1 # Hybridisation and hybrid speciation under global change

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Summary

An unintended consequence of global change is an increase in opportunities for hybridisation among previously isolated lineages. Here we illustrate how global change can facilitate the breakdown of reproductive barriers and the formation of hybrids, drawing on the flora of the British Isles for insight. Although global change may ameliorate some of the barriers preventing hybrid establishment, for example by providing new ecological niches for hybrids, it will have limited effects on environment-independent post-zygotic barriers. For example, genic incompatibilities and differences in chromosome numbers and structure within hybrid genomes are unlikely to be affected by global change. We thus speculate that global change will have a larger effect on eroding pre-zygotic barriers (eco-geographic isolation, phenology) than post-zygotic barriers, shifting the relative importance of these two classes of reproductive barriers from what is usually seen in naturally produced hybrids where pre-zygotic barriers are the largest contributors to reproductive isolation. Although the long-term fate of neo-hybrids is still to be determined, the massive impact of global change on the dynamics and distribution of biodiversity generates an unprecedented opportunity to study large numbers of unpredicted, and often replicated, hybridisation "experiments", allowing us to peer into the birth and death of evolutionary lineages.

I. Introduction

'Global change', the term often used to describe the combination of planetary change and human societal change, is having a profound effect on biodiversity across the globe. Climate change, industrialization, environmental degradation, and global trade and travel have moved plants and animals around the planet, breaking down previous geographic barriers to gene flow between closely related species (Mooney & Cleland 2001). The loss of ecological and geographic barriers that historically kept closely related species apart is creating unprecedented new opportunities for hybridisation (Mable 2013; Chunco 2014, Brennan *et al.,* 2015; Taylor *et al.*, 2015), which could potentially have a profound impact on biodiversity and ecosystems worldwide.

Hybridisation has been defined in many different ways, from crosses between genetically distinct populations (Abbott *et al.,* 2013), to crosses between genetically distinct taxa resulting in the production of viable offspring (Mallet 2007), thus occurring both within and between species (Rieseberg & Carney 1998). Here we focus on inter-specific hybridisation. Interspecific hybridisation is a regular natural phenomenon and it is estimated that as many as 25% of plant species and 10% of animal species hybridise naturally (Mallet 2007). Hybridisation has long attracted the interest of evolutionary biologists, in part because it seems to undo the very process responsible for the generation of species diversity (Dobzhansky 1937, Mayr 1942, Stebbins 1959, Grant 1971, Abbott *et al.,* 2013). However, the consequences of hybridisation between genetically distinct lineages can result in a variety of outcomes that may influence diversity, including gene flow from one taxon to another (introgression) (Rieseberg & Wendel, 1993), the displacement and/or extinction of one or both parental taxa (Ellstrand & Elam, 1993), the fusion of previously divergent taxa (Grant & Grant, 2014), or the creation of new, stable hybrid taxa and, ultimately, speciation (Mallet 2007, Abbott *et al.,* 2013, Thomas 2015). Hybridisation and its consequences have been best studied in plants (Grant, 1971; Arnold, 1997; Rieseberg & Carney, 1998; Abbott *et al.*, 2013), but the biological features and evolutionary mechanisms that contribute to the variation in the occurrence, persistence, and evolution of hybrids are relatively little understood in both plants and animals.

Whether the incidence of hybridisation at a global scale is changing, and if so at what rate, is still unclear, yet analyses of well-studied floras, such as the British Isles, indicate that a significant fraction of hybrid taxa may involve introduced taxa (Stace *et al.,* 2015, Stace & Crawley 2015). Among the flora of Britain and Ireland hybridisation involving introduced (non-native) taxa is well documented (Stace *et al.,* 2015, Stace & Crawley 2015, Table 1), and a recent survey revealed that 33% of hybrid taxa (301/909) involve introduced (non-native) taxa (Preston & Pearman 2015). Approximately half of these non-native hybrids have arisen spontaneously in the wild, while the remainder were introduced as hybrids. Studying the origin and fate of these hybrids is timely and

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108 important if we are to understand the consequences of ongoing changes in the distribution of global

109 biodiversity.

110   Because hybrid formation does not equate to hybrid success, the long term consequences of

111 contemporary hybridisation under global change are hard to predict. Here we present an overview

112 of the consequences of this new era of increased hybridisation opportunities drawing on the flora of

113 Great Britain and Ireland because it is probably the best documented hybrid flora in the world.

114 Specifically, our review addresses the following questions: How does global change alter the

115 likelihood of hybrid formation? Does global change alter the relative importance of prezygotic and

116 postzygotic barriers in maintaining species apart? What is the incidence of human-made hybrids in

117 current floras and is likely to increase or decrease under global change? What are the biological

118 characteristics that make some hybrids more likely to become established than others? Why are

119 recently formed hybrids relatively common, but their allopolyploid derivatives rare? We suggest that

120 global change is clearly increasing opportunities for hybridisation, for example directly by moving

121 species around the world or indirectly by eroding phenological and ecological barriers. Human-made

122 hybrids, created for example for horticultural purposes, can also significantly contribute to current

123 floras as exemplified in the naturalised flora of Britain and Ireland. We speculate that this new

124 scenario is shifting the relative importance of prezygotic and postzygotic barriers from what is

125 generally seen in natural systems, and therefore the consequences of secondary contact in this

126 changing world may depend more heavily on postzygotic than on prezygotic barriers. The

127 persistence of newly formed hybrids, and their evolutionary fate, will hinge on mechanisms that

128 stabilise hybrids including asexual reproduction, selection for increased fertility, polyploidy, and

129 ecological niche diversification, among others. In the next sections, we take a comprehensive

130 approach to understand the processes and mechanisms leading to: (1) hybrid formation, (2) the

131 establishment of hybrids, and (3) the ecological and evolutionary outcomes of hybridisation.

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133 II.   Hybrid formation

134 Understanding hybridisation and the potential for global change to alter its incidence requires

135 analysis of the conditions that allow hybrid formation and establishment. Speciation involves the

136 origin of barriers preventing gene flow between incipient species (Coyne & Orr 2004, Baack *et al.,*

137 2015), but hybridisation bypasses these barriers. Isolating barriers are traditionally classified as

138 those acting before (pre-zygotic) and after zygote formation (post-zygotic) (reviewed in Coyne & Orr

139 2004) (Fig. 1). In plants, pre-zygotic barriers include: (i) Pre-pollination barriers such as geographical

140 and ecological barriers (e.g. habitat preferences), flowering phenology and pollinator preference and

141 behaviour, which prevent or reduce the likelihood of interspecific mating; and (ii) post-pollination

142 barriers which act before zygote formation, such as unilateral interspecific incompatibility (Hiscock

143 *et al.*, 1998), and gametophytic selection, in which conspecific pollen is favoured over heterospecific

144 pollen (Rieseberg & Willis 2007). The breakdown of geographic isolation is usually treated separately

145 from other isolating barriers (Coyne & Orr 2004) as it may reflect both biological differences in

146 habitat preference and non-biological and historical features causing species to occur in different

147 geographic locations (Dobzhansky 1937). However, we include geographic isolation in our discussion

148 because global change, including the movement of species around the globe, has had a direct impact

149 in increasing the opportunities for secondary contact between previously isolated taxa.

150 Although the potential for human-induced environmental change to influence the rate of

151 hybridisation has long been recognised (e.g., Anderson 1948), the evidence to support this view is

152 only just beginning to accumulate (Walther *et al.,* 2009, Garroway *et al.,* 2010, Hoffmann & Sgrò

153 2011, Campbell & Wendlandt 2013, Chunco 2014, Chown *et al.,* 2015). In this section, we discuss

154 how hybrids may overcome pre-zygotic isolating barriers and geographic isolation, paying attention

155 to those barriers that are likely to be affected by global change.

1. Breakdown of geographic isolation

157 The breakdown of geographic isolation when species increase their range, either as a consequence

158 of climate change (e.g. during periods of glaciation), or through anthropogenic dispersal, provides

159 historically isolated taxa with new opportunities for hybridisation. Human-mediated transport of

160 species, either accidental or deliberate, is the most dramatic of these and has been going on for

161 thousands of years, but has been accelerating rapidly in Europe and North America (Hulme *et al.,*

162 2008). Increased international trade and travel in the current era of globalisation means that the

163 spread of non-native species is likely to increase (Hulme *et al.,* 2008). The pathways of human-

164 assisted dispersal are varied, and include accidental long-distance dispersal events that result in

165 translocation of terrestrial plant species across oceanic barriers, facilitated dispersal (e.g., along

166 roadsides and railroads), and intentional introductions (e.g., horticultural trade). Although long

167 distance dispersal events occur with low probability, theoretical analyses have shown that these

168 events can accelerate migration rates in plants by an order of magnitude (Higgins & Richardson

169 1999), greatly increasing opportunities for secondary contact.

170 Long distance dispersal where propagules are transported as contaminants of commodities (e.g.,

171 grains, timber, and wool) and in ships' ballasts is well documented (Stace & Crawley 2015). For

172 instance, in the early 20th century, the River Tweed in the English-Scottish border hosted 384

173 introduced plant species growing along its banks (Myers & Bazely 2003). Many of these plants

174 originated in Europe, Asia, Australasia, and the Americas and were brought as wool contaminants,

175 which were then washed into local rivers during the cleaning process (Silvertown 2011). In addition,

176   anthropogenic activities can also facilitate range expansion by creating dispersal routes that can

177   then be exploited by natural means. Roads and railways provide corridors that can act as dispersal

178   routes allowing species to spread rapidly to new areas. The introduced hybrid *Senecio squalidus*

179   (Oxford ragwort, Asteraceae), originally from Mount Etna, Sicily, escaped from cultivation in the

180   Oxford Botanic Garden in the late 18th century, but it was not until the development of railway lines

181   in the 1800's that it began spreading throughout the British Isles (Abbott *et al.,* 2009). The spread of

182   *S. squalidus* in the British Isles has resulted in novel hybridisation events with native *Senecio* species.

183   Crosses between *S. squalidus* and *S. vulgaris* have given rise to three new fertile hybrid taxa via a

184   sterile triploid intermediate ('bridge') *S. x baxteri*: the tetraploid introgressant 'radiate goundsel',

185   *Senecio vulgaris* var. *hibernicus*, the allohexaploid *S. cambrensis* (Welsh groundsel), and the

186   tetraploid *S. eboracensis* (York radiate groundsel, Abbott & Lowe 2004). Additionally, hybridization

187   between *S. squalidus* and native *S. viscosus* has given rise to the sterile triploid *S. subnebrodensis*

188   (Lousley 1946).

189   In addition to facilitating long distance dispersal events, global change in the form of climate

190   change can increase previously existing areas of sympatry among species, or bring previously

191   isolated taxa together through shifts in their range (Hoffmann & Sgrò 2011, Brennan *et al.,* 2015).

192   For example, changes in temperature or precipitation can result in increased range overlap, and the

193   creation or expansion of hybrid zones (Campbell & Wendlandt 2013, Taylor *et al.,* 2015). Because

194   hybrid zones often occur at the range limits of the parental species, changes to the geographic

195   boundaries where these species occur should have a particularly strong impact on the location and

196   extent of hybrid regions (Chunco 2014). There are a number of historical examples of this within the

197   British flora. For instance, the hybrid waterlily *Nuphar x spenneriana* (Nymphaeaceae) is postulated

198   to have arisen during the Late Glacial period ~10,000 years ago when the ranges of its parental

199   species *N. pumila* (tolerant of cold water) and *N. lutea* (preferring warmer water) overlapped

200   (Preston and Croft 1998). *N x spenneriana* then subsequently displaced *N. lutea* at its more northerly

201   locations due to its competitive advantage in tolerating cooler water. A similar story may account for

202   the current distribution of *Circaea x intermedia* (Onagraceae), a hybrid between *C. alpina* and *C.*

203   *lutetiana* which has all but displaced its cold-loving parent *C. alpina* in northern Britain since the ice

204   retreated (Marren 1999).

205   Climate change can also facilitate the spread of invasive populations which can then contribute

206   to hybridisation events either with native species, or with other invasive taxa. For example, warmer

207   climates may allow the migration of species from warmer regions into regions that were formally

208   too cold for their long-term survival as well as allowing introduced species from warmer regions to

209   overwinter more successfully and extend the growing season, thus facilitating the establishment and

210 spread of non-native taxa (Walther *et al.,* 2009).The increases in populations of *Orchis simia* and *O.*

211 *purpurea* in Kent and their spread north into other counties of southern England provides a good

212 example of the former scenario (Fay 2015). Despite the expected importance of climate change on

213 increasing hybridisation rates (Chunco 2014, Brennan *et al.,* 2015), most evidence in plants remains

214 indirect (e.g., perceived changes on the extent of hybrid zones) or correlative (e.g., comparisons of

215 number of hybrids at different time points), and more detailed case studies are needed (e.g.,

216 Campbell & Wendlandt 2013).

2. Erosion of ecological isolation barriers

218 The production of hybrids may be prevented by pre-zygotic, ecological barriers that have evolved as

219 by-products of adaptation to their local environments (Coyne & Orr 2004, Baack *et al.,* 2015) (Fig. 1).

220 For example, broadly sympatric species may still display ecological preferences for different habitats

221 (e.g., mesic vs. arid habitats and forested vs. open habitats), or be reproductively isolated by virtue

222 of flowering at different times. Habitat modification, including increased disturbance and

223 fragmentation, can erode ecological barriers and facilitate hybrid formation (Anderson 1948,

224 Stebbins 1950, Buggs 2007). Moreover, altered and disturbed habitats, e.g., arising as a consequence

225 of agricultural practices or urbanisation, can provide hybrids with new environments where they can

226 establish. Among the British flora *Silene dioica x S. latifolia (= S. x hampeana,* Caryophyllaceae)

227 (Marren 1999) provides a good example of how habitat modification may facilitate hybridisation,

228 while *Senecio squalidus* and *S. cambrensis* (Asteraceae) are examples of how hybrids can establish in

229 new environments (Abbott *et al.*, 2009). Human-induced breakdown of ecological isolation has also

230 been implied in hybridisation between *Centaurea erythraea* and *C. littorale* (Gentianaceae). These

231 two species generally occur in different habitats, but human disturbance in sand dune systems is

232 thought to have resulted in mixed populations (Ubsdell 1979), which can be found in coastal regions

233 in England and mainland Europe (Brys *et al.,* 2014). Some of these populations harbour nearly sterile

234 F1 hybrids and backcrosses, which appear to differ in fertility (Ubsdell 1979). Nevertheless,

235 differences in floral morphology and mating system are efficient in reducing hybridisation, at least in

236 mixed populations in Belgium, where established hybrids occur at a frequency of approximately 1%

237 (Brys *et al.,* 2014). Ultimately, the outcome of habitat alteration on hybridisation may be dependent

238 on phylogenetic relationships between taxa. For instance, Brennan *et al.*, (2015) suggest that habitat

239 alteration may be more important for hybridisation between younger taxa, as they are expected not

240 yet to have accumulated strong genetic barriers.

241   Climate change affects the phenology of many species (Cleland *et al.,* 2007), for instance by

242 causing plants to flower earlier. Changes in the timing of reproductive events, such as flowering, can

243 directly alter the level of reproductive synchronisation between sympatric species, breaking down

244 temporal isolation barriers, and facilitating hybridisation. For temperature or precipitation changes

245 to increase hybridisation opportunities, it is necessary for one species to change its phenology while

246 the other remains stable or changes more slowly (Chunco 2014). Experimental data shows that this

247 situation may not be uncommon. For instance, artificial warming of experimental plots results in

248 more reproductive overlap among grassland species (Sherry *et al.,* 2007). The breakdown of

249 temporal isolation may be particularly important in sympatric species where phenological isolation is

250 an important reproductive barrier.

251   A further potential barrier to hybridisation in sympatric populations is isolation resulting from

252 pollinator preference (Ramsey *et al.,* 2003). If global change affects the distribution of pollinators

253 and/or their behaviour, we can predict that hybridisation could be facilitated by a breakdown of

254 pollinator isolation barriers (Campbell & Wendlandt 2013). Recent studies show that the distribution

255 of certain species of butterfly in the British Isles and Europe is changing and that their choices of

256 host (brood) plants are changing as a consequence (Bridle *et al.,* 2013). If these changes mean that

257 pollinators visit a broader range of species, it is conceivable that this creates new opportunities for

258 hybridization. Paradoxically, changes in visitation preferences could hypothetically reduce

259 hybridisation, for instance if generalist pollinators change their preference to ignore rarer plant

260 species, thereby resulting in a reduction in heterospecific pollen transfer. In addition to changes in

261 visitation preferences, loss of pollinator diversity may also alter opportunities for hybridization. For

262 example, if pollinators that are specialised on individual plant species are lost, pollen transferred by

263 generalist pollinators may deposit a proportionally higher fraction of heterospecific pollen on

264 stigmas. Clearly, the consequences of changes in pollinator distribution, diversity and abundance can

265 have complex repercussions on hybridization, and these may depend on the particular assemblage

266 of plant and pollinator species. Future studies, akin to that of Bridle *et al.*, should seek to address

267 these questions.

3. Artificial hybridisation

269 Traditionally, artificial hybridisation is not considered in discussions of the ecological and

270 evolutionary significance of hybrids (Rieseberg 1995, Arnold 1997). However, hybrids can be

271 produced artificially and then become part of the natural environment. In the context of global

272 changes to species distribution, the importance of artificial hybrids may be substantial, at least at

273 ecological levels. For example, in the flora of the British Isles, approximately 17% (152/909) of

274 hybrids were introduced as hybrids (Preston & Pearman 2015). Moreover, some of these are

275 deliberate introductions of hybrids of agricultural or horticultural interest such as *Mentha*

276 (Lamiaceae), *Mimulus* (Phrymaceae), and *Verbascum* (Scrophulariaceae). Although most

277 horticultural varieties do not persist outside cultivation (Mack 2005), horticultural introductions do

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278   contribute to the non-native component of many floras, including the British Isles (Clement & Foster

279   1994, Stace 2010).

280   Artificial hybrids may have a disproportionate contribution to hybrid floras, as artificial

281   hybridisation can bypass other pre-zygotic barriers such as pollinator isolation, phenology, habitat

282   isolation and geographic isolation. Furthermore, artificial selection on horticultural hybrids may

283   coincidentally increase their probability of establishment and spread (Ellstrand & Schierenbeck

284   2000). For example, horticultural varieties are often selected on the basis of traits such as hardiness

285   and cold-tolerance (Milne & Abbott 2000), or ease of propagation and large plant size, which may

286   allow the establishment of cultivars beyond the ecological range of their wild parental species (Mack

287   2005). Indeed, it would be interesting to investigate whether artificial hybrids deliberately

288   introduced through the horticulture trade have a larger range than those that originated

289   spontaneously. Artificial crosses from both the horticultural trade and botanic gardens are a

290   relatively unappreciated source of plant hybridisation (Knobloch 1972, Ellstrand & Schierenbeck

291   2000), but the contribution of artificial hybrids to ecological and evolutionary phenomena is likely to

292   increase in an era of global change.

293   ## 4. Weakening of gametic barriers

294   In some cases, pre-zygotic, post-pollination 'gametic' barriers must also be overcome for hybrid

295   formation. This type of post-pollination barrier can arise from gametophytic incompatibilities and

296   conspecific pollen precedence (Hiscock *et al.,* 1998, Howard 1999, Husband *et al.,* 2002).

297   Gametic/gametophytic barriers should be little affected by processes associated with global change,

298   unless these affect the relative receipt of heterospecific pollen (e.g., by increasing the number of

299   heterospecific matings). Because conspecific pollen precedence often depends on the relative

300   amounts of conspecific and heterospecific pollen received on stigmas (Howard 1999), an increase in

301   heterospecific pollen receipt may translate to higher rates of hybrid production. This could be the

302   case if an introduced species becomes invasive and comes to dominate a particular habitat where

303   related native species occur (Morales & Traveset 2009). Pollinators would then carry an increased

304   pollen load of the alien species with corresponding increased chance of cross-pollinating the native

305   species. Thus we predict that the main consequence of global change on pre-zygotic, post-

306   pollination barriers will be through increasing the deposition of heterospecific pollen as non-native

307   species become more abundant, thereby weakening conspecific pollen precedence and facilitating

308   hybridization.

309   In summary, global change is clearly altering the opportunities for hybrid formation, and we

310   would expect the incidence of hybridization to continue increasing. Increased opportunities for

311   hybrid formation may result as a consequence of multiple and not mutually exclusive processes,

312 such as the global re-shuffling of species through human-mediated dispersal, shifting species

313 distributions and phenology as a consequence of climate change, the erosion of ecological barriers,

314 including the "hybridisation of the habitat", changes in pollinator preference and diversity, the

315 expansion of non-native species, and the artificial production and spread of hybrids. Although we

316 think that hybrid formation is likely to increase, an alternative view may be that we have reached

317 "peak" hybridisation, and the rate of hybridisation is slowing down. For example, it may be argued

318 that both species introductions and invasions reduce local biodiversity, and fewer species should

319 mean fewer hybridisation opportunities. Similarly, loss of habitat may reduce contact zones between

320 potentially hybridising taxa. However, we think that the effect of global change in promoting

321 hybridisation through the mechanisms reviewed in this section will by far exceed any hypothetical

322 reduction in hybridisation opportunities. As our records of local floras continues improving,

323 monitoring the temporal patterns of hybridization in floras around the world becomes feasible. Of

324 particular interest will be to monitor floras in developing countries where expansion of international

325 trade, and colossal changes in land use as economies grow, may fast-track the upward trend in the

326 formation of hybrids.

III.   Hybrid establishment: Overcoming intrinsic and extrinsic post-zygotic barriers

329 After hybrids are formed, their short-term fate is partly determined by their ability to overcome both

330 intrinsic (e.g., environment-independent low viability and sterility), and extrinsic (e.g., ecological

331 selection) post-zygotic isolation barriers (Coyne & Orr 2004) (Fig. 1). In natural settings, pre-zygotic

332 barriers may contribute more to total reproductive isolation than post-zygotic barriers (Lowry *et al.,*

333 2008, Baack *et al.,* 2015). However, given the potential for global change to by-pass pre-zygotic

334 barriers and geographic isolation, as argued in the previous section, we speculate that post-zygotic

335 isolation may be the most important hurdle to overcome in the early stages of neo-hybridisation.

336 Intrinsic post-zygotic isolation barriers have been relatively well characterised at the genetic level, in

337 comparison to pre-zygotic barriers (Lowry *et al.,* 2008, Widmer *et al.,* 2009, Rieseberg & Blackman

338 2010). As in animal systems, post-zygotic isolation barriers are expected to increase as a function of

339 genetic divergence (Coyne & Orr 2004), although evidence of this from plant systems is limited

340 (Moyle *et al.,* 2004, Scopece *et al.,* 2007).

5. Escaping hybrid inviability and sterility

342 Hybrids are not uniformly unfit (Arnold & Hodges 1995, Rieseberg & Carney 1998), and within a

343 single hybrid taxon (or sometimes a single hybrid cross) it is possible to find individuals with lower,

344 similar or higher fitness relative to their parents (Taylor *et al.,* 2009). Nevertheless, many hybrids

345 have to overcome severe intrinsic post-zygotic barriers before they can become established (Lowry

346 *et al.,* 2008).

347   Post-zygotic barriers include intrinsic hybrid inviability and sterility (Fig. 1), and may be

348 caused by a variety of mechanisms (Rieseberg & Blackman 2010, Baack *et al.,* 2015), including

349 differences in chromosome structure and ploidy level (Stebbins 1971, Rieseberg 2001, Levin 2002),

350 nuclear-nuclear and nuclear-cytoplasmic genic incompatibilities (Lowry *et al.,* 2008), and parental

351 genomic conflict during seed development (Köhler *et al.,* 2010) (Fig. 1). Genic incompatibilities

352 associated with reduced hybrid fertility and viability have been extensively studied at the genetic

353 level in many plant hybrids (Lowry *et al.,* 2008). The most widely accepted model for the

354 accumulation of such hybrid incompatibilities is the Dobzhansky-Muller (DM) model, in which

355 allopatric populations fix different (and mutually incompatible) alleles at one or more loci (Rieseberg

356 & Willis 2007, Baack *et al.,* 2015). An example of a post-zygotic hybridisation barrier that arises

357 through genic incompatibilities is hybrid necrosis, which resembles plant responses to stress

358 including pathogens (Bomblies & Weigel 2007). Hybrid necrosis can be debilitating or lethal, and

359 generally acts in early (F_1 and sometimes F_2) hybrid generations (Bomblies & Weigel 2007). Other

360 examples of genic-based, intrinsic post-zygotic barriers in inter-specific crosses include hybrid

361 inviability in *Iris* (Iridaceae) (Martin *et al.,* 2008), and hybrid sterility in *Mimulus* (Phrymaceae)

362 (Fishman & Willis 2001) and *Solanum* (Solanaceae) (Moyle & Graham 2005). Postzygotic barriers can

363 thus affect hybrids at different stages (e.g., immediately after zygote formation or in later hybrid

364 generations) and with different consequences (e.g., killing or debilitating hybrids, or rendering them

365 partially or completely sterile).

366   Among the strongest postzygotic barriers are those due to differences in the structure and

367 number of chromosomes of hybridising species (Levin 2002, Husband 2004, Karlsdóttir *et al.,* 2008,

368 Chapman & Abbott 2010). Hybrid viability can be severely affected by differences in the number of

369 chromosomes between hybridising species, especially in interploidy crosses (Stebbins 1958).

370 Hybridisation between a diploid and a tetraploid is often associated with endosperm failure, a

371 phenomenon known as triploid block (Bretagnolle & Thompson 1995, Köhler *et al.,* 2010). However,

372 triploid block may represent a porous hybridisation barrier (Scott *et al.,* 2013). For example, hybrid

373 inviability associated with triploid block is often asymmetric, meaning that the viability of the hybrid

374 depends on whether a given taxon is the maternal or paternal parent (Ramsey & Schemske 1998). In

375 the British Isles a good example of this asymmetry is *Nasturtium x sterile* (2n = 48 (45-58),

376 Brassicaceae), where attempts to produce this well-established hybrid through artificial crosses are

377 successful only if *N. microphyllum* (2n =64) is used as the maternal parent and *N. officinale* (2n = 32)

378 as the paternal (Howard & Manton 1946, Stace *et al.,* 2015). The British flora has many examples of

379 young, established hybrids produced from parents of different ploidy, including a number of species

380 of recent origin, in which at least one of the parents is a non-native taxon such as *Anchusa*

381 *ochroleuca x A. officinalis* (Boraginaceae)*, Brassica napus x B. rapa* (Brassicaceae)*, Fallopia*

382 *baldschuanica x F. japonica* (Polygonaceae)*, Gaultheria mucronata x G. shallon* (Ericaceae)*, Mimulus*

383 *guttatus x M. luteus* (Phrymaceae)*, Rorippa austriaca x R. sylvestris* (Brassicaceae)*, Rumex crispus x*

384 *R. frutescens/R. obovatus, Rumex cristatus x R. palustris* (Polygonaceae)*, Senecio squalidus x S.*

385 *vulgaris* (Asteraceae)*, Spartina alterniflora x S. maritima* (Poaceae)*,* and a number of *Verbascum*

386 (Scrophulariaceae) hybrids (Stace *et al.,* 2015). These taxa provide excellent opportunities to further

387 investigate how inter-ploidy hybrids overcome viability barriers such as triploid block, and to what

388 extent natural hybrid populations have an asymmetric origin.

389   Once a viable hybrid has been produced, a subsequent major challenge is to overcome

390 partial or complete sexual sterility. Interploidy hybrids are often sterile (or have strongly reduced

391 fertility), as a consequence of problems in chromosome pairing during meiosis, leading to

392 unbalanced aneuploid gametes, which are often non-functional (Ramsey & Schemske 1998, Comai

393 2005)(Fig. 2). This pairing problem is expected to be most conspicuous in triploids and hybrids with

394 an odd-number set of chromosomes (Griffiths *et al.,* 2000, Comai 2005), but may also arise in other

395 hybrids in which meiotic pairing results in univalents or odd-numbered multivalents (De Storme &

396 Mason 2014). However, occasionally, interploidy hybrids, such as triploids, produce viable gametes

397 (Ramsey & Schemske 1998, Husband 2004, De Storme & Mason 2014) (Fig. 2). Viable gametes could

398 be produced through multiple routes (Fig. 2), including the production of gametes with the somatic

399 number of chromosomes (unreduced gametes; Köhler *et al.,* 2010, De Storme & Mason 2014, Mason

400 & Pires 2015). Although unreduced gametes are produced at a low rate in non-hybrids (0.0056),

401 their rate of production in hybrids is 50 times higher (0.275) (Ramsey & Schemske 1998). This

402 difference in the rate of production of unreduced gametes is consistent with the observation that

403 polyploids are more common in interspecific hybrid crosses than in crosses within species (Ramsey &

404 Ramsey 2014). Incidentally, the production of unreduced gametes may also help hybrids to bypass

405 the triploid block. For example, mating between the unreduced gametes of a diploid and a tetraploid

406 would produce a hexaploid zygote, without the need to go through a triploid stage. Interestingly, it

407 has been suggested that environmental stress, such as extremes of temperature, may increase the

408 rate at which unreduced, and potentially viable, gametes are formed (De Storme & Mason 2014).

409 Together, this leads to the tantalising idea that global change could facilitate hybridization between

410 diploids and tetraploids—with or without triploids stages—that in the absence of extreme weather

411 events would have remained reproductively isolated.

412   Even in species with the same number of chromosomes, hybrid fertility can be negatively
413   affected by structural differences between parental species including chromosomal re-arrangements
414   (e.g., fusions, fissions, deletions, insertions and inversions) (Rieseberg 2001), and differences in the
415   size of homologous chromosomes (Levin 2002). Reduced fertility among species with structural
416   chromosomal differences is often caused by pairing irregularities during meiosis, which result in
417   unbalanced, and unviable, gametes (Rieseberg 2001, Levin 2002). The strength of this type of
418   chromosomal barrier thus depends on the level of differentiation between the hybridising genomes,
419   and the extent to which these differences cause pairing irregularities (Levin 2002). Bivalent pairing
420   during meiosis may partly reflect structural similarity of the hybridising genomes, although recent
421   work in both auto- and allopolyploid systems suggests that it may also be under the control of a few
422   genes (Hollister 2015). Chromosomal re-arrangements have been shown to contribute to hybrid
423   sterility in many plant groups (Rieseberg & Carney 1998), and they may be particularly important in
424   mediating secondary contact between previously allopatric species (Rieseberg 2001). However,
425   artificial selection on fertility of initially near-sterile hybrids has shown that increased chromosome
426   pairing and more stable meiosis can evolve very rapidly and drastically improve hybrid fertility
427   (Grant 1966). Indeed, some hybrid derivatives between species with divergent chromosome
428   structure have overcome initial reductions in fertility, and formed evolutionarily stable lineages
429   (Rieseberg *et al.,* 1996).

430   ## 6. Persistence of viable but sexually sterile hybrids

431   Hybrids may prevail even when the initial F1 is highly sterile, if they are able to propagate by other
432   means (clonally, apomictically, Fig. 3; Table 1) or by evolving higher sexual fertility (Grant 1966). In
433   addition, polyploid derivatives produced via unreduced gametes (Fig. 2) or somatic mutation can
434   have increased fertility (Rieseberg 2001, Stathos & Fishman 2014); the evolutionary fate of these
435   neo-polyploids is discussed in the *Allopolyploid speciation* section.

436   When sexual reproduction is severely impaired, asexual reproduction can allow individuals
437   to persist and spread (Grant 1971). In the British Isles, perennial hybrids that can propagate through
438   clonal reproduction are more widespread than those that cannot (Preston & Pearman 2015; Table
439   2), showing that the capacity for vegetative reproduction is correlated with spatial coverage.
440   Furthermore, analysis of introduced taxa in the recently published alien flora of the British Isles
441   (Stace & Crawley 2015) indicates that approximately 20% of introduced hybrid taxa reproduce
442   exclusively via clonal propagation, compared to only 8% of non-hybrid introduced taxa (Table 2).
443   Examples of sterile hybrid lineages that persist through vegetative propagation include *Circaea x*
444   *intermedia* (Onagraceae)*, Drosera x obovata* (Droseraceae) and *Stachys x ambigua* (Lamiaceae)
445   (Stace *et al.*, 2015). Some of these sterile lineages, for instance the triploid hybrid *Spartina x*

446 *townsendii* have not spread widely (Strong & Ayres 2013). In contrast other clonal taxa such as the

447 sterile triploid hybrid *Mimulus x robertsii* have spread more extensively (Preston *et al.,* 2002, Vallejo-

448 Marín & Lye 2013; Box 2). The ability to disperse over larger geographic areas through vegetative

449 propagation alone, may depend on the characteristics of the habitat. For instance, hybrids growing

450 along dynamic habitats (e.g., rivers and streams) may disperse clonal propagules more extensively

451 than those where dispersal relies on clonal growth on solid ground. For hybrids exploiting dynamic

452 riparian habitats such as those occupied by introduced *Mimulus* populations in the British Isles,

453 global change may facilitate their spread. For example, if extreme weather events result in more

454 intense or frequent flooding, changes in water flow regimes could help spread clonal propagules

455 further. Thus clonal propagation provides a (temporary) escape route to sexual sterility in hybrids,

456 and extreme weather events brought by global change may amplify its effects on the spatial spread

457 of some hybrids.

458   Another way in which asexuality can bypass sexual failure is through the evolution of

459 agamospermy, i.e., the production of seeds without sex (Fig. 3). Agamospermy (often referred to as

460 apomixis, Whitton *et al.,* 2008) is frequently associated with polyploidy and to a lesser extent

461 hybridisation (Grant 1971, Briggs & Walters 1997, Otto & Whitton 2000). Seeds produced through

462 apomixis are genetically identical to the parental plant, and because there is no sexual reproduction

463 they are reproductively isolated from progenitor taxa, so act as 'good' biological species. For this

464 reason, collections of similar apomictic lineages are sometimes called microspecies, agamospecies,

465 agamocomplexes or syngameons (Briggs & Walters 1997). The flora of the British Isles includes

466 numerous examples of cryptohybrids (ancient hybrids stabilised by apomixis) including syngameons

467 in the genera *Euphrasia* (Orobanchaceae)*, Sorbus* (Rosaceae; see Box 1)*, Rubus* (Rosaceae)*,

468 Hieracium* (Asteraceae)*, Taraxacum* (Asteraceae) and *Rosa* (Rosaceae), all of which are relatively

469 understudied. However, apomixis does not necessarily result in the complete loss of sex. Many

470 facultative apomictic species, such as *Sorbus* (Box 1), display sexual 'leakiness', and produce viable

471 pollen and ovules that can contribute to subsequent hybridisation events (Ludwig *et al.*, 2013). A

472 further example of variation in reproductive system in apomicts is *Hypericum perforatum*

473 (Hypericaceae). This species is a facultative agamospermous tetraploid (2n = 32), where sexual

474 reproduction results in diploid offspring, and agamospermy results in tetraploid and hexaploid plants

475 (Barcaccia *et al.*, 2006). *H. perforatum* hybridises with other taxa, including diploid and tetraploid *H.

476 maculatum* (2n = 16, 32) with which it produces 3x, 4x, and 5x hybrids, which show variable levels of

477 pollen and seed fertility. Yet, it is not known whether these hybrids reproduce sexually,

478 apomictically, or using a combination of both. An unusual form of reproduction via seeds that has

479 evolved in hybrids with odd-numbered chromosomes, occurs in species such as *Rosa canina* (2n = 5x

480   = 35). The *R. canina* complex contains a large number of species in which a permanent state of odd-

481   ploidy is maintained by fusion of gametes with complementary genomes (Grant 1971). Permanent

482   chromosomal heterozygosity results from fertilisation and syngamy between pollen with 7

483   chromosomes and ovules with 28 chromosomes (Grant 1971). Determining the incidence and

484   genetic consequences of occasional bouts of sexuality and recombination in hybrids with facultative

485   apomixis and other unusual forms of reproduction is of key importance to understand their fate, as

486   lack of sex can have both potential benefits and costs for the maintenance of hybrids at ecological

487   and evolutionary timescales.

488   One of the potential benefits that arises from bypassing regular sexual reproduction is that the

489   genetic constitution of the hybrids can be preserved ('fixed heterozygosity'), since gene

490   combinations are not broken down by meiosis and recombination. Viable F1 hybrids have the

491   capacity to show increased vigour (heterosis) relative to their parents (Barton 2001), and this hybrid

492   vigour can be maintained in the absence of sex and recombination. Therefore, clonality and apomixis

493   may be important not only for the persistence of hybrids, but also as a mechanism to maintain

494   hybrid vigour. In the context of global change, the advantages of asexual reproduction, including

495   fixed heterozygosity, may be balanced by a limited ability to deal with rapidly changing

496   environments due to low genotypic diversity. Yet, in recently formed asexual hybrid populations,

497   genotypic diversity may still occur as remnants of the initial hybridisation event (e.g., in hybrids

498   formed from diverse parental stock) or as a consequence of multiple origins of the same hybrid

499   (*Mimulus x robertsii*; Vallejo-Marin and Lye 2013), which may provide enough raw material for short-

500   term evolutionary change through genotypic selection. Moreover, many highly asexual populations

501   can preserve significant levels of genetic and genotypic diversity (Vallejo-Marín *et al.*, 2010), as even

502   rare bouts of sexual reproduction can significantly increase genetic variation (Bengtsson 2003).

503   Populations of asexual hybrids may also be able to deal with rapid environmental change through

504   phenotypic plasticity (Nicotra *et al.*, 2010). Additionally, epigenetic variation can also contribute to

505   rapid adaptation to environmental challenges, as suggested by the ecologically differentiated

506   populations of the invasive species *Fallopia japonica* which display abundant epigenetic diversity,

507   despite genetic uniformity (Kilvitis *et al.*, 2014). The variety of mechanisms available to asexual

508   hybrid populations to adapt to different environments may make hybrids more resilient to the rapid

509   modifications brought by global change.

510   ## 7. Ecological inviability

511   Hybrids may suffer from low fitness when they are unable to find a suitable ecological niche, even if

512   they are not affected by intrinsic developmental problems (Coyne & Orr 2004). For instance, hybrids

513   which have intermediate phenotypes might be selected against in parental habitats, resulting in

514   ecological hybrid inviability (Baack *et al.,* 2015). A potential example of such an extrinsic barrier to

515   hybridisation in the British Isles flora is the hybrid between *Geum urbanum* and *G. Rivale,* (*G. x*

516   *intermedium* (Rosaceae). Although the parents of *G. x intermedium* have broadly overlapping

517   distributions they occupy slightly different habitats: *G. urbanum* occurs in well-drained soils, in areas

518   partially shaded or sometimes in open disturbed habitats, while *G. rivale* is usually found in wetter

519   soils and more open habitats (Stace *et al.*, 2015). Experimental crosses have shown that F1 hybrids

520   are viable and fertile, and that seeds produced in zones of contact contain a variety of hybrid

521   genotypes (Ruhsam *et al.,* 2013). Yet, among adult plants only F1s and backcrosses to *G. rivale* are

522   observed (Ruhsam *et al.,* 2011). Using an experimental garden, Ruhsam *et al.* (2013) showed that

523   hybrids have no intrinsic low fitness, raising the possibility that ecological selection may be filtering

524   out certain hybrid classes (Ruhsam *et al.,* 2013). The agent of selection is unknown, but it is possible

525   that the intermediate morphology of hybrids makes them poorly suited for establishing in parental

526   environments, and explain why some advanced generation hybrid are rare despite obvious intrinsic

527   post-zygotic barriers. The role that ecological barriers play in preventing the establishment of

528   recently formed hybrids remains a relatively unexplored.

IV. Outcomes of hybridisation

530   The long term outcomes of hybridisation can vary from genetic homogenisation of the parental taxa

531   (genetic swamping; Rieseberg & Ellstrand 1993, Wolf *et al.,* 2001), transfer of neutral and adaptive

532   genetic variation across species (introgression and genetic rescue; Rieseberg & Wendel 1993,

533   Ellstrand & Schierenbeck 2000, Baskett & Gomulkiewicz 2011, Gomulkiewicz & Shaw 2013, Hamilton

534   & Miller 2016), and the evolutionary persistence of hybrids as autonomous entities (with or without

535   speciation; Abbott *et al.,* 2013). The persistence of hybrids over evolutionary time can be achieved

536   through four different, non-mutually exclusive pathways: (1) the stabilisation of hybrid zones (Barton

537   & Hewitt 1989, Baack & Rieseberg 2007), (2) spatial displacement of parental taxa, (3) the expansion

538   of hybrids into new ecological niches (hybrids by virtue of transgressive segregation may be pre-

539   adapted to more extreme habitats, Rieseberg 1991, Rieseberg *et al.,* 1999), and (4) hybrid speciation

540   (Rieseberg & Willis 2007, Abbott *et al.,* 2013) (Fig. 3).

541   Global change through its effects on habitat modification, change in climate, and alterations

542   in ecological interactions, has the potential to alter the outcomes of hybridisation (Campbell &

543   Wendlandt 2013, Chunco 2014). Probably the most obvious example of such an effect is the

544   facilitation of hybrid persistence through "hybridisation of the habitat" in which parents occupy

545   different ecological niches and hybrids a third, intermediate, niche (Anderson 1948, Arnold *et al.,*

546   2012). Under the bounded hybrid superiority model, in which hybrids enjoy a higher fitness in

547   intermediate habitats but suffer from lower fitness in parental ones (Moore 1977, Barton & Hewitt

548 1985), the creation and distribution of these intermediate habitats should influence not only hybrid

549 formation, but also their persistence. Despite the intuitive appeal of the effects of global change on

550 plant hybridisation outcomes, there are few good examples from the British Isles flora. Nevertheless,

551 one tantalizing case for the potential of hybridisation to result in adaptive introgression comes from

552 hybridisation between non-native species of *Rhododendron* (Ericaceae). *Rhododendron ponticum* is

553 widespread in the British Isles and displays the signature of hybridisation with other species,

554 including the more cold-tolerant *R. catawbiense* (Milne & Abbott 2000). Introgression from *R.*

555 *catawbiense* to *R. ponticum* is more prevalent in colder regions in eastern Scotland than elsewhere

556 in Britain. Given this, Milne and Abbott (2000) raised the possibility that gene flow may confer

557 increased cold-tolerance. However, the hypothesis that hybridisation results in increased cold

558 tolerance in *R. ponticum*, and that selection (either natural or artificial) is responsible for the

559 geographic distribution of introgression patterns remains to be tested. The success of hybrids under

560 the novel ecological conditions brought by global change will depend on both the availability and

561 type of new habitats and ecological niches, and on whether hybrids are better equipped to deal with

562 these new environments, either via the ability to thrive in "intermediate" habitats or through the

563 expression of new adaptive phenotypes, e.g., due to transgressive segregation in hybrids.

564 8. Hybrid speciation

565 Although hybridisation may result in the reversal of speciation ("speciation undone", Grant & Grant

566 2014, Taylor *et al.,* 2006), in some cases new species can be formed following hybridisation events

567 (Mallet 2007, Abbott *et al.,* 2013). Hybrid speciation can occur with or without whole genome

568 duplication (allopolyploid or homoploid speciation, respectively) (Soltis & Soltis, 2009). The use of

569 genetic and genomic tools has helped to dramatically increase our understanding of hybrid

570 speciation at both homoploid (Rieseberg 1991, 1997, 2006, Gross *et al.,* 2007) and allopolyploid

571 levels (Hegarty & Hiscock, 2008, Soltis *et al.,* 2014a). Below we present a brief overview of these two

572 modes of hybrid speciation, focusing on those speciation events that have been facilitated by the

573 breakdown of reproductive barriers due to global change, particularly the introduction of non-native

574 species into the British Isles flora.

575   (a) **Allopolyploid hybrid speciation**

576 The origin of a new species through hybridisation and polyploidisation can occur rapidly, as taxa with

577 different chromosome numbers are usually characterised by post-zygotic reproductive barriers

578 (Stebbins 1971). In principle, a hybrid can give rise to a new species in a single generation via

579 polyploidisation (Ramsey & Ramsey 2014) leading to a "cataclysmic origin of species" (Dobzhansky

580 1937, p. 192). Such abrupt speciation is well documented for new British allopolyploid species in the

581 genera *Senecio*, *Spartina,* and *Mimulus*, all of which have evolved in the last 200 years (Ainouche *et*

582 *al.,* 2004, Hegarty *et al.,* 2012, Vallejo-Marín 2012), and all of these cases involve hybrids that owe

583 their origin to global change.

584   The genus *Senecio* (ragworts and groundsels) provides some of the best examples of recent

585 hybrid speciation and 'evolution in action' among the British Flora. Within the last 100 years three

586 new polyploid taxa have arisen in the UK as a consequence of hybridization between native tetraploid

587 *Senecio vulgaris* (common groundsel) and the introduced invasive diploid species *S. squalidus* (Abbott

588 & Lowe 2004, Hegarty *et al.*, 2012), providing one of the first examples of hybridization induced by

589 global change. Hybridisation between these two species gave rise to three allopolyploid taxa -

590 allohexaploid *S. cambrensi*s (Welsh groundsel) tetraploid *S. eboracensis* (York radiate groundsel) and

591 tetraploid *S. vulgaris* var. *hibernicus* (radiate groundsel, a stabilized introgressant form of *S. vulgaris*).

592 The latter of these hybrid taxa occurs sporadically throughout the UK, but *S. eboracensis*, first

593 discovered in a York car park, is now probably extinct (Lowe & Abbott 2000, 2003, Abbott *et al.,* 2009).

594 *Senecio cambrensis*, which was discovered in North Wales in 1948 (Rosser 1955) is locally common in

595 North Wales (Hegarty *et al.*, 2012) and most likely arose following a genome duplication event in the

596 sterile triploid hybrid *S. x baxteri*, as hexaploid plants with similar morphology to the wild form of *S.*

597 *cambrensis* can be produced by treating synthetic triploid *S. vulgaris* x *S. squalidus* hybrids with

598 colchicine (Weir & Ingram 1980, Hegarty *et al.*, 2005). In 1982 *S. cambrensis* was found growing in

599 Edinburgh (Abbott *et al.*, 1983) and subsequent molecular marker analyses revealed that this

600 represented an independent second origin in Scotland (probably during the 1970s) rather than

601 dispersal from Wales (Ashton & Abbott 1992, Harris & Ingram 1992). Unfortunately, the Edinburgh

602 lineage may now be extinct as the species has not been recorded in Edinburgh since 1993 (Abbott &

603 Forbes 2002). Independent origins of recently formed allopolyploids are common as has been shown

604 for the allotetraploid *Tragopogon* of the USA (Soltis *et al.,* 2004) – themselves another example of

605 hybridization facilitated by global change.

606   Another recently discovered example of rapid allopolyploid speciation in the British Isles is

607 *Mimulus peregrinus,* which has evolved at least twice, independently in southern Scotland and the

608 Orkney Isles (Vallejo-Marín *et al.,* 2015; Box 2). As in the case of *Senecio*, *M. peregrinus* owes its

609 origin to global change bringing together previously isolated species. This species is derived from a

610 sexually-sterile triploid hybrid (*M. x robertsii*), which is the product of hybridisation between two

611 non-native species: diploid *M. guttatus* (native to North America) and tetraploid *M. luteus* (native to

612 South America). The triploid hybrid persists in the wild and can form large, naturalised populations

613 where it reproduces clonally (Vallejo-Marín & Lye 2013). Unlike the triploid, the allohexaploid

614 derivatives are pollen and seed fertile (Vallejo-Marín 2012). The mechanism for the formation of the

615 allohexaploid from triploid hybrids is unknown, but one likely route is through mating between

616 unreduced gametes (Husband 2004, Mason & Pires 2015). *Mimulus peregrinus* is an example of how

617 a sterile hybrid taxon has escaped the sterility barrier through genome duplication to form a fertile

618 species.

619   Given the young age (<200 years) of allopolyploids such as *Senecio cambrensis* (Hegarty *et*

620 *al.,* 2012)*, Spartina anglica* (Ainouche *et al.,* 2004)*,* and *M. peregrinus* (Vallejo-Marín 2012)*,* their

621 long term persistence is still uncertain. Allopolyploid species often go through severe bottlenecks

622 during their formation (Soltis *et al.,* 2014a), as potentially a single allopolyploid hermaphroditic

623 individual can give rise to a new allopolyploid taxon. This initially low population size may impose

624 severe ecological barriers, such as costs due to matings between individuals with different ploidies

625 (minority cytotype exclusion, Levin 1975; Figure 3), as well as simple stochastic extinction. Yet

626 allopolyploids tend to be formed recurrently, and most allopolyploid species have multiple origins

627 (Soltis *et al.,* 2014a). To the extent that these independently originated allopolyploids are inter-

628 fertile (Modliszewski & Willis 2012), interpopulation crosses could, in principle, increase genetic and

629 phenotypic variation in nascent lineages, as appears to have been the case in *S. cambrensis* (Abbott

630 *et al.*, 2007). Indeed, inter-population crosses between accessions from the two known localities for

631 *M. peregrinus* produce viable and fertile offspring (Vallejo-Marín *et al.*, *unpublished*), although

632 whether this results in increased phenotypic diversity remains to be established. Determining the

633 degree of phenotypic and ecological diversity of recently allopolyploids seems fundamental to

634 predict their long term fate, particularly as global change continues to alter environments and

635 presents novel challenges to their long term survival.

636   It has long been recognised that hybridisation and polyploidy have the potential to generate

637 ecological novelty (Levin 1983, Otto 2007, Soltis *et al.,* 2014a). The development of genomic tools for

638 recently formed allopolyploids opens the opportunity to take these early hypotheses and investigate

639 the underlying genetic changes associated with ecological innovation. Unlike the case of homoploid

640 hybrids, which have been subject of intense scrutiny at the interphase between ecology and

641 genomics (Yakimowski & Rieseberg 2014), genomic approaches have been rarely used to study the

642 ecology of natural populations of allopolyploids (Ramsey & Ramsey 2014, Soltis *et al.,* 2014b). A rare

643 example of such a study from the British flora is a recent study of the allopolyploid species complex

644 *Dactylorhiza majalis, D. traunsteinerii,* and *D. ebudensis* (Orchidaceae) by Paun *et al.*, (2011).

645 Analysis of genome-wide markers (cDNA-AFLPs) showed that these allopolyploids have higher

646 variation in gene expression than their diploid relatives. Paun *et al.,* (2011) suggest that variable

647 gene expression, and potentially having more plastic phenotypes, have facilitated the colonisation of

648 different ecological niches by these three allopolyploid orchids. A similar situation may be

649   responsible for the success of *S. cambrensis* which also shows altered patterns of gene expression

650   relative to its parents (Hegarty *et al.*, 2006) and also changed patterns of DNA methylation (Hegarty

651   *et al.*, 2011) suggesting the possibility of an epigenetic element to the generation of phenotypic

652   novelty in allopolyploids. Studying the genomic basis and evolution of ecological novelty in

653   polyploids is particularly timely, as we have increasing access to genomic tools for non-model

654   organisms, and because understanding ecological adaptation becomes urgent as populations face

655   the challenge to adapt or perish in the new environments brought by global change.

656   Beyond the earliest stages of allopolyploid speciation, the British Isles flora contains several

657   examples of older allopolyploid taxa, including *Mentha spp., Rubus, Euphrasia, Rorippa,* and

658   *Nasturtium* to name just a few (Stace *et al.,* 2015). A particularly intriguing case is the rare Scottish

659   primrose, *Primula scotica* (2n = 6x = 54, Section *Aleuretia*), endemic to the north of Scotland and

660   Orkney Isles. Early genetic work showed clear evidence that *P. scotica* was formed through an

661   allopolyploidisation event (Glover & Abbott 1995). Furthermore, *P. scotica* is thought to have given

662   rise to another allo-octopolyploid species, *P. scandinavica* (2n = 8x = 72), through a hybridisation

663   event with *P. farinosa* (Guggisberg *et al.,* 2009), and it may even be involved in the origin of the 14x-

664   polyploid *P. stricta* (Guggisberg *et al.,* 2006)*.* Currently, the distribution of *P. scotica*'s putative

665   parents, *P. farinosa* and *P. halleri* (Guggisberg *et al.,* 2009)*,* and its potential derivatives *P.*

666   *scandinavica* and *P. stricta*, does not overlap with *P. scotica*, suggesting that they may occupy

667   different ecological niches, a hypothesis that could be tested using ecological niche modelling (e.g.,

668   McIntyre 2012). Recent work on polyploid taxa in *Primula* Sect. *Aleuretia* has shown that polyploids

669   in this section tend to have a smaller distribution and narrower niche breadths than diploids

670   (Theodoridis *et al.,* 2013), which could indicate that genome duplication is associated with habitat

671   specialisation. *Primula scotica* and other taxa in Section *Aleuretia* are wonderful examples of how

672   allopolyploid taxa can not only persist over evolutionary time, but also continue to speciate and

673   diversify. The development of genomic tools in *Primula*, including a draft reference genome for *P.*

674   *veris* (Nowak *et al.,* 2015), has the potential to elevate this group as a system for the study of

675   evolutionarily established allopolyploids. Notably, the timing of the formation of some of these

676   *Primula* allopolyploids seems to coincide with large-scale environmental changes brought by

677   historical climate change (e.g., end of glacial maxima), which spurs further speculation on the

678   incidence and importance of allopolyploid speciation in times of environmental upheaval.

679   Recently formed hybrids, including those potentially associated with global change, are

680   much more common than the new allopolyploid species they have generated (Stace *et al.,* 2015,

681   Thomas 2015), posing the question, why? Polyploids are thought to be most commonly formed by

682   fusion of unreduced gametes (Levin 2013), and the rate of unreduced gamete formation in hybrids

683   can be extremely high in some plant groups (> 25%, Ramsey & Schemske 1998), indicating ample

684   opportunities for their formation. In the case of sterile hybrids, one key hurdle to overcome in order

685   for an allopolyploidisation event to result in a new taxon is the restoration of sexual fertility. In some

686   cases, genome duplication can restore fertility when sterility arises from structural or numeric

687   differences in the parental chromosome sets (Stebbins 1958, Coyne & Orr 2004). However, if sterility

688   is rooted in genic incompatibilities between the parents, genome duplication alone will not be

689   sufficient for fertility restoration. Thus one would expect that those hybrids that have produced

690   allopolyploid species should tend to show few or no genic incompatibilities. This hypothesis can be

691   experimentally tested by inducing polyploidy in sterile hybrids (Hegarty *et al.,* 2013) and assessing

692   their level of fertility (Stathos & Fishman 2014). If synthetic polyploids are as sterile as their parents,

693   then genic incompatibilities can be inferred (Coyne & Orr 2004). If polyploidy restores fertility, the

694   absence of allopolyploids in nature could be explained by ecological or perhaps stochastic causes.

695   Ultimately, determining why allopolyploids are rarer than their hybrid ancestors remains an

696   open question. Their absence is puzzling given the high rate of unreduced gamete production in

697   hybrids (Ramsey and Schemske 1998). Beyond the barrier of overcoming sexual sterility in

698   allopolyploids, other extrinsic post-zygotic mechanisms such as minority cytotype disadvantage

699   (Levin 1975), and ecological competition with their hybrid ancestors, may pose significant barriers to

700   the establishment of recently formed allopolyploids. It is also possible that our records of neo-

701   allopolyploids will continue to accumulate as more recently formed hybrids come of age (Thomas

702   2015). Recently formed allopolyploids may be difficult to spot in botanical surveys, particularly in the

703   initial stages when they are quite rare. Large-scale searches for cryptic allopolyploids among recently

704   formed hybrids, using direct or indirect measurements of genome size such as flow cytometry (e.g.,

705   Castro *et al.*, 2012), and pollen and stomata size, may provide an effective way to establish if recent

706   allopolyploids are indeed as rare as they appear to be.

707   (b) **Homoploid hybrid speciation**

708   In contrast to allopolyploid speciation, homoploid hybrid speciation appears less common, although

709   the number of confirmed cases of homoploid hybrid speciation is increasing (Gross & Rieseberg

710   2005, Abbott *et al.,* 2013, Schumer *et al.,* 2014). This apparent rarity of homoploid speciation may

711   therefore simply reflect the difficulty in detecting it (Abbott *et al.,* 2013). Homoploid hybrid

712   speciation requires the evolution of a hybrid that is reproductively isolated from its parents in the

713   absence of differences in ploidy level (Gross & Rieseberg 2005, Yakimowski & Rieseberg 2014).

714   Reproductive isolation between parental and hybrid taxa could thus be based on differences in

715   chromosome structure, ecological divergences, spatial isolation, or a combination of the above

716   (Rieseberg 1997). In the British Isles flora, the only confirmed case of recent homoploid hybrid

717 speciation is *Senecio squalidus* (Oxford ragwort) although the place of origin of the hybrid plants was

718 Sicily (reviewed in Abbott *et al.,* 2013). *S. squalidus* is a recent homoploid hybrid species, which

719 evolved in the UK following its introduction from a *S. aethnensis* x *S. chrysanthemifolius* hybrid zone

720 on Mt Etna ~300 years ago.

721   *Senecio aethnensis* is endemic to of high altitudes of Mt Etna, whereas *S. chrysanthemifolius*, a

722 native of Sicily, is more widespread at lower altitudes. At mid altitudes on the volcano the

723 distribution of the two species frequently overlaps leading to the formation of stable hybrid zones

724 around the circumference of the volcano (Brennan *et al.,* 2009). Material from this hybrid zone was

725 introduced to the Oxford Botanic Garden in the early 1700s from where plants subsequently

726 escaped and colonized the masonry of college walls and roadsides. During the industrial revolution

727 the clinker beds of the expanding railway network provided an ideal habitat (akin to the volcanic

728 slopes of Mt Etna) for *S. squalidus* to thrive and spread. During the next 300 years *S. squalidus*

729 diverged phenotypically and to a lesser extent genetically in allopatric isolation from its parental

730 species, such that it now meets the criteria for a new homoploid species (James & Abbott 2005,

731 Abbott *et al.,* 2013). *S. squalidus* and its parental species are divergent in their morphology,

732 flowering phenology, and in ecologically important traits even though they are genetically very

733 similar and completely interfertile (Chapman *et al.,* 2013, Osborne *et al.,* 2013). Nevertheless, the

734 ~300 years of allopatric isolation have allowed *S. squalidus* to adapt to the cooler climate and non-

735 volcanic soils of the UK, resulting in the ecological and phenotypic divergence seen today.

736   Other potential candidates for homoploid hybrids in the British flora are found in the complex

737 assemblage of interspecific hybrids in annual, hemiparasitic *Euphrasia*. Species of *Euphrasia* in the

738 British Isles include both diploid and tetraploid taxa, which are strongly reproductively isolated

739 across ploidy levels (Yeo 1968). However, hybrids within ploidy levels are often fertile (Stace *et al.,*

740 2015), and Yeo (1968) suggests that some of these hybrids may have produced new hybrid taxa. The

741 high fertility of within-ploidy hybrids means that for homoploid speciation to occur, reproductive

742 isolation between parental and hybrid taxa will depend on ecological divergence and spatial isolation

743 (Yeo 1968, Rieseberg 1997). Future studies of *Euphrasia* will determine whether the hypothesis of

744 homoploid hybrid speciation holds for this group.

745   Taken at face value, homoploid hybrid speciation seems to be rarer than polyploid hybrid

746 speciation (Abbott *et al.*, 2013). The apparent higher facility of allopolyploids to form and establish

747 may be linked, in part, to the reproductive isolation barriers introduced by differences in ploidy level

748 between derivative and parental taxa. But allopolyploid taxa also have the added feature of

749 genome-wide redundancy conferred by polyploidisation (Soltis *et al.*, 2014a). It is tempting to

750 speculate that genomic redundancy is causally linked to evolutionary success in allopolyploids, for

751 example by allowing duplicated subgenomes to differentiate and specialise in different functions

752 (subfunctionalisation and neofunctionalisation; Ohno 1970; Soltis *et al.*, 2014b). This higher

753 evolutionary potential of allopolyploids may not be immediately realised upon genome duplication

754 (or seen in synthetic allopolyploids; Hegarty *et al.*, 2006), but instead may accumulate as lineages

755 diverge. The higher incidence of allopolyploids *vs.* homoploid hybrid species may thus reflect an

756 evolutionary advantage over the longer term in taxa with highly redundant and malleable genomes.

757 Whether recently formed allopolyploids can keep up with rapid global change remains to be seen.

V. Outlook

759 Hybridisation and hybrid speciation permeates the evolutionary history of plants, and is certainly not

760 a novel phenomenon. However, the accelerating transformation of the world's biota brought about

761 by global change make the study of hybridisation both current and urgent if we are to understand

762 the consequences of eroding reproductive barriers between evolutionarily distinct lineages. For

763 instance, hybridisation can threaten the conservation of local biodiversity by altering the genetic

764 integrity of native species through introgression, and genetically swamp rare ones (Vilà *et al.*, 2000,

765 Wolf *et al.*, 2001, Brennan *et al.*, 2015, Chown *et al.*, 2015, Taylor *et al.*, 2015). Changes to ecological

766 communities due to hybridisation involving non-native species can also change ecological

767 communities, and hybrids can compete for pollination services and fruit dispersers (Vilà *et al.*, 2000).

768 Moreover, hybrids themselves can become significant invasive pests (Ellstrand & Schierenbeck 2000)

769 with negative effects for local economies and biodiversity. On the other hand, the consequences of

770 hybridisation and its effects on biodiversity need not necessarily always be negative since

771 hybridisation can spur accelerated rates of speciation and thus increase biodiversity (Thomas 2015).

772 More studies are needed to gauge whether the balance of neo-hybridisation is positive or negative.

773 Although the ecological and evolutionary consequences of recently formed hybrids are hard to

774 predict, many hybrids are probably innocuous to local ecosystems as we know them. Regardless of

775 their perceived costs and benefits, recently formed hybrids provide us with the unique opportunity

776 to study, in real time, the breakdown of reproductive barriers and the processes allowing newly

777 formed hybrids to become established, and in some cases speciate (Mallet 2007, Abbott *et al.*,

778 2013). The dynamic and rapidly changing nature of our planet's biota has created a global

779 experiment on hybridisation. We now have the timely opportunity to use this unplanned experiment

780 to study the basic biological phenomena responsible for the birth and death of species. Capitalizing

781 on this opportunity needs a multidisciplinary approach, combining classic taxonomic studies and

782 catalogues of hybrid floras (Stace *et al.*, 2015) with ecological experiments, phylogenetic analysis,

783 quantitative genetics, and bioinformatics to exploit the new genomic resources available for non-

784 model organisms.

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References

792 **Abbott R, Albach D, Ansell S, Arntzen JW, Baird SJE, Bierne N, Boughman JW, Brelsford A, Buerkle**

793   **CA, Buggs R, Butlin RK, Dieckmann U, Eroukhmanoff F, Grill A, Cahan SH, Hermansen JS,**

794   **Hewitt G, Hudson AG, Jiggins C, Jones J, Keller B, Marczewski T, Mallet J, Martínez-Rodríguez**

795   **P, Moest M, Mullen S, Nichols R, Nolte AW, Parisod C, Pfennig K, Rice AM, Ritchie MG, Seifert**

796   **B, Smadja CM, Stelkens R, Szymura JM, Vainola R, Wolf JBW, and Zinner D. 2013.**

797   Hybridization and speciation. *J. Evol. Biol.* **26**: 229-246. 10.1111/j.1420-9101.2012.02599.x.

798 **Abbott RJ, Brennan AC, James JK, Forbes DG, Hegarty MJ, and Hiscock SJ. 2009.** Recent hybrid

799   origin and invasion of the British Isles by a self-incompatible species, Oxford ragwort (*Senecio*

800   *squalidus* L., Asteraceae). *Biol. Invasions* **11**: 1145-1158. 10.1007/s10530-008-9382-3.

801 **Abbott RJ, and Forbes DG. 2002.** Extinction of the Edinburgh lineage of the allopolyploid neospecies,

802   *Senecio cambrensis* Rosser (Asteraceae). *Heredity* **88:** 267-269.

803 **Abbott RJ, Ireland HE, and Rogers HJ. 2007.** Population decline despite high genetic diversity in the

804   new allopolyploid species *Senecio cambrensis* (Asteraceae). *Molecular Ecology*, **16:** 1023-1033.

805 **Abbott RJ, and Lowe AJ. 2004.** Origins, establishment and evolution of new polyploid species:

806   *Senecio cambrensis* and *S. eboracensis* in the British Isles. *Biological Journal of the Linnean*

807   *Society* **82**: 467-474.

808 **Abbott RJ, Noltie HJ, and Ingram R. 1983.** The origin and distribution of *Senecio cambrensis* Rosser

809   in Edinburgh. *Transactions of the Botanical Society of* Edinburgh **44:** 103-106.

810 **Ainouche M, Baumel A, Salmon A, and Yannic G. 2004.** Hybridization, polyploidy and speciation in

811   *Spartina* (Poaceae). *New Phytol.* **161**: 165-172. 10.1046/j.1469-8137.2003.00926.x.

812 **Anderson E. 1948.** Hybridization of the habitat. *Evolution*: 1-9.

813 **Arnold ML. 1997.** *Natural Hybridization and Evolution.* Oxford University Press.

814 **Arnold ML, Ballerini ES, and Brothers AN. 2012.** Hybrid fitness, adaptation and evolutionary
815   diversification: lessons learned from Louisiana Irises. *Heredity* **108**: 159-166.
816   10.1038/hdy.2011.65.

817 **Arnold ML, and Hodges SA. 1995.** Are natural hybrids fit or unfit relative to their parents? *Trends in*
818   *Ecology & Evolution* **10**: 67-71. 10.1016/S0169-5347(00)88979-X.

819 **Ashton PA, and Abbott RJ. 1992.** Multiple origins and genetic diversity in the newly arisen
820   allopolyploid species, *Senecio cambrensis* Rosser (Compositae). *Heredity* **68:** 25-32.

821 **Baack E, Melo MC, Rieseberg LH, and Ortiz-Barrientos D. 2015.** The origins of reproductive isolation
822   in plants. *New Phytologist.* **207**: 968-984.

823 **Baack EJ, and Rieseberg LH. 2007.** A genomic view of introgression and hybrid speciation. *Current*
824   *Opinion in Genetics & Development* **17**: 513-518.

825 **Bakker FT, Lei D, Yu J, Mohammadin S, Wei Z, Kerke S, Gravendeel B, Nieuwenhuis M, Staats M,**
826   **and Alquezar-Planas DE. 2016.** Herbarium genomics: plastome sequence assembly from a
827   range of herbarium specimens using an Iterative Organelle Genome Assembly pipeline.
828   *Biological Journal of the Linnaean Society* **117:** 33-43. doi: 10.1111/bij.12642

829 **Barcaccia G, Arzenton F, Sharbel TF, Varotto S, Parrini P, and Lucchin M. 2006.** Genetic diversity
830   and reproductive biology in ecotypes of the facultative apomict *Hypericum perforatum* L.
831   *Heredity*, **96**: 322-334.

832 **Barton NH. 2001.** The role of hybridization in evolution. *Molecular Ecology* **10**: 551-568.

833 **Barton NH, and Hewitt G. 1985.** Analysis of hybrid zones. *Annual Review of Ecology and Systematics*:
834   113-148.

835 **Barton NH, and Hewitt GM. 1989.** Adaptation, speciation and hybrid zones. *Nature* **341**: 497-503.
836   doi: 10.1038/341497a0

837 **Baskett ML, and Gomulkiewicz R. 2011.** Introgressive hybridization as a mechanism for species
838   rescue. *Theoretical Ecology* **4**: 223-239.

839 **Bengtsson BO. 2003.** Genetic variation in organisms with sexual and asexual reproduction. *Journal of*
840   *Evolutionary Biology* **16**: 189-199.

841 **Bomblies K, and Weigel D. 2007.** Hybrid necrosis: autoimmunity as a potential gene-flow barrier in
842   plant species. *Nature Reviews Genetics* **8**: 382-393.

843 **Bradshaw HD, Wilbert SM, Otto KG, and Schemske DW. 1995.** Genetic-mapping of floral traits
844   associated with reproductive isolation in monkeyflowers (*Mimulus*). *Nature* **376**: 762-765.

845 **Brandvain Y, Kenney AM, Flagel L, Coop G, and Sweigart AL. 2014.** Speciation and introgression
846   between *Mimulus nasutus* and *Mimulus guttatus*. *PLoS Genetics.* 10(6): e1004410. doi:
847   10.1371/journal.pgen.1004410

848   **Brennan AC, Bridle JR, Wang A, Hiscock SJ, and Abbott RJ. 2009.** Adaptation and selection in the
849   *Senecio* (Asteraceae) hybrid zone on Mount Etna, Sicily. *New Phytologist* **183**: 702-717. doi:
850   10.1111/j.1469-8137.2009.02944.x

851   **Brennan AC, Woodward G, Seehausen O, Muñoz-Fuentes V, Moritz C, Guelmami A, Abbott RJ, and**
852   **Edelaar P. 2015.** Hybridization due to changing species distributions: adding problems or
853   solutions to conservation of biodiversity during global change? *Evolutionary Ecology Research*
854   **16**: 475-491.

855   **Bretagnolle FA, and Thompson J. 1995.** Gametes with the somatic chromosome number:
856   mechanisms of their formation and role in the evolution of autopolyploid plants. *New*
857   *Phytologist.* **129**:1-22. doi: 10.1111/j.1469-8137.1995.tb03005.x

858   **Bridle JR, Buckley J, Bodsworth EJ, and Thomas CD. 2013.** Evolution on the move: specialization on
859   widespread resources associated with rapid range expansion in response to climate change.
860   *Proceedings of the Royal Society. B: Biological Sciences.* **281**: 20131800. doi:
861   10.1098/rspb.2013.1800.

862   **Briggs D, and Walters SM. 1997.** *Plant Variation and Evolution.* Cambridge, U.K.: Cambridge
863   University Press.

864   **Brys R, Broeck AV, Mergeay J, and Jacquemyn H. 2014.** The contribution of mating system variation
865   to reproductive isolation in two closely related *Centaurium* species (Gentianaceae) with a
866   generalized flower morphology. *Evolution* **68**: 1281-1293.

867   **Buggs R. 2007.** Empirical study of hybrid zone movement. *Heredity* **99**: 301-312.

868   **Campbell DR, and Wendlandt C. 2013.** Altered precipitation affects plant hybrids differently than
869   their parental species. *American Journal of Botany* **100**: 1322-1331. doi: 10.3732/ajb.1200473

870   **Castro M, Castro S, and Loureiro J. 2012.** Genome size variation and incidence of polyploidy in
871   Scrophulariaceae *sensu lato* from the Iberian Peninsula. *AoB plants*. **2012:** pls037. doi:
872   10.1093/aobpla/pls037

873   **Chapman MA, and Abbott RJ. 2010.** Introgression of fitness genes across a ploidy barrier. *New*
874   *Phytologist.* **186**: 63-71. doi: 10.1111/j.1469-8137.2009.03091.x

875   **Chapman MA, Hiscock SJ, and Filatov DA. 2013.** Genomic divergence during speciation driven by
876   adaptation to altitude. *Molecular Biology and Evolution* **30**: 2553-2567. doi:
877   10.1093/molbev/mst168

878   **Chown SL, Hodgins KA, Griffin PC, Oakeshott JG, Byrne M, and Hoffmann AA. 2015.** Biological
879   invasions, climate change and genomics. *Evolutionary Applications* **8**: 23-46.
880   10.1111/eva.12234.

881 **Chunco AJ. 2014.** Hybridization in a warmer world. *Ecology and Evolution* **4**: 2019-2031. doi:
882   10.1002/ece3.1052

883 **Cleland EE, Chuine I, Menzel A, Mooney HA, and Schwartz MD. 2007.** Shifting plant phenology in
884   response to global change. *Trends in Ecology and Evolution* **22**: 357-365.

885 **Clement EJ, and Foster M. 1994.** *Alien Plants of the British Isles.* London: Botanical Society of the
886   British Isles.

887 **Comai L. 2005.** The advantages and disadvantages of being polyploid. *Nature Reviews Genetics* **6**:
888   836-846.

889 **Coyne JA, and Orr HA. 2004.** *Speciation.* Sunderland, Massachusetts, U.S.A.: Sinauer Associates.

890 **De Storme N, Mason A. 2014.** Plant speciation through chromosome instability and ploidy change:
891   Cellular mechanisms, molecular factors and evolutionary relevance. *Current Plant Biology* **1**: 10-
892   33.

893 **Dobzhansky TG. 1937.** *Genetics and the Origin of Species.* New York: Columbia University Press.

894 **Dodsworth S. 2015.** Genome skimming for next-generation biodiversity analysis. *Trends in Plant*
895   *Sciences* **20**: 525-527.

896 **Ellstrand NC, and Schierenbeck KA. 2000.** Hybridization as a stimulus for the evolution of
897   invasiveness in plants? *Proceedings of the National Academy of Sciences U. S. A.* **97**: 7043-7050.

898 **Ellstrand NC, and Elam DR. 1993.** Population genetic consequences of small population size:
899   implications for plant conservation. *Annual Review of Ecology and Systematics* **24:** 217-242.

900 **Fay MF 2015**. British and Irish orchids in a changing world. *Curtis's Botanical Magazine* **32**: 3-23.

901 **Fishman L, and Willis JH. 2001.** Evidence for Dobzhansky-Muller incompatibilities contributing to the
902   sterility of hybrids between *Mimulus guttatus* and *M. nasutus*. *Evolution* **55**: 1932-1942.

903 **Garroway CJ, Bowman J, Cascaden TJ, Holloway GL, Mahan CG, Malcolm JR, Steele MA, Turner G,**
904   **and Wilson PJ. 2010.** Climate change induced hybridization in flying squirrels. *Global Change*
905   *Biology* **16**: 113-121.

906 **Glover BJ, and Abbott RJ. 1995.** Low genetic diversity in the Scottish endemic *Primula scotica* Hook.
907   *New Phytologist* **129**: 147-153.

908 **Gomulkiewicz R, and Shaw RG. 2013.** Evolutionary rescue beyond the models. *Philosophical*
909   *Transactions of the Royal Society of London. B: Biological Sciences.* **368**: 20120093. doi:
910   10.1098/rstb.2012.0093.

911 **Grant PR, and Grant BR. 2014.** Evolutionary biology: Speciation undone. *Nature* **507**: 178-179.

912 **Grant V. 1966.** Selection for vigor and fertility in the progeny of a highly sterile species hybrid in
913   *Gilia*. *Genetics* **53**: 757-776.

914 **Grant V. 1971.** *Plant Speciation.* New York: Columbia University Press.

915 **Griffiths AJ, Miller JH, Lewontin RC, and Suzuki DT. 2000.** *An Introduction to Genetic Analysis.* New
916   York: WH Freeman.

917 **Gross BL, and Rieseberg LH. 2005.** The ecological genetics of homoploid hybrid speciation. *Journal of*
918   *Heredity.* **96**: 241-252. 10.1093/jhered/esi026

919 **Gross BL, Turner KG, and Rieseberg LH. 2007.** Selective sweeps in the homoploid hybrid species
920   *Helianthus deserticola*: evolution in concert across populations and across origins. *Molecular*
921   *Ecology* **16**: 5246-5258. 10.1111/j.1365-294X.2007.03574.x

922 **Guggisberg A, Mansion G, Kelso S, and Conti E. 2006.** Evolution of biogeographic patterns, ploidy
923   levels, and breeding systems in a diploid–polyploid species complex of Primula. *New Phytologist*
924   **171**: 617-632.

925 **Guggisberg A, Mansion G, and Conti E. 2009.** Disentangling reticulate evolution in an arctic–alpine
926   polyploid complex. *Systematic Biology.* **58:** 55-73. doi: 10.1093/sysbio/syp010

927 **Hamilton JA, and Miller JM. 2016.** Adaptive introgression: A resource for management and genetic
928   conservation in a changing climate. *Conservation Biology* **30:** 33–41. doi: 10.1111/cobi.12574

929 **Harris SA, and Ingram R. 1992.** Molecular systematics of the genus *Senecio* L. 1. Hybridization in a
930   British polyploid complex. *Heredity* **69:** 1-10.

931 **Hegarty MJ, Batstone TOM, Barker GL, Edwards KJ, Abbott RJ, and Hiscock SJ. 2011.** Nonadditive
932   changes to cytosine methylation as a consequence of hybridization and genome duplication in
933   *Senecio* (Asteraceae). *Molecular Ecology*, **20:** 105-113.

934 **Hegarty MJ, Jones JM, Wilson ID, Barker GL, Coghill JA, Sanchez-Baracaldo P, Buggs RJA, Liu G,**
935   **Abbott RJ, Edwards KJ, Hiscock SJ. 2005.** Development of anonymous cDNA microarrays to
936   study changes to the *Senecio* floral transcriptome during hybrid speciation. *Molecular Ecology*
937   **14**: 2493-2510.

938 **Hegarty MJ, Barker GL, Wilson ID, Abbott RJ, Edwards KJ, Hiscock SJ. 2006** Transcriptome shock
939 after interspecific hybridization in *Senecio* is ameliorated by genome duplication. *Current Biology* **16**:
940 1652-1659.

941 **Hegarty MJ, Hiscock SJ. 2008.** Genomic clues to the evolutionary success of polyploid plants. *Current*
942 *Biology* **18**: R435-R444.

943 **Hegarty MJ, Abbott RJ, and Hiscock SJ. 2012.** Allopolyploid speciation in action: The origins and
944   evolution of *Senecio cambrensis*. Pages 245-270 In: Soltis PS, and Soltis DE, eds. *Polyploidy and*
945   *genome evolution.* Berlin: Springer-Verlag. 245-270.

946 **Hegarty M, Coate J, Sherman-Broyles S, Abbott R, Hiscock S, and Doyle J. 2013.** Lessons from
947   natural and artificial polyploids in higher plants. *Cytogenetics and Genome Research.* **140**: 204-
948   225. doi: 10.1159/000353361

949   **Higgins SI, and Richardson DM. 1999.** Predicting plant migration rates in a changing world: the role

950   of long-distance dispersal. *American Naturalist* **153**: 464-475.

951   **Hiscock SJ, Doughty J, and Dickinson HG. 1998.** Unilateral incompatibility and the *S* (self-

952   incompatibility) locus. In: Owens SJ and Rudall PJ. *Reproductive Biology in Systematics,*

953   *Conservation and Economic Botany.* Richmond, Surrey, UK: Royal Botanic Gardens, Kew. 31-46.

954   **Hoffmann AA, and Sgrò CM. 2011.** Climate change and evolutionary adaptation. *Nature* **470**: 479-

955   485.

956   **Hollister JD. 2015.** Polyploidy: adaptation to the genomic environment. *New Phytologist* **205**: 1034-

957   1039. doi: 10.1111/nph.12939

958   **Howard DJ. 1999.** Conspecific sperm and pollen precedence and speciation. *Annual Review of*

959   *Ecology and Systematics* **30**: 109-132.

960   **Howard H, and Manton I. 1946.** Autopolyploid and allopolyploid watercress with the description of

961   a new species. *Annals of Botany* **10**: 1-13.

962   **Hulme PE, Bacher S, Kenis M, Klotz S, Kühn I, Minchin D, Nentwig W, Olenin S, Panov V, and Pergl**

963   **J. 2008.** Grasping at the routes of biological invasions: a framework for integrating pathways

964   into policy. *Journal of Applied Ecology* **45**: 403-414.

965   **Husband BC. 2004.** The role of triploid hybrids in the evolutionary dynamics of mixed-ploidy

966   populations. *Biological Journal of the Linnean Society* **82**: 537-546.

967   **Husband BC, Schemske DW, Burton TL, and Goodwillie C. 2002.** Pollen competition as a unilateral

968   reproductive barrier between sympatric *Chamerion angustifolium*. *Proceedings of the Royal*

969   *Society of London Series B: Biological Sciences* **269**: 2565-2571.

970   **James JK, and Abbott RJ. 2005.** Recent, allopatric, homoploid hybrid speciation: The origin of

971   *Senecio squalidus* (Asteraceae) in the British Isles from a hybrid zone on Mount Etna, Sicily.

972   *Evolution* **59**: 2533-2547.

973   **Karlsdóttir L, Hallsdóttir M, Thórsson AT, and Anamthawat-Jónsson K. 2008.** Characteristics of

974   pollen from natural triploid *Betula* hybrids. *Grana* **47**: 52-59.

975   **Kilvitis HJ, Alvarez M, Foust CM, Schrey AW, Robertson M., and Richards CL, 2014.** Ecological

976   epigenetics. *In*: Landy CR, and Aubin-Hort N (eds), *Ecological Genomics: Ecology and the*

977   *Evolution of Genes and Genomes.* Springer Netherlands. pp. 191-210.

978   **Knobloch IW. 1972.** Intergeneric hybridization in flowering plants. *Taxon*: 97-103.

979   **Köhler C, Scheid OM, and Erilova A. 2010.** The impact of the triploid block on the origin and

980   evolution of polyploid plants. *Trends in Genetics* **26**: 142-148. 10.1016/j.tig.2009.12.006.

981   **Levin DA. 1975.** Minority cytotype exclusion in local plant populations. *Taxon*: 35-43.

982 **Levin DA. 1983.** Polyploidy and novelty in flowering plants. *American Naturalist*: 1-25.

983   http://www.jstor.org/stable/2461003

984 **Levin DA. 2002.** *The Role of Chromosomal Change in Plant Evolution.* Oxford, UK: Oxford University

985   Press.

986 **Levin DA. 2013.** The timetable for allopolyploidy in flowering plants. *Annals of Botany*, **112:** 1201-

987   1208.

988 **Lousley J. 1946.** A new hybrid Senecio from the London area. *Report of the Botanical Society and*

989   *Exchange Club of the British Isles* **12**: 869-874.

990 **Lowe AJ, and Abbott RJ. 2000.** Routes of origin of two recently evolved hybrid taxa: *Senecio vulgaris*

991   *var. hibernicus* and York radiate groundsel (Asteraceae). *American Journal of Botany* **87:** 1159-

992   1167.

993 **Lowe AJ, and Abbott RJ. 2003.** A new British species, *Senecio eboracensis* (Asteraceae), another

994   hybrid derivative of *S. vulgaris* L. and *S. squalidus* L. *Watsonia* **24:** 375-388.

995 **Lowry DB, Modliszewski JL, Wright KM, Wu CA, and Willis JH. 2008.** The strength and genetic basis

996   of reproductive isolating barriers in flowering plants. *Philosophical Transactions of the Royal*

997   *Society B-Biological Sciences* **363**: 3009-3021.

998 **Ludwig S, Robertson A, Rich TCG, Djordjevic M, Cerovic R, Houston L, Harris SA, Hiscock SJ (2013)**

999   Breeding systems, hybridization and continuing evolution in Avon Gorge *Sorbus*. *Annals of*

1000   *Botany* **111**: 563-575.

1001 **Mable BK. 2013**. Polyploids and hybrids in changing environments: winners or losers in the struggle

1002   for adaptation? *Heredity* **110**: 95-96. doi: 10.1038/hdy.2012.105

1003 **Mack RN. 2005.** Predicting the identity of plant invaders: future contributions from horticulture.

1004   *HortScience* **40**: 1168-1174.

1005 **Mallet J. 2007.** Hybrid speciation. *Nature* **446**: 279-283. doi: 10.1038/nature05706.

1006 **Marren P. 1999.** Britain's Rare Flowers. London: A & C Black Publishers Ltd.

1007 **Martin NH, Sapir Y, and Arnold ML. 2008.** The genetic architecture of reproductive isolation in

1008   Louisiana irises: Pollination syndromes and pollinator preferences. *Evolution* **62**: 740-752.

1009 **Mason AS, and Pires JC. 2015.** Unreduced gametes: meiotic mishap or evolutionary mechanism?

1010   *Trends in Genetics* **31:** 5-10. doi: 10.1016/j.tig.2014.09.011

1011 **Mayr E. 1942.** *Systematics and the Origin of Species, from the Viewpoint of a Zoologist.* Cambridge,

1012   Massachusetts: Harvard University Press.

1013 **McIntyre PJ. 2012.** Polyploidy associated with altered and broader ecological niches in the *Claytonia*

1014   *perfoliata* (Portulacaceae) species complex. *American Journal of Botany* **99**: 655-662. doi:

1015   10.3732/ajb.1100466.

1016 **Milne RI, and Abbott RJ. 2000.** Origin and evolution of invasive naturalized material of

1017   *Rhododendron ponticum* L. in the British Isles. *Molecular Ecology* **9**: 541-556. doi:

1018   10.1046/j.1365-294x.2000.00906.x.

1019 **Modliszewski JL, and Willis JH. 2012.** Allotetraploid *Mimulus sookensis* are highly interfertile despite

1020   independent origins. *Molecular Ecology* **21**: 5280-5298. doi: 10.1111/j.1365-294X.2012.05706.x.

1021 **Mooney H, and Cleland E. 2001.** The evolutionary impact of invasive species. *Proceedings of the*

1022   *National Academy of Sciences, U. S. A.* **98**: 5446-5451. 10.1073/pnas.091093398.

1023 **Moore WS. 1977.** An evaluation of narrow hybrid zones in vertebrates. *Quarterly Review of Biology*

1024   **52**: 263-277.

1025 **Morales CL, and Traveset A. 2009.** A meta-analysis of impacts of alien vs. native plants on pollinator

1026   visitation and reproductive success of co-flowering native plants. *Ecology Letters* **12**: 716-728.

1027 **Moyle LC, and Graham EB. 2005.** Genetics of hybrid incompatibility *between Lycopersicon*

1028   *esculentum* and *L. hirsutum. Genetics* **169**: 355-373. doi: 10.1534/genetics.104.029546

1029 **Moyle LC, Olson MS, and Tiffin P. 2004.** Patterns of reproductive isolation in three angiosperm

1030   genera. *Evolution* **58**: 1195-1208.

1031 **Myers JH, and Bazely D. 2003.** *Ecology and Control of Introduced Plants.* Cambridge, U.K.: Cambridge

1032   University Press.

1033 **Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U, Poot P, Purugganan MD,**

1034   **Richards CL, Valladares F, and van Kleunen M. 2010.** Plant phenotypic plasticity in a changing

1035   climate. *Trends in Plant Science* **15:** 684-692.

1036 **Nowak MD, Russo G, Schlapbach R, Huu CN, Lenhard M, and Conti E. 2015.** The draft genome of

1037   *Primula veris* yields insights into the molecular basis of heterostyly. *Genome Biology* **16**: 12 doi:

1038   10.1186/s13059-014-0567-z

1039 **Ohno S. 1970.** *Evolution by Gene Duplication*. Springer-Verlag. New York.

1040 **Osborne OG, Batstone TE, Hiscock SJ, and Filatov DA. 2013.** Rapid speciation with gene flow

1041   following the formation of Mt. Etna. *Genome Biology and Evolution* **5**: 1704-1715. doi:

1042   10.1093/gbe/evt127

1043 **Otto SP. 2007.** The evolutionary consequences of polyploidy. *Cell* **131**: 452-462. doi:

1044   10.1016/j.cell.2007.10.022

1045 **Otto SP, and Whitton J. 2000.** Polyploid incidence and evolution. *Annual Review of Genetics* **34**: 401-

1046   437. 10.1146/annurev.genet.34.1.401

1047 **Paun O, Bateman RM, Fay MF, Luna JA, Moat J, Hedren M, and Chase MW. 2011.** Altered gene

1048   expression and ecological divergence in sibling allopolyploids of *Dactylorhiza* (Orchidaceae).

1049   *BMC Evolutionary Biology* **11**: 113. doi: 10.1186/1471-2148-11-113

1050   **Preston CD, and Croft JM. 1998.** Britain's changing aquatic flora. *British Wildlife* **10**: 18-28.

1051   **Preston CD, Pearman DA, and Dines TD. 2002.** New Atlas of the British and Irish Flora. Oxford, UK:
1052   Oxford University Press.

1053   **Preston CD, and Pearman DA. 2015.** Plant hybrids in the wild: evidence from biological recording.
1054   *Biological Journal of the Linnean Society.* **115**: 555-572.

1055   **Ramsey J, Bradshaw HD, and Schemske DW. 2003.** Components of reproductive isolation between
1056   the monkeyflowers *Mimulus lewisii* and *M. cardinalis* (Phrymaceae). *Evolution* **57**: 1520-1534.

1057   **Ramsey J, and Ramsey TS. 2014.** Ecological studies of polyploidy in the 100 years following its
1058   discovery. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* **369**:
1059   20130352. doi: 10.1098/rstb.2013.0352

1060   **Ramsey J, and Schemske DW. 1998.** Pathways, mechanisms, and rates of polyploid formation in
1061   flowering plants. *Annual Review of Ecology and Systematics* **29**: 467-501.

1062   **Rich TCG, Houston L, Robertson A, and Proctor MCF. 2010.** *Whitebeams, Rowans and Service Trees*
1063   *of Britain and Ireland. A monograph of British and Irish Sorbus L.* London: Botanical Society of
1064   the British Isles.

1065   **Rich TCG, and Robertson A. 2015.** *Sorbus* L. In: *Hybrid Flora of the British Isles.* Stace CA, Preston CD,
1066   and Pearman DA (eds). Bristol, U.K.: Botanical Society of Britain and Ireland.

1067   **Rieseberg LH. 1991.** Homoploid reticulate evolution in *Helianthus* (Asteraceae). Evidence from
1068   ribosomal genes. *American Journal of Botany* **78**: 1218-1237. doi: 10.2307/2444926.

1069   **Rieseberg LH. 1997.** Hybrid origins of plant species. *Annual Review of Ecology and Systematics* **28**:
1070   359-389. 10.1146/annurev.ecolsys.28.1.359

1071   **Rieseberg LH. 1995.** The role of hybridization in evolution. Old wine in new skins. *American Journal*
1072   *of Botany* **82**: 944-953. doi: 10.2307/2445981

1073   **Rieseberg LH. 2001.** Chromosomal rearrangements and speciation. *Trends in Ecology & Evolution* **16**:
1074   351-358. doi: 10.1016/S0169-5347(01)02187-5

1075   **Rieseberg LH. 2006.** Hybrid speciation in wild sunflowers. *Annals of the Missouri Botanical Garden*
1076   **93**: 34-48.

1077   **Rieseberg LH, Archer MA, and Wayne RK. 1999.** Transgressive segregation, adaptation and
1078   speciation. *Heredity* **83**: 363-372.

1079   **Rieseberg LH, and Blackman BK. 2010.** Speciation genes in plants. *Annals of Botany* **106**: 439-455.
1080   doi: 10.1093/aob/mcq126.

1081   **Rieseberg LH, and Brunsfeld SJ. 1992.** Molecular evidence and plant introgression. Pages 151-176 In:
1082   Soltis PS, Soltis DE, and Doyle JJ. *Molecular systematics of plants.* New York, U.S.A.: Springer,
1083   151-176.

1084 **Rieseberg LH, and Carney SE. 1998.** Plant hybridization. *New Phytologist* **140**: 599-624.

1085 **Rieseberg LH, and Ellstrand NC. 1993.** What can molecular and morphological markers tell us about

1086   plant hybridization. *Critical Reviews in Plant Sciences* **12**: 213-241. doi: 10.1080/713608045

1087 **Rieseberg LH, Sinervo B, Linder CR, Ungerer MC, and Arias DM. 1996.** Role of gene interactions in

1088   hybrid speciation: Evidence from ancient and experimental hybrids. *Science* **272**: 741-745. doi:

1089   10.1126/science.272.5262.741

1090 **Rieseberg LH, and Wendel JF. 1993.** Introgression and its Consequences in Plants. In: Harrison R, ed.

1091   *Hybrid Zones and the Evolutionary Process*. New York: Oxford University Press. 70-109.

1092 **Rieseberg LH, and Willis JH. 2007.** Plant speciation. *Science* **317**: 910-914.

1093 **Roberts RH. 1964.** *Mimulus* hybrids in Britain. *Watsonia* **6**: 70-75.

1094 **Robertson A, Rich TCG, Allen AM, Houston L, Roberts C, Bridle JR, Harris SA, and Hiscock, SJ. 2010.**

1095   Hybridization and polyploidy as drivers of continuing evolution and speciation in *Sorbus* L.

1096   (Rosaceae). *Molecular Ecology* **19**: 1675-1690.

1097 **Rosser EM. 1955.** A new British species of *Senecio*. *Watsonia* **3:** 228-232.

1098 **Ruhsam M, Hollingsworth P, and Ennos R. 2011.** Early evolution in a hybrid swarm between

1099   outcrossing and selfing lineages in *Geum*. *Heredity* **107**: 246-255.

1100 **Ruhsam M, Hollingsworth PM, and Ennos RA. 2013.** Patterns of mating, generation of diversity, and

1101   fitness of offspring in a *Geum* hybrid swarm. *Evolution* **67**: 2728-2740.

1102 **Schumer M, Rosenthal GG, and Andolfatto P. 2014.** How common is homoploid hybrid speciation?

1103   *Evolution* **68**: 1553-1560. doi: 10.1111/evo.12399

1104 **Scopece G, Musacchio A, Widmer A, and Cozzolino S. 2007.** Patterns of reproductive isolation in

1105   Mediterranean deceptive orchids. *Evolution* **61**: 2623-2642.

1106 **Scott RJ, Tratt JL, and Bolbol A. 2013.** Seed development in interploidy hybrids. In Chen ZJ and

1107   Birchler JA, eds. *Polyploid and Hybrid Genomics.* Oxford, UK: John Wiley & Sons. 271-290.

1108 **Sherry RA, Zhou X, Gu S, Arnone JA,3rd, Schimel DS, Verburg PS, Wallace LL, and Luo Y. 2007.**

1109   Divergence of reproductive phenology under climate warming. *Proceedings of the National*

1110   *Academy of Sciences, U. S. A.* **104**: 198-202.

1111 **Silvertown J. 2011.** *Demons in Eden: The Paradox of Plant Diversity.* Chicago, U.S.A.: University of

1112   Chicago Press.

1113 **Soltis DE, Soltis PS, Pires JC, Kovarik A, Tate JA, and Mavrodiev E. 2004.** Recent and recurrent

1114   polyploidy in *Tragopogon* (Asteraceae): Cytogenetic, genomic and genetic comparisons.

1115   *Biological Journal of the Linnean Society* **82**: 485-501.

1116 **Soltis PS, and Soltis DE. 2009.** The role of hybridization in plant speciation. *Annual Review of Plant*

1117   *Biology* **60:** 561-588.

1118 **Soltis DE, Visger CJ, and Soltis PS. 2014a.** The polyploidy revolution then...and now: Stebbins
1119   revisited. *American Journal of Botany.* **101**: 1057-1078. doi: 10.3732/ajb.1400178

1120 **Soltis PS, Liu X, Marchant DB, Visger CJ, and Soltis DE. 2014b.** Polyploidy and novelty: Gottlieb's
1121   legacy. *Philosophical Transactions of the Royal Society of London. B: Biological Sciences.* **369**:
1122   20130351. doi: 10.1098/rstb.2013.0351

1123 **Stace CA. 2010.** *New Flora of the British Isles.* Cambridge: Cambridge University Press.

1124 **Stace CA, and Crawley MJ. 2015.** *Alien Plants.* London, U.K.: William Collins.

1125 **Stace CA, Preston CD, and Pearman DA. 2015.** *Hybrid flora of the British Isles.* Bristol, U.K.: Botanical
1126   Society of Britain and Ireland.

1127 **Stathos A, and Fishman L. 2014.** Chromosomal rearrangements directly cause underdominant F1
1128   pollen sterility in *Mimulus lewisii–Mimulus cardinalis* hybrids. *Evolution* **68**: 3109-3119.

1129 **Stebbins GL. 1950.** *Variation and Evolution in Plants.* New York: Columbia University Press.

1130 **Stebbins GL. 1958.** The inviability, weakness, and sterility of interspecific hybrids. *Advances in*
1131   *Genetics Incorporating Molecular Genetic Medicine* **9**: 147-215. doi: 10.1016/S0065-
1132   2660(08)60162-5.

1133 **Stebbins GL. 1959.** The role of hybridization in evolution. *Proceedings of the American Philosophical*
1134   *Society* **103:** 231-251.

1135 **Stebbins GL. 1971.** *Chromosomal Evolution in Higher Plants.* London: Edward Arnold.

1136 **Straub SC, Parks M, Weitemier K, Fishbein M, Cronn RC, and Liston A. 2012.** Navigating the tip of
1137   the genomic iceberg: Next-generation sequencing for plant systematics. *American Journal of*
1138   *Botany.* **99**: 349-364. doi: 10.3732/ajb.1100335

1139 **Strong DR, and Ayres DR. 2013.** Ecological and evolutionary misadventures of *Spartina*. *Annual*
1140   *Review of Ecology, Evolution, and Systematics* **44**: 389-410.

1141 **Taylor E, Boughman J, Groenenboom M, Sniatynski M, Schluter D, and Gow J. 2006.** Speciation in
1142   reverse: morphological and genetic evidence of the collapse of a three-spined stickleback
1143   (*Gasterosteus aculeatus*) species pair. *Molecular Ecology* **15**: 343-355.

1144 **Taylor SA, Larson EL, and Harrison RG. 2015.** Hybrid zones: windows on climate change. *Trends in*
1145   *Ecology & Evolution* **30:** 198-406. doi: 10.1016/j.tree.2015.04.010

1146 **Taylor SJ, Arnold M, and Martin NH. 2009.** The genetic architecture of reproductive isolation in
1147   Louisiana irises: hybrid fitness in nature. *Evolution* **63**: 2581-2594.

1148 **Theodoridis S, Randin C, Broennimann O, Patsiou T, and Conti E. 2013.** Divergent and narrower
1149   climatic niches characterize polyploid species of European primroses in *Primula* sect*. Aleuritia*.
1150   *J. Biogeogr.* **40**: 1278-1289.

1151 **Thomas CD. 2015.** Rapid acceleration of plant speciation during the Anthropocene. *Trends in Ecology*
1152   *& Evolution* **30**: 448-455.

1153 **Twyford AD, and Ennos RA. 2012.** Next-generation hybridization and introgression. *Heredity* **108**:
1154   179-189. 10.1038/hdy.2011.68.

1155 **Ubsdell R. 1979.** Studies on variation and evolution in *Centaurium erythraea* Rafn and *C. littorale* (D.
1156   Turner) Gilmour in the British Isles, 3. Breeding systems, floral biology and general discussion.
1157   *Watsonia* **12**: 225-232.

1158 **Vallejo-Marín M. 2012.** *Mimulus peregrinus* (Phrymaceae): A new British allopolyploid species.
1159   *PhytoKeys* **14**: 1-14. 10.3897/phytokeys.14.3305.

1160 **Vallejo-Marín M, and Lye GC. 2013.** Hybridisation and genetic diversity in introduced *Mimulus*
1161   (Phrymaceae). *Heredity* **110**: 111-122.

1162 **Vallejo-Marín M, Buggs RJ, Cooley AM, and Puzey JR. 2015.** Speciation by genome duplication:
1163   Repeated origins and genomic composition of the recently formed allopolyploid species
1164   *Mimulus peregrinus*. *Evolution* **69**: 1487-1500.

1165 **Vallejo-Marín M, Cooley AM, Qi MLY, Folmer M, McKain MR, and Puzey JR. *in press*.** Strongly
1166   asymmetric hybridization barriers shape the origin of a new polyploid species and its hybrid
1167   ancestor. *American Journal of Botany.*

1168 **Vallejo-Marín M, Dorken ME, and Barrett SCH. 2010.** The ecological and evolutionary consequences
1169   of clonality for plant mating. *Annual Review of Ecology, Evolution, and Systematics* **41:** 193-213.

1170 **Vickery RK. 1959.** Barriers to gene exchange within *Mimulus guttatus* (Scrophulariaceae). *Evolution*
1171   **13**: 300-310.

1172 **Vilà M, Weber E, and Antonio CMD. 2000.** Conservation implications of invasion by plant
1173   hybridization. *Biological Invasions* **2**: 207-217.

1174 **Walther G, Roques A, Hulme PE, Sykes MT, Pysek P, Kuehn I, Zobel M, Bacher S, Botta-Dukat Z,**
1175   **Bugmann H, Czucz B, Dauber J, Hickler T, Jarosik V, Kenis M, Klotz S, Minchin D, Moora M,**
1176   **Nentwig W, Ott J, Panov VE, Reineking B, Robinet C, Semenchenko V, Solarz W, Thuiller W,**
1177   **Vila M, Vohland K, and Settele J. 2009.** Alien species in a warmer world: risks and
1178   opportunities. *Trends in Ecology and Evolution* **24**: 686-693. doi: 10.1016/j.tree.2009.06.008.

1179 **Weir J, and Ingram R. 1980.** Ray morphology and cytological investigations of *Senecio cambrensis*
1180   Rosser. *New Phytologist* **86:** 237-241.

1181 **Whitton J, Sears CJ, Baack EJ, and Otto SP. 2008.** The dynamic nature of apomixis in the
1182   angiosperms. *International Journal of Plant Sciences* **169**: 169-182.

1183 **Widmer A, Lexer C, and Cozzolino S. 2009.** Evolution of reproductive isolation in plants. *Heredity*
1184   **102**: 31-38.

1185   **Wolf DE, Takebayashi N, and Rieseberg LH. 2001.** Predicting the risk of extinction through

1186   hybridization. *Conservation Biology* **15**: 1039-1053. doi: 10.1046/j.1523-

1187   1739.2001.0150041039.x.

1188   **Yakimowski SB, and Rieseberg LH. 2014.** The role of homoploid hybridization in evolution: a century

1189   of studies synthesizing genetics and ecology. *American Journal of Botany* **101**: 1247-1258. doi:

1190   10.3732/ajb.1400201

1191   **Yeo P. 1968.** The evolutionary significance of the speciation of *Euphrasia* in Europe. *Evolution*: 736-

1192   747.

Figure Legends

Figure 1. Reproductive isolation barriers preventing the formation and establishment of hybrids.

Reproductive isolation barriers are usually divided in pre-zygotic and post-zygotic. Notice that in

plants, some barriers such as pollen precedence act after mating (pollination) but before zygote

formation.

Figure 2. Meiosis in hybrids is adversely affected by differences in chromosome number and

structure between parental species. The result will generally be gamete inviability, but, rarely,

gametes with atypical chromosome numbers (aneuploid and unreduced gametes) may be produced

and viable. The figure shows ways in which viable gametes could be produced in a hypothetical

triploid hybrid (2n = 3x). Unpaired chromosomes during meiosis (in both univalents and

multivalents) migrate to either cell pole at random, resulting in unbalanced gametes (top pathway).

Most of these unbalanced gametes will be inviable, causing a very severe reduction in fertility. A

small fraction of these gametes may by chance end up with the parental cytotype, and are thus

viable. Another small fraction may be viable despite carrying new aneuploid combinations.

Occasionally, gametes may be produced with the entire set of chromosomes present in the hybrid

(unreduced gametes), which may be subsequently involved in polyploid formation.

Figure 3. Potential fate of viable hybrids that are able to persist over ecological or evolutionary

timescales after their origin and initial establishment.

Table 1. Examples of the "Significant Eight" alien hybrid complexes of the British Flora recognised by Stace & Crawley (2015). Introduced taxa (neophytes) are indicated with *. Data compiled by Stace & Crawley (2015) and Stace *et al.* (2015).

Hybrid	Parents		Sexual Fertility	Clonal reproduction	Allopolyploid derivative
Fallopia x bohemica (2n = 66)	*F. japonica** (2n = 88)	F. sachalinensis* (2n = 44)	Low	Yes	--
Senecio x baxteri (2n = 30)	*S. vulgaris* (2n = 40)	*S. squalidus** (2n = 20)	Very low	No	*S. cambrensis* (2n = 60)
Spartina x townsendii (2n = 62)	*S. maritima* (2n = 60)	*S. alterniflora** (2n = 62)	Very low	Yes	*S. anglica* (2n = 124)
Mimulus x robertsii (2n = 44-46)	*M. guttatus** (2n = 28)	*M. luteus** (2n = 60-62)	Very low	Yes	*M. peregrinus* (2n = 92)
Calystegia x lucana (2n = 22)	*C. sepium* (2n = 22)	*C. sylvatica** (2n = 22)	High	Yes	--
Hyacinthoides x massartiana (2n = 16, 24)	*H. non-scripta* (2n = 16, 24)	*H. hispanica** (2n = 16, 24)	High	Yes	--
Rhododendron x superponticum (2n = 26)	*R. ponticum** (2n = 26)	*R. catawbiense*, R. maximum*, R. macrophyllum** (2n = 26)	High	Yes	--

Centaurea x gerstlaueri **(2n = 44)**	*C. nigra* (2n = 44)	*C. jaceae** (2n = 44)		High?	Yes (but limited lateral spread)	--

Table 2. Reproductive mode and occurrence of 274 alien angiosperm hybrids and 1,590 non-hybrid aliens in the British and Irish flora. Data from Stace and Crawley (2015). Hectad = 10km x 10km square.

Propagation Mode	# Hybrid alien taxa (proportion)	Average # hectads per hybrid taxon (maximum)	# Non-hybrid alien taxa (proportion)	Average # hectads per non-hybrid taxon (maximum)
Seeds only	48 (0.175)	180 (2,242)	559 (0.351)	224 (3,530)
Clonal only	55 (0.200)	157 (1,147)	122 (0.077)	212 (2,723)
Seeds and clonal	71 (0.259)	199 (2,419)	330 (0.207)	218 (2,535)
No reproduction in the British isles	100 (0.365)	32 (444)	579 (0.364)	37 (1,738)
Total	**274**	**126 (2,419)**	**1,590**	**153 (3,530)**

Box 1. The Avon Gorge: A hotspot of hybrid diversity

The genus *Sorbus* (Rosaceae) includes sexual diploid species and apomictic species/taxa with varying ploidy (usually 3x and 4x) that have arisen through hybridisation between sexual species and rare backcrosses with their apomictic allopolyploid derivatives (Rich & Robertson 2015). There are 31 *Sorbus* agamospecies native to the British Isles, 10 of which are reported in *Stace et al.*, (2015). The parentage of these hybrids is often complex but molecular data is helping to resolve their origins (Robertson *et al.*, 2010). 'Hotspots' for *Sorbus* diversity in the British Isles include: the Isle of Arran, Cheddar Gorge, the Wye Valley and the Avon Gorge (Rich & Robertson 2015) – the latter being by far the richest, possibly the richest in the world.

The Avon Gorge contains at least 21 *Sorbus* taxa, six of which are endemic and shown to have evolved within the Gorge, probably within the last 10,000 years since the last ice-age (Robertson *et al.*, 2010; Ludwig *et al.*, 2013). Studies using molecular markers revealed that this *Sorbus* diversification has been driven primarily by a series of interspecific hybridisations and backcrosses among closely related taxa, with each new genotype being fixed and perpetuated via apomixis (Robertson *et al.*, 2010; Ludwig *et al.*, 2013). Once established these new microspecies then occasionally participate in further rare hybridisation events leading to a complex pattern of ongoing reticulate evolution (Robertson et al. 2010; Ludwig *et al.*, 2013). This occurs because of subtle variation ('leakiness') in the apomictic mating system that permits occasional sexual unions, usually involving pollen from the most common species, *S. aria*, which is diploid and sexual (outcrossing) and its close tetraploid relative *S. porrigentiformis*, apomictic. Pollen from these taxa is essential for apomictic seed production by the triploid apomictic hybrid taxa (*e.g.*, *S. bristoliensis* and *S. wilmottiana*) because apomixis is pseudogamous (endosperm formation requires fertilization of the central cell by sperm) and triploids (unlike tetraploids) are self-incompatible (SI) (Ludwig *et al.*, 2013; Ludwig & Hiscock in preparation). This unusual mating system therefore fuels the possibility of rare hybridisations being successful because interspecific pollination is required for apomictic seed production in the triploids, which in all but one microspecies tested, are genetically identical (indicating a single hybrid origin event) and therefore incompatible because they all share the same self-incompatibility (S) genotype (Ludwig *et al.*, 2013). Populations of at least two undescribed microspecies have been identified together with numerous puzzling individuals, which do not fit known taxa (Tim Rich pers. comm.) indicating that these evolutionary processes are ongoing. The 2km long Avon Gorge which cuts through Carboniferous Limestone to a depth of 80m, displays a great diversity of habitats (including woodland, scrub, open rocks and quarries) which has probably facilitated and continues to facilitate the survival of newly divergent (perhaps pre-adapted) *Sorbus* taxa (Rich *et al.*, 2010). Conservation strategies for the rare *Sorbus* taxa endemic to the Avon Gorge have taken these findings on board in revising management regimes such that all *Sorbus* taxa are considered so as to conserve the evolutionary process rather than just the rare ones.

Box 2. Monkeyflower hybrids: The birth of a new species

Mimulus has a long tradition in studies of hybridisation, reproductive isolation and speciation (Vickery 1959, Bradshaw *et al.,* 1995, Fishman & Willis 2001, Brandvain *et al.,* 2014). The spread of some *Mimulus* species beyond their native range thanks to human-assisted dispersal, has created new opportunities for hybridisation between previously isolated *Mimulus* taxa (Vallejo-Marín & Lye 2013).

The best studied case is the invasion of the British Isles by two related monkeyflower taxa: The North American diploid *M. guttatus*, and the South American tetraploid *M. luteus*. Both species were introduced into the British Isles in the early 19th century as botanical curiosities, and quickly became naturalised (Stace *et al.,* 2015). Hybridisation between these two taxa produce a highly sterile triploid, which is nevertheless capable of vegetative growth. The hybrid has been established in the wild since at least the 1870's, and it is currently the second most abundant *Mimulus* in the UK after *M. guttatus* being present in approximately 40% of extant populations (Vallejo-Marín & Lye 2013). The triploid hybrid *M. x robertsii*, has produced the new allopolyploid species *M. peregrinus*, which has originated at least twice, independently, in Scotland (Vallejo-Marín 2012, Vallejo-Marín *et al.,* 2015). *M. peregrinus* produces both viable pollen and seeds, as well as retaining its capacity for vegetative reproduction. Multiple origins seem to be the rule rather than the exception for the formation of allopolyploids (Soltis *et al.,* 2014b), and this provides a natural system to investigate hybrid speciation in a replicated fashion.

Although the parentage of *Mimulus* hybrids and the origin of *M. peregrinus* has been recently documented using genome-wide analyses (Vallejo-Marín *et al.,* 2015), a missing piece of the puzzle is to determine which taxon has served as the maternal parent and which as the paternal parent in hybrid formation. Inter-ploidy hybridisation often results in asymmetric reproductive barriers, and it is likely that hybridisation between *M. guttatus* and *M. luteus* does not work equally well in both directions (Roberts 1964). Determining the ancestry of inter-specific hybrids can be done by analysing uniparentally inherited genomes (e.g., chloroplast and mitochondria) (Rieseberg & Brunsfeld 1992, Twyford & Ennos 2012). However, genetic analysis of hybrid ancestry can be complicated by limited availability of species-diagnostic polymorphism in uniparentally-inherited genomes. Recent studies have pointed out how whole genomes of cytoplasmic organelles are recovered in both targeted and whole-genome sequence projects, as by-products of even low-depth sequencing efforts (genome skimming, Dodsworth 2015). This trove of genetic information can be used to identify the taxon that acted as the maternal parent of hybrids, even in the absence of closely related reference genomes (Bakker *et al.,* 2016). Vallejo-Marín *et al.*, (*in press*) applied a genome skimming approach to determine the ancestry of *Mimulus* hybrids in the British Isles. They used data obtained from both whole-genome and targeted sequencing projects, to rescue chloroplast and mitochondrial genomes of *M. x robertsii* and *M. peregrinus* and their parental taxa. They showed that hybrids have been produced unidirectionally, with *M. guttatus* as the maternal and *M. luteus* as the paternal parent. Genome skimming (Straub *et al.,* 2012) holds great potential to investigate the ancestry of neo-hybrids, and we expect similar approaches to be increasingly exploited in the near future.