.68	4.07	6.20	1.02	2.68	4.07	6.20	1.02	2.68	4.07	6.20
<i>Feed specifications</i>												
DE content of feed (MJ/kg)	12	12	12	12	16	16	16	16	20	20	20	20
Feed intake (g/fish/day)	2.66	9.13	15.55	26.46	2.00	6.85	11.66	19.85	1.60	5.48	9.33	15.88
Feed intake (%BW)	2.7	1.8	1.6	1.3	2.0	1.4	1.2	1.0	1.6	1.1	0.9	0.8
DPro content of feed (g/kg)	383	294	262	234	511	391	350	312	639	489	437	391
Expected FCR	1.19	1.62	1.86	2.12	0.89	1.22	1.39	1.59	0.72	0.97	1.11	1.27
DPro : DE ratio (g/MJ)	32	24	22	20	32	24	22	20	32	24	22	20

4 <sup>a</sup>Based on Equation 10 and a temperature of 25.6°C. <sup>b</sup>Digestible energy required for maintenance = 74.3 kJ / kg<sup>0.80</sup> /day. <sup>c</sup>Energy content of body based on Equation 9. <sup>d</sup>Amount of digestible  
 5 energy required for growth based on a partial energy utilization efficiency of 0.651. <sup>e</sup>Total digestible energy required per day = DE<sub>maint</sub> + DE<sub>growth</sub>. <sup>f</sup>Digestible protein required for maintenance =  
 6 0.99 g /kg<sup>0.70</sup> /day. <sup>g</sup>Protein content of body based on 16.89% of live-weight. <sup>h</sup>Amount of digestible protein required for growth based on a partial protein utilization efficiency of 0.456. <sup>i</sup>Total  
 7 digestible protein required per day = DP<sub>maint</sub> + DP<sub>growth</sub>

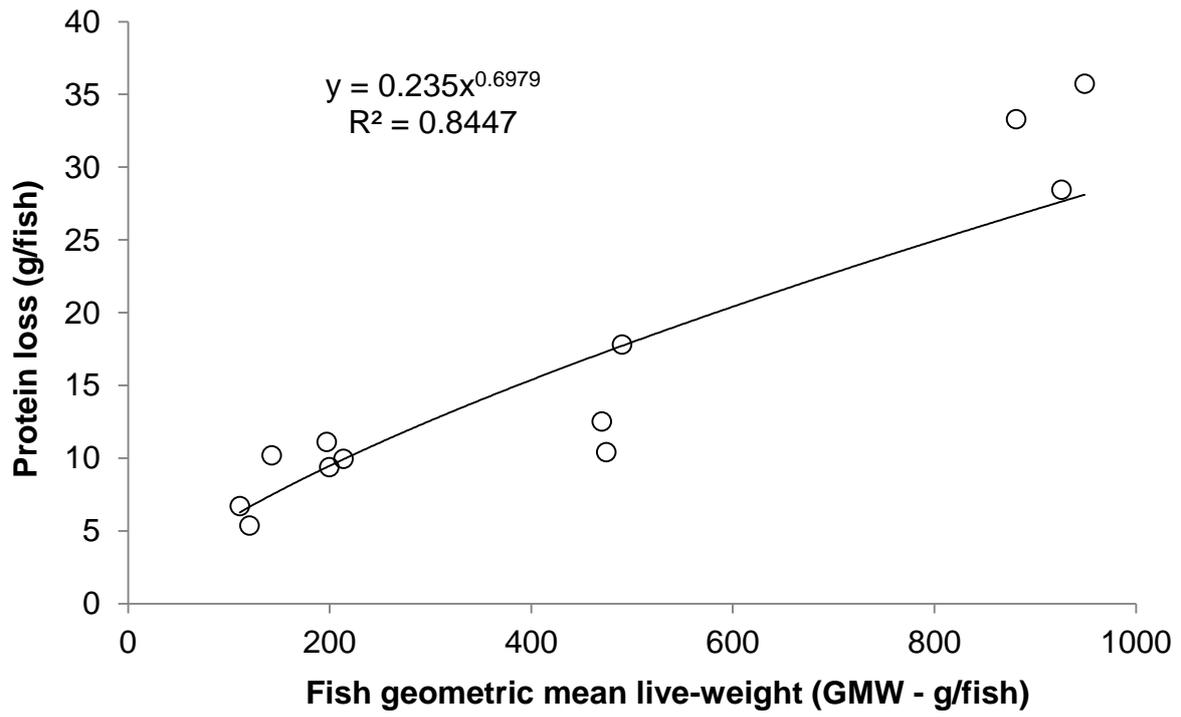


Figure 1. Protein loss (g/fish) by cobia starved for 21 days at 28°C. Regression equation is: Protein loss = 0.235\*(fish geometric mean live-weight)<sup>0.697</sup>, (R<sup>2</sup> = 0.844)

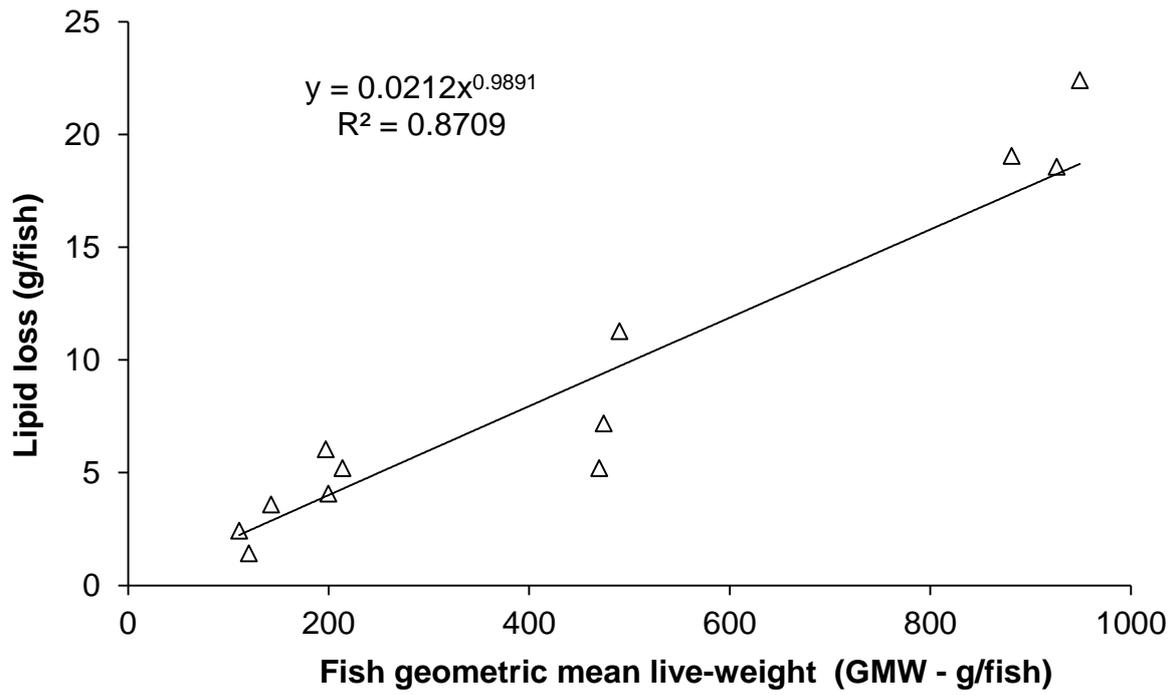


Figure 2. Lipid loss (g/fish) by cobia starved for 21 days at 28°C. Regression equation is: Lipid loss (g/fish) = 0.021\*(fish geometric mean live-weight)<sup>0.989</sup>, (R<sup>2</sup> = 0.870).

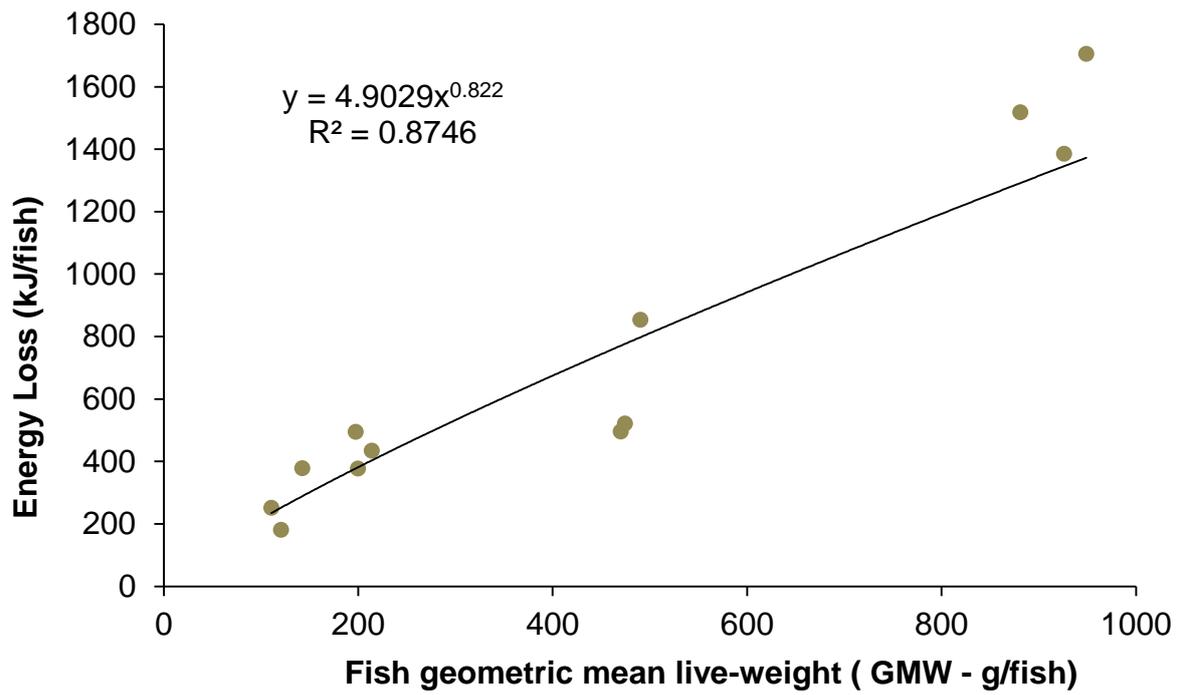


Figure 3. Energy loss (kJ/fish) by cobia starved for 21 days at 28°C. Regression equation is: Energy loss (kJ/fish) = 4.902\*(fish geometric mean live weight)<sup>0.822</sup>, (R<sup>2</sup> = 0.874).

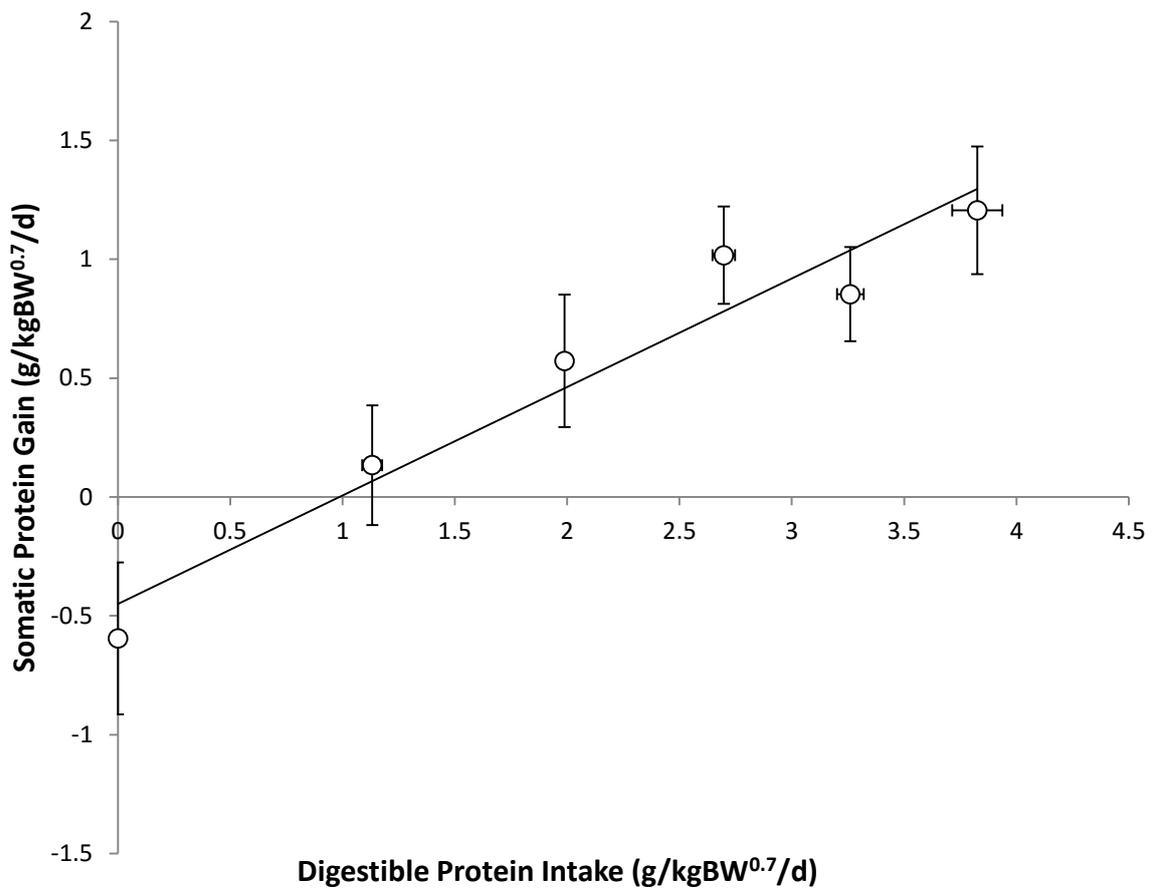


Figure 4 Protein gain (g/kgBW<sup>0.7</sup>/d) by cobia fed increasing amounts of an experimental feed at 27.9 ± 0.3°C. Overall regression equation is: Protein gain = 0.456•(digestible protein intake) – 0.450, (R<sup>2</sup> = 0.938). Maintenance digestible protein intake level is estimated at 0.99 g/kgBW<sup>0.7</sup>/d.

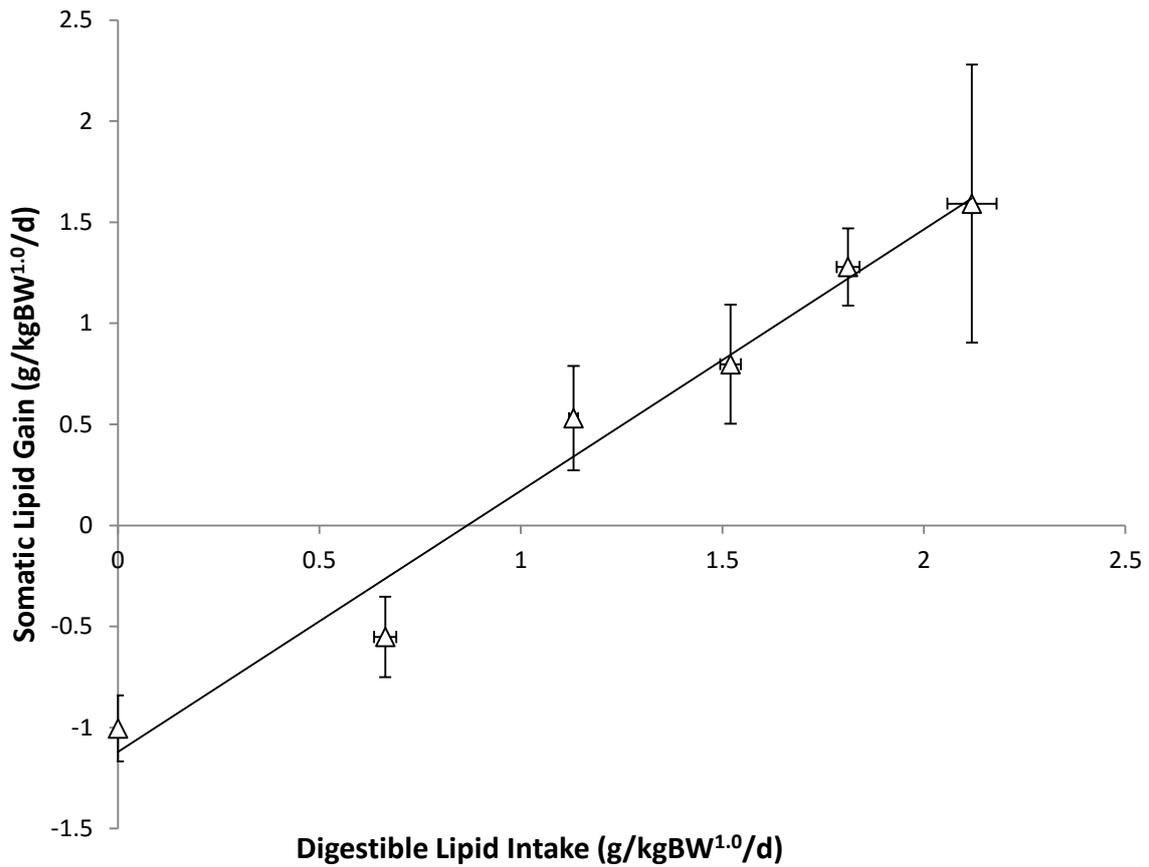


Figure 5 Lipid gain (g/kgBW<sup>1.0</sup>/d) by cobia fed increasing amounts of an experimental feed at 27.9 ± 0.3°C. Overall regression equation is: Lipid gain (g/kgBW<sup>1.0</sup>/d) = 1.292•(digestible lipid intake) – 1.120, (R<sup>2</sup> = 0.973). Maintenance digestible lipid intake level is estimated at 0.87 g/kgBW<sup>1.0</sup>/d

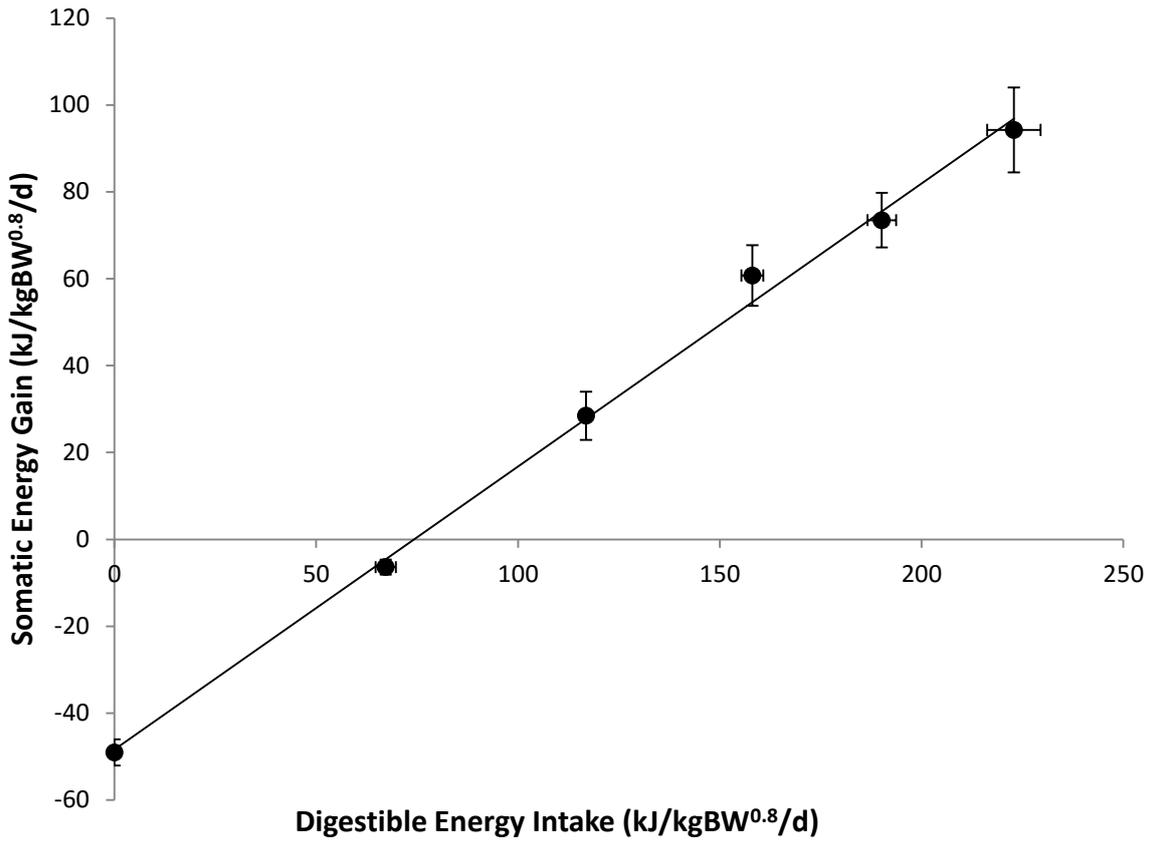


Figure 6 Energy gain (kJ/kgBW<sup>0.8</sup>/d) by cobia fed increasing amounts of an experimental feed at 27.9 ± 0.3°C. Overall regression equation is: Energy gain = 0.651 • (Digestible energy intake) – 48.411, (R<sup>2</sup> = 0.996). Maintenance digestible energy intake level is estimated at 74.3 kJ/kgBW<sup>0.8</sup>/d.

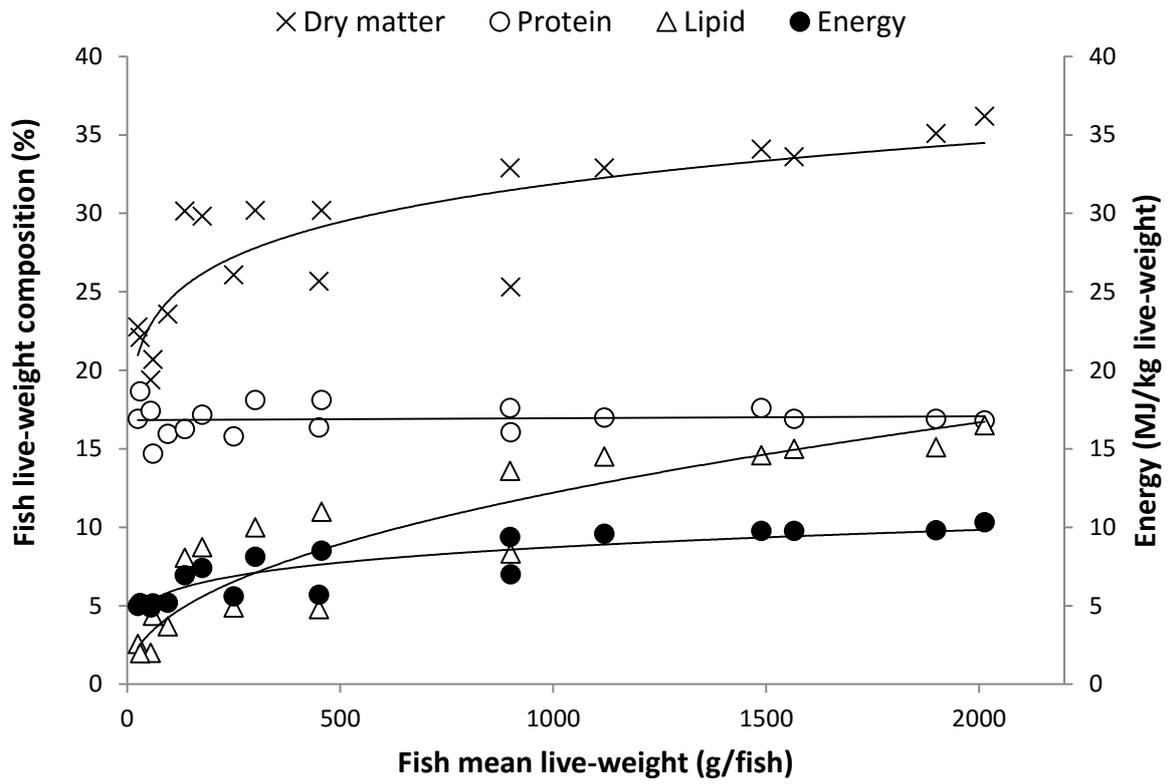


Figure 7. Live-weight body compositions of cobia (n = 18) from 25g to 2013g. Fish were a combination of laboratory and commercial farmed stocks fed either laboratory or commercial diets.

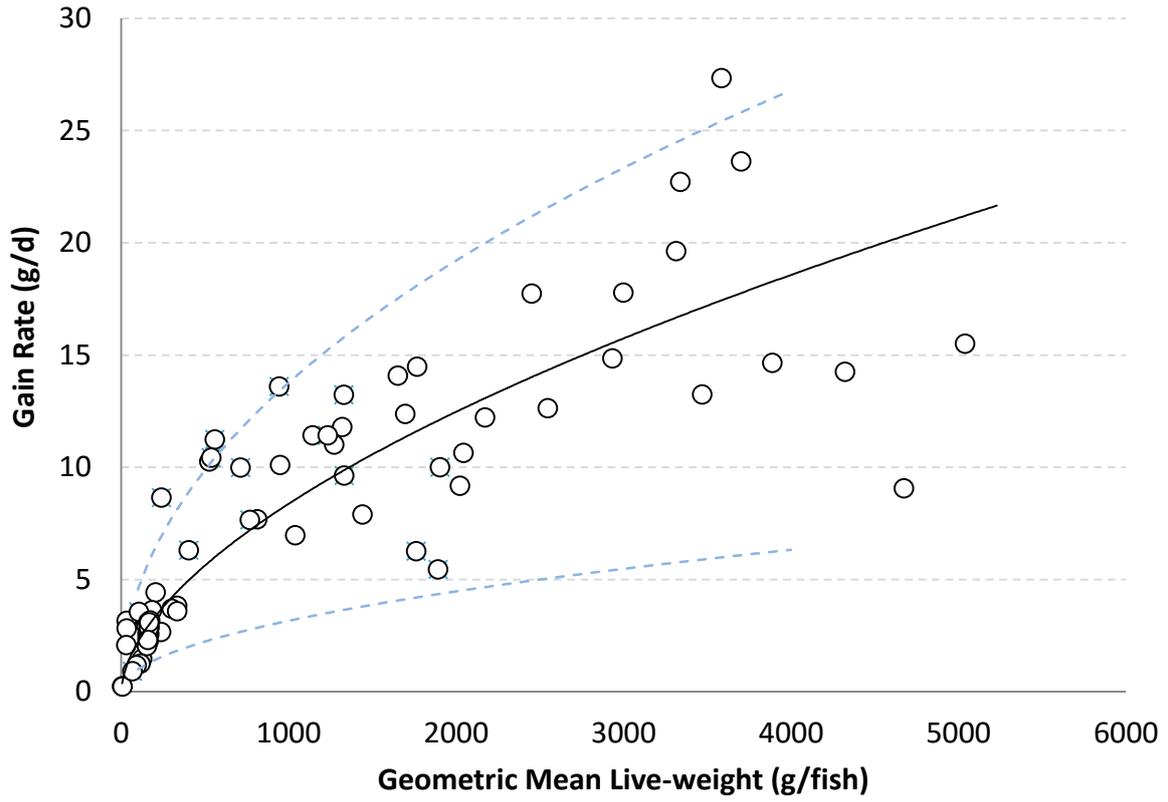


Figure 8 Growth rates of cobia with varying live-weight size range (expressed as geometric mean live-weight). Growth rate (g/d) was defined by the equation  $y = 0.1586x^{0.574}$  ( $R^2$  0.847). Average temperature across all data is  $25.6 \pm 2.8^\circ\text{C}$  (mean  $\pm$  SD).

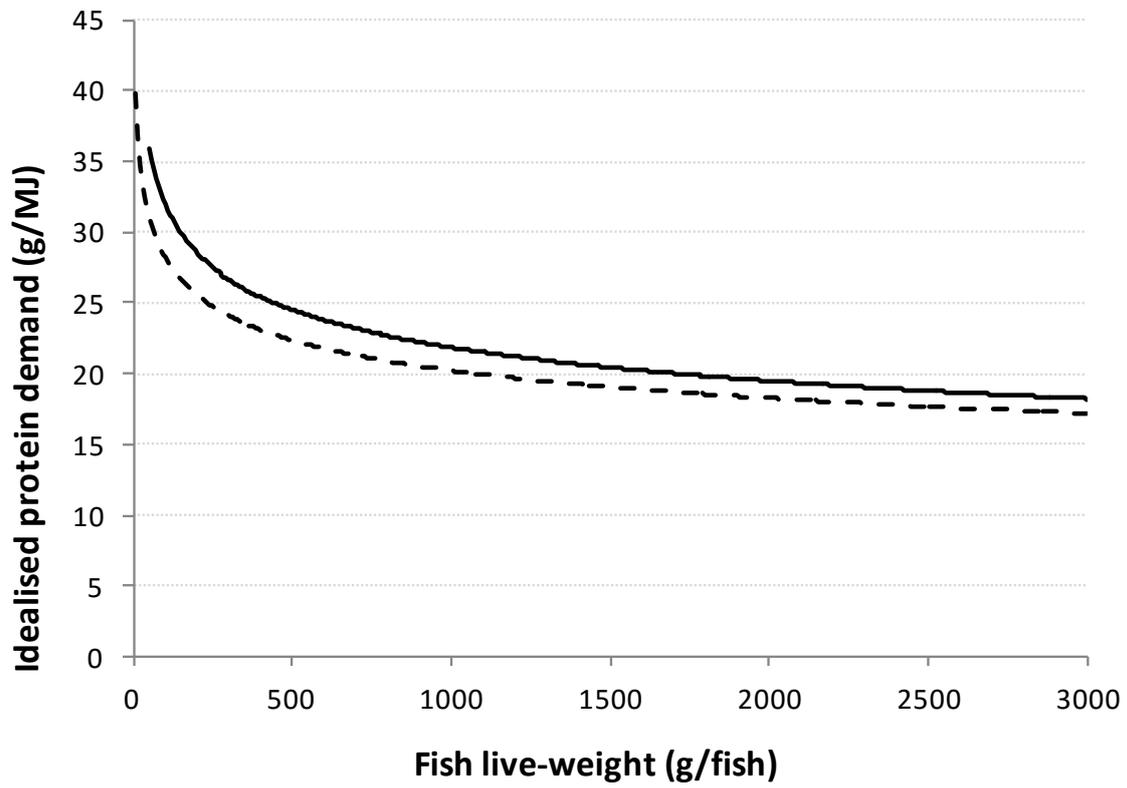


Figure 9 A comparison of the idealised protein demand (g/MJ) of cobia (solid line) with Asian seabass (dashed line) with varying live-weight size range (Asian seabass data derived from Glencross and Bermudes, 2012). Shown is the marginally higher demand for protein by cobia relative to this other tropical carnivorous species.