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1	(Comparative evidence supports a role for reproductive allocation in the
2		evolution of female ornament diversity
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10		
11	Abstr	act
12	1.	Sexually selected ornaments are highly variable, even among closely related
13		species, and the ultimate causes of variation in ornament evolution are unclear,
14		including in rare cases of female ornament expression. One hypothesis is that
15		differences across species in female reproductive allocation may help explain
16		patterns of female ornament expression among insects with nuptial gifts.
17		
18	2.	Dance flies (Diptera: Empididae: Empidinae) vary considerably among species
19		in the presence and extravagance of female ornaments, which probably evolved
20		through female contests for mates. In most dance flies, adult females appear to
21		acquire all their dietary protein from nuptial gifts provided by males during
22		mating. The importance of nuptial feeding on egg development is not yet
23		known.
24		
25	3.	To test the prediction that the presence of female ornaments reflects differences
26		in the degree to which females rely on nuptial feeding for egg development, we

27	examined egg development in wild females of two species, one ornamented
28	and the other unornamented. We validated an ageing technique based on
29	cuticular bands which permitted a regression of egg size on adult age.
30	
31	4. We found that egg development depended on mating status in the ornamented
32	species alone, meaning the eggs of unmated females of the ornamented species
33	did not develop. This contrast across species is consistent with expectations that
34	females of different species vary in their dependence on nuptial gifts for egg
35	development.
36 37	5. Our findings provide preliminary support for the hypothesis that differences in
38	reproductive allocation mediate the intensity of female contests for nuptial gifts.
39	
40	Key words - courtship feeding, female ornaments, mate choice, reproductive
41	allocation, anautogeny, sexual competition
42	
43	Introduction
44	In some unusual mating systems, female fitness is limited by male monopolization of
45	resources required for reproduction, and females may consequently compete for mates.
46	In such mating systems, females can evolve secondary sexual traits if the advantages of

winning contests for mates is sufficiently large (Clutton-Brock 2009; Gwynne and
Simmons 1990; Herridge et al., 2016). However, while sexual selection and ornament

49 expression are common in males of many taxa (Janicke et al., 2016), even when females
50 experience strong sexual selection they rarely have extravagant ornaments (Amundsen
51 2000). One possible explanation for this disparity is that female fitness tends to be
52 resource-limited to a greater degree than male fitness; by definition, females must make

53 large investments in eggs, which might trade-off with any investment in ornamentation

54 (Fitzpatrick et al., 1995). Male choice for adorned females is probably also constrained 55 by trade-offs between ornaments and offspring, as males should prefer mates who 56 invest in offspring rather than ornaments. Moreover, when females store sperm, 57 attractive females may actually present a higher risk of sperm competition, such that 58 males might avoid rather than prefer showy females (Herridge et al., 2016). Together, 59 these arguments make rare species with male choice for showy female ornaments 60 perplexing, and good candidates for testing theories about what regulates interspecific 61 diversity in male choice and ornament expression.

62

63 Male insects often provide nutrition to females during courtship in the form of "nuptial 64 gifts", and these material donations are commonly used to initiate or accelerate egg 65 production (Lewis et al., 2014). The degree to which females rely on nuptial gifts for 66 egg production should covary with the sexual receptivity of females, because hungry 67 females might use sex as a foraging technique. An increase in sexual receptivity can in 68 turn lead to increased competition among females, especially if the preferred mating 69 rate of females begins to exceed the rate at which males can provide gifts (Arnqvist and 70 Nilsson 2000; Simmons and Gwynne 1993). Increased sexual selection on females 71 arising from this enhanced competition could under some circumstances lead to the 72 evolution of extravagant traits that improve female attractiveness and therefore female 73 access to limiting nuptial gift nutrients.

74

While the role of nuptial gifts in promoting female contests and ornament evolution is relatively uncontroversial, the extravagance of ornaments often varies even among closely related species that share the same geographic distribution and mating behavior, which remains unexplained (Cumming 1994; Downes 1970). Cumming (1994) has hypothesized that the presence and level of expression of female ornaments may depend 80 on the intensity of female competition for nuptial gifts, which in turn might be 81 determined by the degree to which females rely on male gifts for egg development. The 82 allocation of resources to eggs prior to mating may therefore mediate the intensity of 83 female competition for nuptial gifts, influencing the strength of selection on female 84 investment into ornamental traits.

85

86 Female insects can allocate resources obtained during both larval and adult stages to 87 reproduction, and the timing of acquisition and source of resources have important 88 consequences on the reproductive and foraging strategies of animals (Boggs 1997a). 89 Exclusively using larval-derived resources to invest into egg development is termed 90 autogeny, and is common among Ephemeroptera and in many species of Lepidoptera 91 and Diptera (Engelmann 1970). By contrast, anautogeny describes the condition in 92 which females require some adult nutrients to mature eggs, e.g., as in mosquitoes that 93 act as vectors for human diseases including malaria. The level of dependence on adult 94 derived resources for egg production varies continuously across anautogenous species 95 (Boggs 1981; Jervis 2012). While variation in anautogeny remains largely unexplained, 96 the availability of resources at different stages in insect life cycles likely strongly 97 influences the pattern of resource partitioning, and therefore resources available for 98 reproductive allocation (Jervis et al., 2001). Nuptial gifts, being often of substantial 99 size, may influence the partitioning of resources towards growth and reproduction in 100 life stages prior to adult stages. This may occur because of female expectation of nuptial 101 resources in the adult stage. However, to our knowledge the extent to which selection 102 arising from the presence of nuptial gifts affects resource partitioning has not been 103 investigated.

By definition anautogenous species should have females that depend more on adult derived resources for egg production when compared to others. In these species, therefore, nuptial gifts (should they be present) may represent essential resources for female reproductive success (Fritzsche et al., 2016). As such females of anautogenous species may experience greater competition for mating in order to acquire nuptial gifts, and subsequently have elevated selection for sexual trait investment (Cumming 1994).

111

One group of taxa with nuptial gift giving which exhibits extraordinary interspecific 112 113 diversity in the extravagance of female ornaments are the dance flies (Diptera; 114 Empididae; Empidinae) (Collin 1961; Cumming 1994; Downes 1970). Despite closely 115 related species having similar courtship behaviour, females of different species display 116 varying levels of ornamentation (Funk and Tallamy 2000; Murray 2015). Ornamental 117 traits include pinnate leg scales, enlarged and or darkened wings, and inflatable 118 abdominal sacs, although their presence and extravagance varies substantially among 119 taxa (Collin 1961; Downes 1970). Within species variation in ornament size is known 120 to influence male mate choice (Funk and Tallamy 2002: Murray et al 2018). Ornaments 121 appear to have evolved independently multiple times across the dance fly phylogeny 122 (Murray 2015). We do not yet know what regulates these many evolutionary transitions 123 in ornament expression across dance flies, but we expect higher levels of ornament 124 expression in species subject to more intense sexual selection on females. Therefore, it 125 is reasonable that the fitness of females in species with substantial sexual ornaments is 126 more strongly constrained by dietary protein than unadorned species. In such species, 127 investment in ornaments may be justified because the returns on investment in 128 ornaments (through the accrual of nuptial gifts) more than offset the cost of construction 129 (Fitzpatrick et al., 1995).

In roughly a third of dance fly species of the subfamily Empidinae, females are more numerous than males in lek-like mating swarms (Cumming 1994; Downes 1970). Males approach swarms with prey items as nuptial gifts, typically another dipteran (Cumming 1994). Males typically assess females from below, apparently evaluating the gravidity of females (Funk and Tallamy 2000). In most species, females do not appear to hunt as adults, and therefore derive all of their dietary protein from nuptial gifts.

138

The strong link between mating and foraging in dance flies suggest that egg development may be copulation-dependent (Cumming, 1994), but this hypothesis has never been rigorously tested. One limitation has been an inability to rear dance flies in the lab, and thereby study ovarian physiology in individuals of known age and mating history. Additionally nuptial gifts vary in size and quality, and therefore females may not receive the same volume of resources in each mating (Svensson et al., 1990), which may cause fitness received by mating to depend on nuptial gift characteristics.

146

147 In this study we circumvent the inability to rear and manipulate flies by predicting that 148 mating status (rather than mate number) covaries with egg development in ornamented 149 and non-ornamented taxa. It is possible to distinguish mated from non-mated wild 150 females by inspecting their sperm storage organs for the presence of sperm, and 151 therefore to know whether a female has or has not received adult dietary protein, in the 152 form of at least one nuptial gift. In order to assess the temporal dimension of ovarian 153 maturation, we first adopt and validate an ageing method for wild caught dance flies. 154 We compare two species that are locally abundant for long periods near our University 155 in central Scotland, facilitating the collection of mated and non-mated individuals at a 156 range of different ages. The two species are *Empis aestiva*, which has females with

extensive pennate scales on their mid and hind legs (see figure 1), and *Rhamphomyia crassirostris* with no obvious sexual ornaments on either sex (see figure 1). In both species, males obligately provide nutritious nuptial gifts to females (males are not known to provide non-nutritious "sham" gifts in these species). We predicted that when unmated, the eggs ornamented *E. aestiva* females would develop at a slower rate than the eggs of unadorned *R. crassirostris* females, whereas mated females with access to dietary protein might show no such differences.

164

165 Materials and methods

166 Aim 1: Validating methods for ageing wild flies

167 The thoracic apodemes of flies are known to continue growing even after eclosion, 168 leaving evidence of time passed since eclosion (Schlein and Gratz 1973). A distinct line 169 marks the extent of the apodeme structure upon eclosion (see supplementary Figure 1). 170 After eclosion, the density of the cuticle deposited at the cortical part of apodemes is 171 influenced by temperature. Diurnal temperature fluctuations cause banding to occur and 172 therefore theoretically the number of apodeme bands covaries with the number of days 173 since eclosion (Johnston and Ellison 1982; Schlein and Gratz 1973). However, it is 174 likely that cuticle is not laid down on the apodemes indefinitely, and that the age at 175 which the apodemes cease to grow varies among species and even individuals. Using 176 apodeme bands to age flies has been validated for a number of Dipteran species, but 177 not previously for any empid species (Neville 1983).

178

179 In order to test whether our focal species produce apodeme bands that reliably reflect 180 adult age, individuals of known age were required. We were unable to collect a 181 sufficient number of the two focal species (the precise habitats of larval empids remain 182 unclear, and our sampling was unable to improve this knowledge, but it is likely that

larval empids are relatively well dispersed since they are thought to be predators). We
therefore determine whether the thoracic apodemes of dance flies accumulate daily
cortical growth bands using a range of Empidinae species caught in emergence traps.
We also assessed which particular regions of the thoracic apodemes have the most
distinguishable bands.

188

189 To ensure that the flies caught for this experiment were of known age, we used 190 emergence traps (ground area 1 m. X 2 m., height 1 m.). The traps were deployed from 191 the 13th of June till the 20th of July 2016 at three locations in central Scotland, UK: 192 Stirling University campus, (56°14'81.10"N, 003°90'52.02"W), Kippenross forest, 193 (56°17'04.07"N, 003°95'52.56"W) and a field near Enochdhu, (56°74'20.21"N, 194 003°52'21.00"W). The traps were searched twice per day, once before 12pm and again 195 after 6pm, which ensured that any flies found in the traps had emerged within the last 196 24 hours. We aimed to collect as many flies as could be caught in the traps during the 197 period. Flies in the dance fly sub-family Empidinae were retained and kept alive 198 between zero and ten days, as experimentally determined after capture using a random 199 number generator (pilot experiments suggested that individuals were unlikely to live in 200 captivity beyond ten days). The captive flies were placed in individual plastic containers 201 closed with cotton stoppers. Each container had two cuvettes placed inside, one with 202 cotton wool soaked in water and another with sugar granules. The containers were kept 203 sheltered, but out-of-doors so that the flies would experience natural diurnal 204 temperature fluctuations, which are critical for the formation of the thoracic bands 205 (Schlein and Gratz 1973; Johnston and Ellison 1982; Neville 1983). On the appointed day of sacrificing, the flies were killed by freezing ($C^{\circ} \leq -12$). 206

208 The method used to dissect flies and count apodeme bands was modified based on the 209 protocol published in Schlein and Gratz (1973). The head, abdomen, wings and legs of 210 the flies were removed and the thorax bisected. The bisected thorax halves were placed 211 in distilled water and, using a dissecting microscope (Lecia MZ12) and titanium 212 forceps, the anterior and posterior thoracic apodemes extracted. The majority of the 213 muscle tissue was removed from the apodemes with forceps, and the rest dissolved in 214 potassium hydroxide (10%) for 150 minutes. The specimen was then rinsed in distilled 215 water, stained in potassium permanganate (10%) for two minutes and rinsed again in distilled water. The anterior and posterior apodemes were bisected mid-sagitally with a 216 217 razor blade to form four fragments. The two fragments of the anterior apodeme were 218 bisected transversely into rostral and caudal halves. The resulting six fragments (two 219 from the posterior and four from the anterior apodemes) were then mounted in DPX 220 medium (Fisher Scientific, Loughborough, UK) on a microscope slide.

221

Taking care not to count the eclosion line, the bands on all six fragments of apodeme were counted using a light microscope (Olympus BX-41; see supplementary figure S1 for the locations of bands on the three apodeme fragments). To prevent bias, we counted blind to the knowledge of how many days the fly had been kept alive. Recounting to quantify repeatability using intra-class correlation was performed blind to both the real adult age and the previous count.

228

Statistics were performed in R version 3.3.2 (R Core Team 2016). To validate the age estimates obtained from apodeme bands, we used major axis (model 2) linear modeling to regress the maximum number of bands counted from any apodeme per fly against known age. Major axis regression was used because we recognize the possible error in both the predictor and response, and because least squares regression is known to

234 negatively bias slope estimates in such cases. Because bands are often hard to 235 distinguish, and their formation is unlikely to be as regular as theorized (especially in 236 Scotland where daily temperature fluctuations are often modest), we minimized the 237 resulting bias by using maximum band number instead of the average across apodemes. 238 We used age as the y-axis in this case because we experimentally manipulated the age 239 of these flies, having collected them on the day of eclosion; therefore the band number 240 was the response variable of interest even though our motivation is to assess if band 241 number reflects age. Models were validated by visually assessing diagnostic plots to 242 confirm normality of residuals and homoscedascity, and to ensure that no records had 243 unduly high influence. We used the ICC package for R to obtain repeatabilities (Wolak 244 et al., 2012).

245

Aim 2: The effect of age and mating on ovarian maturation

247 In order to investigate the relationship between ovarian development and female 248 resource allocation, we collected data on egg area (as a proxy for the stage of 249 vitellogenesis), mating status (mated or unmated, based on the absence or presence of 250 sperm in the spermatheca) and estimates of age using thoracic apodeme counts from 251 wild females of unknown age captured in mating swarms or on vegetation using hand-252 held nets. We aimed to collect 30 unmated and mated females of both species. Females 253 of R. crassirostris were caught from May to July in 2016 in central Scotland, on the 254 University of Stirling campus, (56°14'88.01"N, 003°90'56.02"W,) and in Kippenross 255 forest, (56°17'01.50"N, 003°95'51.22"W). Females of *E. aestiva* were caught in 2015 256 in central Scotland, near the Scottish Centre for Ecology and Natural Environment 257 (SCENE) (56°09'06.56"N, 004°38'36.16"W), and in 2016 in a forest near Enochdhu, 258 (56°74'20.21"N, 003°52'21.00"W). The flies were all frozen immediately on the day 259 of capture ($C^{\circ} \leq -12$).

Flies were dissected using a Leica MZ12 light microscope and titanium forceps. The

abdomen was removed and placed in distilled water and opened. We photographed between three and five of the largest eggs (dance flies mature eggs in clutches, such that within female variation in egg size is negligible, pers. obs.) using a microscope mounted camera (Olympus SP-500UZ) and measured egg area using ImageJ version 1.51h (Schindelin et al., 2015).

267

Like most other insects, female dance flies are able to store sperm, and eggs are 268 269 fertilized only at the time of oviposition. We checked the mating status of females by 270 splitting the spermatheca with forceps and visually assessing the presence of sperm 271 (females captured while mating invariably contain sperm, suggesting that failures to 272 transfer ejaculate are rare in these taxa, pers. obs.). Since females in the two species 273 being investigated have never been seen mating without a nuptial gift (pers. obs.), by 274 comparing ovarian development in mated and unmated females, we are able to 275 distinguish the development of eggs in females with and without access to adult dietary 276 protein. The thoracic apodemes of the species were dissected and stained using the 277 method outlined above. We used the maximum number of bands counted on any of the 278 apodeme types as our best estimate of age.

279

To test whether the two species of dance fly differ in the effect of mating status on egg development, we fit a multiple regression model including a three-way interaction between age (in days), mating status (mated or unmated as a categorical variable) and species (*E. aestiva* or *R. crassirostris*). The two species differed substantially in the size of mature eggs, so average egg sizes were standardized independently for each species, which facilitates interpretability of model coefficients (Schielzeth 2010) by providing

a strong test of whether the interaction between mating status, species and age represents a difference in the rate of maturation, rather than merely reflecting the different egg sizes across species. We assessed model quality by visually inspecting diagnostic plots. In order to achieve homoscedascity and improve fit, we used a naturallog-transformed average egg size for each female in our model.

291

292 Results

Among flies caught in emergence traps (see table 1) and reared in captivity for up to ten days, the number of bands counted on the rostral and caudal parts of their anterior apodemes ranged from zero to eight and zero to five, respectively. For the posterior apodemes the number of bands ranged from zero to nine.

297

The repeatabilities in table 2 show that estimates of age are most consistent across blind trials for the maximum band number, followed by the rostral fraction of anterior apodemes, then posterior apodemes and finally the caudal part of the anterior apodemes. We therefore estimated the age of field-caught dance flies using the maximum number of bands counted on any apodeme. This subset of counts produced an age predicting model with the lowest AIC and highest R squared when compared to models using counts from only one apodeme type (see figure 2).

305

306 As predicted, we found a strongly significant positive association between maximum

307 band number and fly age (Intercept = 0.7478, 95% CI (lower = 0.0538, upper = 1.3341),

308 Slope = 0.7788, 95% CI (lower = 0.627, upper = 0.9562), p-value <0.0001, R² adjusted

309 = 0.7485, see figure 2).

310

The age estimates for *R. crassirostris* females ranged from zero to ten days, with a broadly Gaussian distribution notwithstanding the strict bound at zero, see figure 3. Nineteen of the 69 females were unmated, and average egg sizes ranged from 0.0092mm^2 to 0.1305mm^2 , with mean egg size at $0.0500 \pm \text{SE} 0.0036$. The age estimates for *E. aestiva* ranged from zero to five days; see figure 3. Eighteen of 49 females were unmated. Average egg sizes ranged from 0.0068mm^2 to 0.0523mm^2 , with mean egg size = 0.0181 ± 0.0014 .

318

The effect of mating status on egg development differed between the two dance fly species. In the unadorned *R. crassirostris*, eggs increased gradually in size for both mated and unmated females (although the intercept for unmated females was lower, as might be expected if male nuptial gifts boosted vitellogenesis). In contrast and as predicted, unmated *E. aestiva* demonstrated no change in egg size with age (Partial F test of whether the removal of the 3-way interaction term results in a poorer model fit: F = 4.1919, P = 0.0430; see Figure 4).

326

327 Discussion

328 We examined egg development in mated and unmated dance flies from two species that 329 contrast sharply in female ornamentation, and found evidence consistent with an 330 association between anautogeny and ornamentation, supporting the hypothesis that 331 interspecific differences in dance fly sexual ornament expression derive at least in part 332 from differences in ovarian physiology. Mindful of the limitations on inference that are 333 inherent with two-species comparisons, we discuss the implications of our findings for 334 mating system diversity in this group and female ornament expression in general, as 335 well as the validated ageing method that we modified from previous work.

337 How mate acquisition relates to investment in ornaments

338 Sexually selected ornaments are highly variable, even among some closely related 339 species (Murray 2015). Ornaments evolve as a consequence of competition for mates, 340 but the underlying causes of competition are unclear in many cases. Female ornaments 341 are particularly curious adaptations, because general conditions that favour their 342 evolution appear to be exceedingly rare (Clutton-Brock 2009). In spite of this rarity, 343 some taxa like the dance flies possess extravagant variation in female ornaments, which 344 challenges our understanding of the general rules that are thought to regulate diversity 345 in mating systems (Janicke et al 2016).

346

347 We tested Cumming's (1994) prediction that interspecific variation in how and when 348 female dance flies allocated resources to eggs plays a key role in mediating the intensity 349 of female contests for mates. Although we did not directly manipulate allocation (such 350 experiments remain impossible given the current state of knowledge for culturing dance 351 flies), we nevertheless generated a priori predictions for patterns of ovarian maturation 352 in females of two different species that differed in ornament expression. We found that, 353 as predicted, the eggs of unmated females of the ornamented species developed at much 354 slower rates compared to the unornamented species. This pattern suggests that females 355 of the ornamented species rely to a greater degree on male nuptial gifts than females 356 from unadorned species (see table 3), which provides long-awaited support for 357 Cumming's hypothesis, and can help explain the otherwise confounding diversity of 358 sexual trait expression among dance flies (Herridge et al., 2016: Houslay and Bussière 359 2012).

360

In light of the observational nature of our work, there are several factors in addition tothe hypothesized mechanism which could be at play. For example, we cannot rule out

363 that females may receive other nutrition or simulants other than prey item nuptial gifts 364 from mating, as is common in other insect species, which may also influence the 365 development of eggs (Lewis et al., 2014). However, even if this were true it would not 366 explain the difference in ovarian maturation seen among unmated females in the two 367 focal species. Likewise, our estimates of egg size, mating status and age were all 368 measured with error. While these sources of error do not lead to systematic biases in 369 our estimates, they nevertheless make our central conclusions tentative, especially in 370 light of the marginal significance of our key result. More information on how these 371 errors contribute to patterns in the data would be welcome. For example, we did not 372 know the number of times a female had mated, and the volume of resources acquired 373 by the female during each mating. Some of these matings may have been too recent for 374 the effective conversion of nuptial gifts to egg maturation, and others may have resulted 375 in minimal resource intake. More detailed information on mating history rather than mere mating status (mated or unmated) would clearly provide more resolution for this 376 377 kind of analysis.

378

379 Another possibility is that species differences in mating system caused differences in 380 the representativeness of subsamples of unmated females. For example, if male choice 381 is stronger in the ornamented *E. aestiva*, it might lead to a stronger difference in average 382 condition between the mated and unmated fractions of females. The fact that low-383 condition females in that species are less likely to mature eggs over time could therefore 384 be due to their lower condition rather than to species differences in reproductive 385 allocation. Although we cannot rule out this possible alternative explanation, it is worth 386 noting that the proportions of unmated females were similar in the two species, 387 suggesting no large difference in the chances of mating (19/69 or 27.5% of females for

- *R. crassirostris*, compared to 18/49 or 36% for *E. aestiva*). It seems unlikely that this
 small difference could explain our observed differences in ovarian physiology.
- 390

Another intriguing contrast across species involved the difference in female age range, as estimated by apodeme bands. *E. aestiva* ages ranged from zero to five days and *R. crassirostris*, zero to ten days. While these differences in apparent age structure are intriguing and unexplained, they may simply reflect differences across species in the deposition of cuticular bands. Once again, we think this is unlikely, because if many individuals were older than the maximum number of bands possible, we would expect a left-skewed distribution of apparent ages, which we did not observe in Figure 3.

399 In species with relatively brief adult life span, it is probably important for females to 400 mate quickly in order to produce eggs and oviposit. Selection for females to mate 401 quickly after emerging may act as an important factor leading to the expression of 402 ornaments. In the apparently longer-lived R. crassirostris, females may be able to 403 remain virgins for a longer period. Comprehensively disentangling the possible causes 404 of species differences in sexual receptivity requires more information on the individual 405 species in question as well as more comparative work on further species. Herridge 406 (2016) found significant differences in the number of matings obtained by females of 407 three nuptial gift giving species of dance fly, including our study species E. aestiva. 408 Interestingly mate number in these species did not straightforwardly reflect ornament 409 expression. While the most ornamented species (R. longicauda) does appear to have 410 the most sexually receptive females, female *E. aestiva* appeared to mate less often than 411 in a completely unornamented species (E. tessellata). The longevity of R. longicauda 412 and *E. tesselata* are unknown, but it is possible that the combination of female size,

413 longevity and reproductive allocation all affect sexual receptivity and help further414 explain cross-species differences in female ornament expression.

415

Inferring adaptive patterns from two-species comparisons is difficult (Garland and Adolf 1994). We recognize that our study would benefit from comparing additional species, however the collection of sufficient numbers of mated and unmated females of even the two species used here was difficult, despite the fact that these species are abundant, and we know a reasonable amount about both their mating swarm locations and habitat use. Although we remain cautious in our conclusions, we agree with Cooper (1999) that two species comparisons help identify promising avenues of research.

423

Interspecific variation in anautogeny remains largely unexplained (Jervis et al., 2001). However resource allocation to reproduction is expected to relate to the nutritional ecology of the adult (Jervis et al., 2001). The quantity, quality and predictability of resources available to the adult in the environment are factors that likely influence the evolution of reproductive allocation across life history stages (Karlsson 1994). This fact supports the notion that nuptial gifts are in a unique position to influence both sexual selection and life history (Lewis et al., 2014).

431

432 Validation of an ageing method for wild dance flies

A technique for ageing wild subjects is one of the prerequisites for measuring egg development rates in dance flies, which have heretofore been difficult to culture under lab conditions. We have demonstrated that using thoracic apodeme bands works at least as well for the Dipteran subfamily Empidinae as it has previously been shown for other groups of flies (Neville 1982), in spite of the relatively slight daily fluctuations in temperature that are routine in Scottish summers. We hope this demonstration will 439 stimulate more investigations of an ecological and evolutionary nature on the 440 demographic patterns and life history traits of wild flies, which remain heavily reliant 441 on lab studies with few exceptions (see Bonduriansky and Brassil 2002 for an 442 exception).

443

444 In previous studies that evaluated the accuracy of apodeme bands to age other species 445 of fly, flies were kept in controlled temperature regimes (Neville 1983); hence flies 446 experienced constant temperature fluctuation between day and night for an extended 447 period of time, which likely influenced how distinguishable the bands were (Johnson 448 and Ellison 1982; Schlein and Gratz 1972). In our study, recently emerged flies were 449 kept outside and experienced natural diurnal fluctuations in temperature. Although it 450 was difficult to count bands in some specimens, which inflated uncertainty around age 451 estimates, our estimates were nevertheless consistently close to the ages of known-age 452 specimens (see figure 2). Regardless of the exact linear modeling approach (ordinary 453 least squares, major axis, or forcing the intercept through the origin) our slope estimate 454 was consistently more than one, indicating that band numbers are a good proxy for age 455 but are likely to systematically underestimate it. This is not surprising since it is easier 456 to imagine environmental conditions that obscure bands than those that might create 457 additional bands.

458

Because of an inability to collect large numbers of individual species in emergence traps, we had to pool individuals of all dance fly species for our analysis. It is of course possible that species differ in the rate of deposition of cuticle, although the theoretical link between temperature and band deposition makes it unlikely that any such difference is a cause of bias in our study. Nevertheless, it would be useful to supplement our data with collections of more known age flies in the future to test this possibility.

466 It is also possible that the deposition of cuticle on apodemes ceases in the adult at a 467 particular age, which is known to occur in several Drosophila species (Johnson and 468 Ellison 1982), and that the age at which this occurs varies across species and 469 individuals. Our data in figure 2 show a linear relationship of band number on age, with 470 no identifiable upper limit to age resolution (if the number of bands were saturating, we 471 might have expected a nonlinear relationship between age and band number). However 472 only one fly was kept alive for ten days, and it exhibited nine bands on its posterior 473 apodeme. Whether band deposition continues beyond the maximum of nine we 474 observed is unclear. However, in the vast majority of flies used to measure egg 475 development, individuals had fewer than nine bands and so were unlikely to be older 476 than the maximum age that this method may provide. Furthermore, the histograms 477 describing the distributions of bands in wild flies were not left-skewed (as might be 478 expected if there were many individuals at or near an upper bound of band number), 479 which suggests that most wild females were still accumulating bands at the time of 480 capture (see figure 3).

481

482 The validation of an ageing technique for further groups of wild insects may be 483 important to future investigations in evolutionary ecology. Estimates of age and 484 longevity of wild populations are often the focus of studies of natural and sexual 485 selection (Endler et al., 1986). The apodeme band ageing technique may successfully 486 be applied in studies examining differences in the age structure of the sexes. For 487 example, male dance flies are hypothesized to incur higher mortality rates than females 488 due to the male sex-specific behavior of acquiring nuptial gifts (Murray 2015). 489 However, the extent to which sexual differences may incur survival costs in other 490 contexts remains unclear (e.g., due to the risk of entanglement for ornamented females

491	in spider webs, for example (Gwynne et al., 2015)). The consequences of sex
492	differences in mortality could be evaluated by comparing the age profiles of the sexes
493	in more wild populations.

- 494
- 495

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- 502
- 503 Supplementary Figures S1
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645	Tables

Table 1: The empidinae species and counts of individuals caught in the emergence traps.

Species	Number Caught
Rhamphomyia longipes	5
Rhamphomyia flava	1
Hilara rejecta	5
Hilara manicata	2
Hilara litorea	1
Hilara intermidia	2
Hilara hirta	1
Hilara fulvibarba	2
Hilara clypeata	7
Hilara chorica	1
Hilara apta	6
Empis albinervis	1

647

648 Table 2: The intra-class correlation (ICC) for thoracic apodeme band counts on three different apodemes

Apodeme Type	ICC	Lower 95% CI	Upper 95% CI	Ν
Rostral Anterior Apodeme	0.8431	0.7104	0.9181	34
Caudal Anterior Apodeme	0.4344	0.1156	0.6730	33
Posterior Apodeme	0.4560	0.0.965	0.7118	26
Maximum Band Number	0.8468	0.6343	0.9411	17

types and for the maximum band number obtained from any apodeme type per fly.

650

Table 3: Parameter estimates describing the effect of age, mating status and species on egg development,

652 X denotes interaction terms. The model uses treatment contrasts to compare categories; the reference

653 levels are *Empis aestiva* and mated flies (see Schielzeth 2010). This model explained a significant

fraction of the variation in natural log standardised egg sizes (mm^2) (R square (adj) = 0.1756, p-value =

 $655 \quad 0.0002, n = 118$).

Source	Estimate	SE	t-value	p-value
Intercept	-0.7761	0.4074	-1.905	0.0594
Species (R. crassirostris)	-0.0262	0.5391	-0.049	0.9613
Age	0.4599	0.1565	2.938	0.0040
Age X Species (R. crassirostris)	-0.2729	0.1708	-1.598	0.1129
Mating Status (Unmated)	0.3713	0.6185	0.600	0.5496
Mating Status (Unmated) X Species (R. crassirostris)	-1.2807	1.1416	-1.122	0.2643
Age X Mating Status (Unmated)	-0.5239	0.2365	-2.215	0.0288
Age X Mating Status (Unmated) X Species (<i>R. crassirostris</i>)	0.5946	0.2904	2.047	0.0430

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657

658 Figure Legends

659	
660	Figure 1: The sexual dimorphism of Empis aestiva (Top) and Rhamphomyia crassirostris (Bottom).
661	Males and females are shown in full, as are their disembodied legs (Photo credit Frederick Hunter).
662	
663	Figure 2: The association between dance fly age and greatest apodeme band number. Lines represent
664	back transformed model predictions, and shaded area shows 95% confidence intervals. Transparency and
665	small random horizontal deviations have been added to the points to facilitate visualisation of
666	overlapping points.
667	
668	Figure 3: The range and frequency of age estimates of wild caught R. crassirostris (left) and E. aestiva
669	(right).
670	
671	Figure 4: The egg development of mated and unmated females of E. aestiva (left), and R. crassirostris
672	(right). Lines represent back-transformed model predictions, with shaded areas showing 95% CIs. Small
673	deviations have been added to data points to reduce overlap.
674	
675	Figure S1: The location of the eclosion line and apodeme bands on (top) rostral anterior apodeme,
676	(middle) caudal anterior apodeme and (bottom) anterior apodeme.
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