



## Role of CH<sub>4</sub> oxidation, production and transport in forest soil CH<sub>4</sub> flux

M.A. Bradford<sup>a,b,\*</sup>, P. Ineson<sup>a,1</sup>, P.A. Wookey<sup>c</sup>, H.M. Lappin-Scott<sup>b</sup>

<sup>a</sup>*Institute of Terrestrial Ecology, Merlewood Research Station, Windermere Road, Grange-over-Sands, Cumbria LA11 6JU, UK*

<sup>b</sup>*University of Exeter, Department of Biological Sciences, Prince of Wales Road, Exeter, Devon EX4 4PS, UK*

<sup>c</sup>*Uppsala University, Institute of Earth Sciences, Physical Geography, Villavägen 16, S-752 36 Uppsala, Sweden*

Received 17 April 2000; received in revised form 30 January 2001; accepted 27 February 2001

---

### Abstract

Forest soils are an important sink for atmospheric CH<sub>4</sub> but the contribution of CH<sub>4</sub> oxidation, production and transport to the overall CH<sub>4</sub> flux is difficult to quantify. It is important to understand the role these processes play in CH<sub>4</sub> dynamics of forest soils, to enable prediction of how the size of this sink will respond to future environmental change. Methane oxidation, production and transport were investigated for a temperate forest soil, previously shown to be a net CH<sub>4</sub> consumer, to determine the extent to which physical and biological processes contributed to the net flux. The sum of oxidation rates for soil layers were significantly greater ( $P < 0.05$ ) than for the intact soil cores from which the layers were taken. Combined with the immediate inhibition of CH<sub>4</sub> uptake on waterlogging soils, the findings suggested that soil CH<sub>4</sub> diffusion was an important regulator of CH<sub>4</sub> uptake. In support of this, a subsurface maximum for CH<sub>4</sub> oxidation was observed, but the exact depth of the maximum differed when rates were calculated on a mass or on an areal basis. Markedly varying potential CH<sub>4</sub> uptake activities between soil cores were masked in intact core rates. Potential CH<sub>4</sub> oxidation conformed well to Michaelis–Menten kinetics but  $V_{max}$ ,  $K_t$  and  $a_A^{\circ}$  values varied with depth, suggesting different functional methanotrophic communities were active in the profile. The presence of monophasic kinetics in fresh soil could not be used to infer that the soil was exposed only to CH<sub>4</sub> mixing ratios  $\leq$  atmospheric, as challenging soils with 20% CH<sub>4</sub> in air did not induce low-affinity oxidation kinetics. Atmospheric CH<sub>4</sub> oxidation potentials exceeded production potentials by 10–220 times. The results show that the forest soil CH<sub>4</sub> flux was dominated by CH<sub>4</sub> oxidation and transport, methanogenesis played only a minor role. © 2001 Elsevier Science Ltd. All rights reserved.

*Keywords:* Methane oxidation; Methane consumption; Forest soil; Methane transport; Kinetics

---

### 1. Introduction

Atmospheric concentrations of CH<sub>4</sub>, an important greenhouse gas (Prinn, 1994), have increased by approximately 2.45 fold since the Industrial Revolution (IPCC, 1995). Unsaturated soils represent the only net biological sink for atmospheric CH<sub>4</sub> and any effects on the size of this sink will markedly affect the rate of change in atmospheric CH<sub>4</sub> concentration (Reeburgh et al., 1993). In peatlands the CH<sub>4</sub> flux from the soil surface is often reported to be the resultant of CH<sub>4</sub> oxidation, CH<sub>4</sub> production and CH<sub>4</sub> transport within the soil (Moore and Dalva, 1997), and the same assumption is applicable to unsaturated soils. It is important to understand and quantify the role these three processes play in the CH<sub>4</sub> dynamics of unsaturated soils, to enable

prediction of how the size of this sink will respond to future environmental change.

The characteristics of the CH<sub>4</sub> oxidising and producing communities and factors which affect these characteristics and CH<sub>4</sub> transport determine the magnitude of the surface CH<sub>4</sub> flux. The ‘characteristics’ include the potential activity and depth distribution of CH<sub>4</sub> oxidation and production and the factors may include events such as inorganic N fertilisation and changes in soil water content. For example, inorganic N additions can decrease CH<sub>4</sub> oxidation potentials (Gulledge and Schimel, 1998) and similar additions in the field have decreased soil surface CH<sub>4</sub> uptake (Stuedler et al., 1989). Similarly, forest soils consume less CH<sub>4</sub> as the soil is made wetter (Castro et al., 1994) but rather than an inhibition of the soil CH<sub>4</sub> oxidation potential, the effect may be the result of slower CH<sub>4</sub> diffusion to the site of oxidation or a stimulation of CH<sub>4</sub> production. Slower CH<sub>4</sub> diffusion would be an important factor if a sub-surface maximum for CH<sub>4</sub> oxidation exists within the soil (Kruse et al., 1996), as CH<sub>4</sub> has to diffuse down through the soil and does so 10<sup>4</sup> times slower through water than air (Whalen et al., 1990). To

---

\* Corresponding author: NERC Centre for Population Biology, Imperial College at Silwood Park, Ascot, Berks SL5 7PY, UK.

*E-mail address:* m.a.bradford@ic.ac.uk (M.A. Bradford).

<sup>1</sup> Present address: The Department of Biology, University of York, PO Box 373, York, YO10 5YW, UK.

determine whether changes in CH<sub>4</sub> production or rate of CH<sub>4</sub> diffusion are most important in reduced net soil CH<sub>4</sub> uptake in wetted soils, determination of the potential activities and vertical distribution of oxidation and production within the soil is useful.

The kinetic parameters for soil CH<sub>4</sub> oxidation also provide valuable information regarding the dynamics of CH<sub>4</sub> oxidation and production within a soil profile. By quantifying the kinetics, Bender and Conrad (1992) suggested that it could be determined whether a soil was exposed only to atmospheric and sub-atmospheric concentrations of CH<sub>4</sub>, with high-affinity activity then dominating. The additional presence of low-affinity activity (Bender and Conrad, 1992, 1995; Conrad, 1996) would imply that a significant CH<sub>4</sub> source was present within the soil, possibly due to biological CH<sub>4</sub> production, as has been observed for many peat systems (Hall et al., 1996).

Field studies showed a forest soil to be a consistent net CH<sub>4</sub> oxidiser across 1 year with the rate of CH<sub>4</sub> uptake primarily controlled by soil water potential (Bradford et al., 2001). Our purpose was to determine the extent to which CH<sub>4</sub> oxidation, production and transport contributed to the observed temporal changes in net CH<sub>4</sub> uptake of this soil. To achieve this the vertical distribution of CH<sub>4</sub> oxidation potentials and kinetic parameters within the soil profile were characterised to determine the depth of maximum oxidation and whether the CH<sub>4</sub> mixing ratio experienced *in situ* exceeded atmospheric concentrations. Oxidation kinetics were determined for soils experimentally challenged with 20% CH<sub>4</sub> in air to assess the validity of using kinetics to infer *in situ* mixing ratios. The potential of the soil to produce CH<sub>4</sub> was assessed by waterlogging intact soil cores and incubating sieved soil under an O<sub>2</sub> free atmosphere. In addition, it was hypothesised that if soil CH<sub>4</sub> diffusion was an important determinant of soil CH<sub>4</sub> flux, then the sum of the soil CH<sub>4</sub> fluxes for undisturbed soil layers would be greater than the CH<sub>4</sub> flux rate of an intact (i.e. not divided into soil layers) soil core.

## 2. Methods and materials

### 2.1. Soil

Soil cores were collected in 25 cm tall PVC cylinders (15 cm dia.) from Perridge Forest (NGR SX 869908), a temperate mixed deciduous woodland consisting predominantly of mature (ca. 80 years old) oak (*Quercus robur* L.). The soil was a freely draining, low-base status, acidic brown earth, mapped within the Denbigh 1 Association (Findlay et al., 1984). For a full site and soil description see Bradford et al. (2001).

### 2.2. Determining CH<sub>4</sub> flux

Methane flux for sieved soil (to 2 mm) was determined using a closed chamber technique, and for soil cores or

undisturbed (i.e. non-sieved/unbroken) soil core layers using an open chamber technique. All experiments were performed at 20°C.

For the closed chamber technique, CH<sub>4</sub> concentrations were first standardised within a Wheaton bottle (120 ml) containing 10 g fresh weight (gfw) soil. A headspace gas sample was taken immediately upon sealing of the bottle. A second headspace sample was taken after 1 h, except for the kinetic determinations where repeated sampling over time was used. Although the soils follow first-order reaction kinetics (see Section 3 of this paper), a two-point rate calculation over a 1 h incubation provided rates representative of those calculated using rate constants derived from log-transformed time course data (M. A. Bradford, unpublished PhD thesis, Exeter University, 1999). Methane concentrations were determined on a Perkin Elmer Autosystem XL GC fitted with a FID operated at 200°C. Methane was separated isothermally on a Porapak Q (50–80 mesh) 2.5 m packed stainless steel column, with N<sub>2</sub> carrier gas flowing at 20 ml min<sup>-1</sup>. The detector response was calibrated using certified gas standards (British Oxygen Company, Special Gases, UK). Rates of CH<sub>4</sub> oxidation for sieved soils are expressed as nmol CH<sub>4</sub> consumed g<sup>-1</sup> dry weight (gdw) soil d<sup>-1</sup>.

For the open chamber technique, PVC cylinders containing soil cores were sealed at the base and capped with a Perspex lid with two air inlets and one air outlet. External ambient air, supplied via a single mixing-chamber to ensure all cores received input air with the same CH<sub>4</sub> concentrations, was drawn through the chamber headspaces at 50 ml min<sup>-1</sup>. The gas stream was automatically monitored for CH<sub>4</sub> by gas chromatography; for a full description of the gas analysis and data storage see Ineson et al. (1998).

### 2.3. CH<sub>4</sub> flux of intact soil cores, and undisturbed and sieved soil layers

This experiment was performed to determine: (1) the depth profile of soil CH<sub>4</sub> oxidation; (2) if soil CH<sub>4</sub> diffusion was an important determinant of soil CH<sub>4</sub> flux. Due to CH<sub>4</sub> flux sampling constraints, the experiment was performed in two halves (*A* + *B*). In parts *A* and *B*, CH<sub>4</sub> fluxes of intact soil cores were monitored and the cores were then split with a knife into defined soil horizons/layers (see below) and the CH<sub>4</sub> flux rates of these horizons/layers assessed. In part *A*, the intact cores were first split into the two constituent soil horizons, H and A. These two horizons were monitored, and then the H horizon was split into the H<sub>t</sub> (*t* = top; 4 cm deep) and H<sub>b</sub> (*b* = bottom; ca. 6 cm deep) soil layers and monitored. Three control cores were not split. In part *B*, intact cores were split into H<sub>b</sub> and A<sub>t</sub> (top 4 cm of A horizon) layers and monitored. Intact soil cores were analysed as controls. Lastly, to determine the depth profile of CH<sub>4</sub> oxidation for the upper 25 cm of soil, the H<sub>t</sub>, H<sub>b</sub>, A<sub>t</sub> and A<sub>b</sub> (bottom ca. 9 cm of A horizon) soil layers were sieved and their CH<sub>4</sub> flux rates determined using the described closed chamber technique. In addition, 5 gfw of the litter layer was finely cut

and analysed as for soil to determine CH<sub>4</sub> flux. Empty Wheaton bottles were analysed as blanks.

#### 2.4. Kinetics of CH<sub>4</sub> oxidation

Sieved soils from the H<sub>t</sub>, H<sub>b</sub> and A<sub>t</sub> layers of 20 cores were thoroughly mixed by layer and 10 gfw soil placed into Wheaton bottles. Six CH<sub>4</sub> oxidation rates were measured at increasing initial CH<sub>4</sub> mixing ratios in air (to 132 μl l<sup>-1</sup>) and from these rates the kinetic parameters  $K_t$  and  $V_{max}$  were determined using Lineweaver–Burk plots. Three replicates were sampled per time point for each initial CH<sub>4</sub> concentration; rate constants were calculated from log-transformed time course data. Incubation time increased from 2 to 10 h as the initial CH<sub>4</sub> concentration was increased, facilitating measurement of a marked and repeatable decrease in headspace CH<sub>4</sub> concentration. Soil water content was determined gravimetrically and maintained by deionised water addition. Control soils were not exposed to higher than ambient CH<sub>4</sub> mixing ratios; ambient oxidation rates of controls were compared with soils exposed to higher than ambient CH<sub>4</sub> concentrations to ensure oxidation activities did not change over the course of the experiment.

$V_{max}$  is defined here as the maximum specific uptake rate and  $K_t$  the half saturation concentration for uptake, after converting maximum uptake rates to units of CH<sub>4</sub> accumulated g<sup>-1</sup> soil f.w. h<sup>-1</sup>, similar to the methodology applied by Button (1991) for whole-cell uptake kinetics.  $V_{max}$  is reached at the substrate concentration where an organism can no longer increase its substrate utilisation rate and, thus, is a measure of the maximal rate of substrate uptake per unit of biomass.  $K_t$  is generally used as a measure for affinity of an organism for a substrate but is often misleading (Button, 1991) and so specific affinity ( $a_A^{\circ}$ ) was also determined.  $a_A^{\circ}$  is the best indicator of the ability of microorganisms to collect substrate from a dilute solution (Button, 1991) and is calculated from the relationship:  $a_A^{\circ} = V_{max}/K_t$  (Schut et al., 1993). Soil water CH<sub>4</sub> concentrations were calculated using the Ideal Gas Law and the Bunsen solubility coefficient for CH<sub>4</sub> at 20°C and 0‰ salinity (Yamamoto et al., 1976).

To determine the threshold ( $Th$ ) for CH<sub>4</sub> oxidation, the point at which the substrate is at too low a concentration to be assimilated, soil bottles initially at ambient CH<sub>4</sub> mixing ratios were sampled after 2, 3, 5 and 15 days.

To evaluate the CH<sub>4</sub> producing potential of the soil, four additional Wheaton bottles per soil layer were flushed with N<sub>2</sub> gas and the headspace CH<sub>4</sub> concentrations re-sampled after 5 days. Anaerobic indicator (Don Whitley Scientific Ltd, W. Yorkshire, UK) was used to establish whether O<sub>2</sub>-free conditions persisted in five control bottles containing soil.

Soil from H<sub>b</sub> and A<sub>t</sub> layers was autoclaved for 1 h at 121°C to determine if oxidation was a biological process. These soils were chosen because they exhibited the highest CH<sub>4</sub> oxidation potentials. Oxidation rates were assessed at

the start and end of the kinetic experiment and autoclaved, soil free Wheaton bottles were analysed as blanks.

To establish whether low-affinity CH<sub>4</sub> oxidising activity could develop when the soil was exposed to high CH<sub>4</sub> mixing ratios, A<sub>t</sub> layer soil (chosen because it represented the maximum for CH<sub>4</sub> oxidation on a volume basis) was challenged with 20% CH<sub>4</sub> in air for 40 days and the kinetic parameters  $V_{max}$ ,  $K_t$  and  $a_A^{\circ}$  determined as above. For the 40 days incubation, four replicates of 130 gfw soil aliquots in 1.7 l Kilner jars were used for the ambient and elevated CH<sub>4</sub> treatments. Soils were exposed to laboratory air for 15 min every 2–3 days to restore O<sub>2</sub> concentrations to those in ambient air and then headspace CH<sub>4</sub> concentrations were readjusted to restore treatments. Soil water content was determined gravimetrically and maintained by deionised water addition.

#### 2.5. Waterlogging soil cores

The potential for soil to produce CH<sub>4</sub> under waterlogged conditions was determined by comparing CH<sub>4</sub> flux from four waterlogged soil cores and four non-waterlogged soil cores (control). Soils were monitored to quantify baseline CH<sub>4</sub> flux rates prior to treatment assignment, to enable blocking in the experimental design. Waterlogged cores were maintained in polythene buckets filled with deionised water to the height of the top of the soil core. The height of the water was maintained for 30 days during which soil CH<sub>4</sub> flux was periodically sampled.

#### 2.6. Statistical analysis

All data analyses and statistical comparisons were performed using SAS (SAS Institute, 1988). Repeated measures blocked ANOVA were used to assess the effects of waterlogging cores and of splitting intact cores and soil horizons. Paired t-tests, or blocked ANOVA without repeated measures, were performed for sieved soil experiments and for comparing means for split core units assessed for CH<sub>4</sub> flux at separate times. Frequency distributions of the CH<sub>4</sub> fluxes or model residuals were tested for normality ( $\alpha = 0.1$ ) using the Shapiro–Wilk test. Non-normal data were normalised by log<sub>10</sub>-transformation prior to analysis. Coefficients of variation (CV) were calculated for CH<sub>4</sub> flux data in the core splitting experiments to provide a measure of the spatial heterogeneity of flux rates.

### 3. Results

#### 3.1. CH<sub>4</sub> flux of intact soil cores, and undisturbed and sieved soil layers

The sum of net CH<sub>4</sub> oxidation rates for constituent soil layers of whole soil cores and H horizons were significantly greater ( $P < 0.05$ ) than the measured oxidation rates of intact whole soil cores and intact whole H horizons (Fig. 1). When

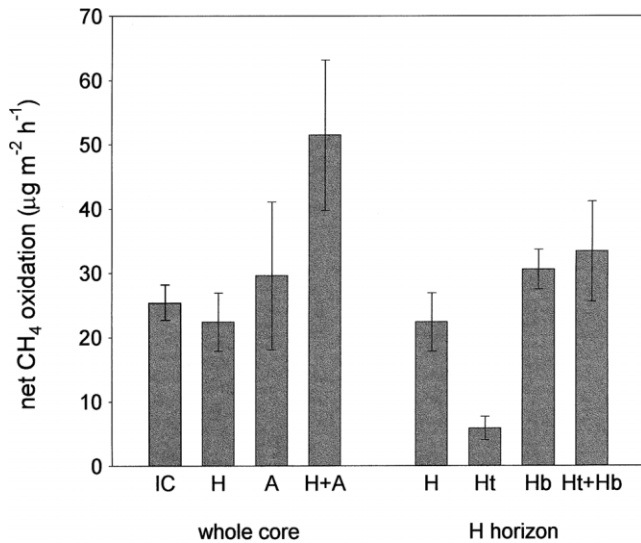


Fig. 1. Net CH<sub>4</sub> oxidation of intact soil core (IC) and undisturbed H horizon and the individual and summed fluxes from the constituent soil layers (H, A; H<sub>t</sub>, H<sub>b</sub>). The sum of the fluxes from the constituent layers (H + A; H<sub>t</sub> + H<sub>b</sub>) were significantly greater ( $P < 0.05$ ) than the fluxes from the soil units from which they were taken (IC; H). Flux rates are means  $\pm 1$  SEM ( $n = 3$ ). H<sub>t</sub>, top of H horizon; H<sub>b</sub>, bottom of H horizon.

other soil layers apart from H<sub>b</sub> and A<sub>t</sub> were discarded (Part B), an increased CH<sub>4</sub> consumption for the sum of these two layers over the intact core rate was observed (data not shown), but the increase was not significant ( $P = 0.09$ ). No changes in flux rates of parallel intact control cores were observed and, thus, the shift in rates was associated with the splitting of cores and horizons. High potential CH<sub>4</sub> oxidation rates in one layer of a soil core corresponded with high potential oxidation in another layer of the same core (data not shown); the same was true for low potential rates.

The CVs for net CH<sub>4</sub> oxidation rates across replicates markedly increased as soil cores were split into horizons and then into layers (Table 1).

Except for the litter layer, all sieved soil layers consistently oxidised CH<sub>4</sub> under the study conditions (Fig. 2). Change in headspace CH<sub>4</sub> concentrations in litter layer

Table 1

Coefficients of variation (CV) for net CH<sub>4</sub> oxidation rates of intact soil cores (IC), undisturbed H and A horizons and the constituent soil layers (A<sub>t</sub>, H<sub>t</sub>, H<sub>b</sub>) of the horizons, ( $n = 3$ ). The experiment was performed in two halves (A + B). CVs increased as soils were broken down from intact cores into horizons and layers (A<sub>t</sub>, top of A horizon; H<sub>t</sub>, top of H horizon; H<sub>b</sub>, bottom of H horizon)

Soil layer	CV part A	CV part B
IC	19.5	19.8
H	34.6	ND
A	27.4	ND
A <sub>t</sub>	ND	39.1
H <sub>t</sub>	66.5	ND
H <sub>b</sub>	41.1	56.4

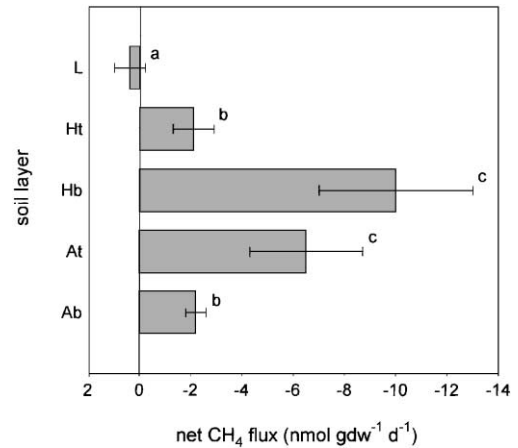


Fig. 2. Depth profile of potential net CH<sub>4</sub> flux (net CH<sub>4</sub> oxidation is shown by negative values). Bars marked with different letters (a–c) have significantly different flux rates ( $P < 0.05$ ). Changes in headspace CH<sub>4</sub> concentrations in soil free bottles were not significantly different from changes in L layer bottles ( $P > 0.05$ ). Flux rates are means  $\pm 1$  SEM ( $n = 4$ ). L, litter layer; H<sub>t</sub> and H<sub>b</sub>, top and bottom of H horizon; A<sub>t</sub> and A<sub>b</sub>, top and bottom of A horizon.

bottles were not significantly different to those for soil free bottles ( $P > 0.05$ ) and were significantly lower ( $P < 0.05$ ) than for all other soil layers. The H<sub>b</sub> and A<sub>t</sub> soil layers had the highest oxidation potentials, which were significantly greater ( $P < 0.05$ ) than the measured potentials for the H<sub>t</sub> and A<sub>b</sub> layers. Although sieved H<sub>b</sub> layer soil had a higher oxidation potential than sieved A<sub>t</sub> layer soil (Fig. 2), there was no significant difference between the rates ( $P > 0.05$ ). On a mass basis in undisturbed soil, oxidation potentials were also highest for H<sub>b</sub> soil and this was significant ( $P < 0.05$ ; Table 2), but on an areal basis, the maximum potential CH<sub>4</sub> oxidation occurred in A<sub>t</sub> soil ( $P < 0.05$ ; Table 2). Oxidation potentials on a mass basis for H<sub>b</sub> and A<sub>t</sub> soil increased when assessed for sieved rather than undisturbed soil (Table 2).

### 3.2. Kinetics of CH<sub>4</sub> oxidation

Potential CH<sub>4</sub> oxidation in H<sub>t</sub>, H<sub>b</sub> and A<sub>t</sub> soils conformed

Table 2

Net CH<sub>4</sub> oxidation by area (µg CH<sub>4</sub> m<sup>-2</sup> h<sup>-1</sup>) and by mass (nmol CH<sub>4</sub> gdw<sup>-1</sup> d<sup>-1</sup>) of undisturbed and sieved soil layers (H<sub>b</sub>, A<sub>t</sub>;  $n = 4$ ). (Values linked by different superscript letters are significantly different within the same row ( $P < 0.05$ ). Oxidation was significantly greater by mass for sieved A<sub>t</sub> soil than undisturbed A<sub>t</sub> soil ( $P < 0.05$ ); no significant difference between sieved and undisturbed H<sub>b</sub> soil was observed. (H<sub>b</sub>, bottom of H horizon; A<sub>t</sub>, top of A horizon)

Soil layer	H <sub>b</sub>	A <sub>t</sub>
Net CH <sub>4</sub> oxidation $\pm 1$ SEM		
By area (undisturbed soil)	25.6 $\pm$ 3.3 <sup>a</sup>	35.4 $\pm$ 3.1 <sup>b</sup>
By mass (undisturbed soil)	4.9 $\pm$ 0.1 <sup>a</sup>	1.1 $\pm$ 0.6 <sup>b</sup>
By mass (sieved soil)	10.0 $\pm$ 3.0	6.5 $\pm$ 2.2

Table 3

Kinetic parameters ( $V_{\max}$ ,  $a_A^{\circ}$ ,  $K_t$ ,  $Th$ ) of potential net CH<sub>4</sub> oxidation for fresh soil layers (H<sub>t</sub>, H<sub>b</sub>, A<sub>t</sub>) and the ratio of potential atmospheric CH<sub>4</sub> oxidation to potential CH<sub>4</sub> production ( $n = 3$ ) (H<sub>t</sub>, top of H horizon; H<sub>b</sub>, bottom of H horizon; A<sub>t</sub>, top of A horizon)

Soil layer (parameter)	H <sub>t</sub>	H <sub>b</sub>	A <sub>t</sub>
$V_{\max}$ (nmol gdw <sup>-1</sup> h <sup>-1</sup> )	0.7	13.0	0.3
$a_A^{\circ}$ (l gdw <sup>-1</sup> h <sup>-1</sup> )	0.02	0.19	0.03
$K_t$ (nM or μl l <sup>-1</sup> ) <sup>a</sup>	36 or 25	69 or 48	9 or 6
$Th^b$ (μl l <sup>-1</sup> ) ± 1 SEM	0.14 ± 0.03	0.04 ± 0.03	0.04 ± 0.02
Oxidation/production	10	221	105

<sup>a</sup> nM in soil water or μl l<sup>-1</sup> in headspace.

<sup>b</sup>  $P > 0.05$ ; no significant difference between  $Th$  values was observed.

well to typical Michaelis–Menten kinetics ( $r^2 > 0.98$  in all cases). The  $V_{\max}$ ,  $K_t$  and  $a_A^{\circ}$  values all markedly varied with depth, with the highest values being observed in the H<sub>b</sub> layer soil (Table 3). Although  $V_{\max}$  and  $K_t$  values were higher in the H<sub>t</sub> than A<sub>t</sub> soil, the  $a_A^{\circ}$  was greater in the A<sub>t</sub> than H<sub>t</sub> soil (Table 3). There was no significant difference ( $P > 0.05$ ) in  $Th$  values between soils (Table 3).

Oxidation potentials exceeded production potentials in all soils (Table 3). Highest CH<sub>4</sub> production potentials were observed in H<sub>t</sub> soil and had a mean (±1 SEM) rate of  $4.4 \pm 0.4$  pmol CH<sub>4</sub> gdw<sup>-1</sup> h<sup>-1</sup>, which was significantly greater ( $P < 0.05$ ) than the production potentials for H<sub>b</sub> and A<sub>t</sub> soils. The highest CH<sub>4</sub> production rate, combined with the lowest potential CH<sub>4</sub> oxidation, resulted in H<sub>t</sub> soil having the lowest ratio of oxidation to production (Table 3). H<sub>b</sub> soil produced significantly more CH<sub>4</sub> than A<sub>t</sub> soil ( $P < 0.05$ ; data not shown), but the greater oxidation potential in the lower organic layer resulted in a much greater ratio of potential oxidation to production (Table 3).

Ambient oxidation rates in control soils and soils exposed to CH<sub>4</sub> concentrations up to 132 μl l<sup>-1</sup> did not significantly differ ( $P > 0.05$ ) at the start or end of the kinetic experiments (data not shown). The net CH<sub>4</sub> oxidation potential for autoclaved soil was significantly lower than that for non-autoclaved control soil, 24 h and 14 days after autoclaving ( $P < 0.001$ ). Change in headspace CH<sub>4</sub> concentrations for autoclaved soil bottles were not significantly different ( $P > 0.05$ ) from those in soil free bottles (blanks; data not shown). The complete inhibition of CH<sub>4</sub> oxidation through autoclaving confirmed that oxidation within the soil was biologically mediated, as reported by Crill (1991) and Bender and Conrad (1992, 1995).

### 3.3. Kinetics of CH<sub>4</sub> oxidation for CH<sub>4</sub>-enriched soil

Both  $V_{\max}$  and  $K_t$  values for soil CH<sub>4</sub> uptake were greater after exposure to a 20%, rather than an ambient, CH<sub>4</sub> headspace, but the  $a_A^{\circ}$  remained approximately the same (Table 4). There was no significant difference between the ambient CH<sub>4</sub> oxidation rate of the two treatments ( $P > 0.05$ ), the mean values being equal (Table 4).

Table 4

Kinetic parameters ( $V_{\max}$ ,  $a_A^{\circ}$ ,  $K_t$ ,  $Th$ ) and rates of potential atmospheric CH<sub>4</sub> oxidation (pmol gdw<sup>-1</sup> h<sup>-1</sup>) for A<sub>t</sub> layer soil incubated with ambient and 20% CH<sub>4</sub> in the headspace for 40 days ( $n = 4$ )

Soil incubation (parameter)	Ambient CH <sub>4</sub>	20% CH <sub>4</sub>
$V_{\max}$ (nmol gdw <sup>-1</sup> h <sup>-1</sup> )	0.4	0.7
$a_A^{\circ}$ (l gdw <sup>-1</sup> h <sup>-1</sup> )	0.04	0.04
$K_t$ (nM or μl l <sup>-1</sup> ) <sup>a</sup>	13 or 9	25 or 17
Oxidation at ambient ± 1 SEM <sup>b</sup>	60 ± 1	60 ± 1

<sup>a</sup> nM in soil water or μl l<sup>-1</sup> in headspace.

<sup>b</sup>  $P > 0.05$ ; no significant treatment effect was detected.

### 3.4. Waterlogging soil cores

Net CH<sub>4</sub> oxidation rates were significantly greater ( $P < 0.01$ ) in control than waterlogged cores, with mean uptake rates (±1 SEM) of  $-30 \pm 3$  and  $-2 \pm 1$  μg CH<sub>4</sub> m<sup>-2</sup> h<sup>-1</sup> across the 30 d period, respectively. Net CH<sub>4</sub> oxidation dropped immediately on waterlogging and no consistent emissions of CH<sub>4</sub> were observed over the 30 d study (data not shown).

## 4. Discussion

### 4.1. CH<sub>4</sub> flux of intact soil cores, and undisturbed and sieved soil layers

Kruse et al. (1996) predicted that CH<sub>4</sub> diffusion was an important controller of CH<sub>4</sub> oxidation rate in forest soils similar to Perridge. We hypothesised that if CH<sub>4</sub> supply to oxidisers was important then the sum of the CH<sub>4</sub> uptake rates for soil horizons and soil layers would be greater than the oxidation rates for the intact cores or H horizons from which they were taken. Greater uptake was observed and, also, oxidation potentials were greater in sieved than undisturbed H<sub>b</sub> and A<sub>t</sub> soils. As well as improved CH<sub>4</sub> supply to methanotrophs as soils were broken down into horizons, then layers and then sieved, O<sub>2</sub> supply would also have improved. Due to the fact that Mancinelli (1995) proposed that O<sub>2</sub> is not likely to be limiting in systems where the CH<sub>4</sub> source is atmospheric, we suggest that the increased oxidation was caused by increased CH<sub>4</sub> supply.

Variability between net CH<sub>4</sub> oxidation rates for separated soil units increased as they were divided from cores, to horizons, to layers (Table 1). Thus, the full spatial heterogeneity in potential CH<sub>4</sub> oxidation rates for specific soil layers was not exhibited in intact core CH<sub>4</sub> uptake rates. We propose that environmental constraints, such as soil water status, regulate net CH<sub>4</sub> uptake so efficiently that markedly different potential uptake activities between cores are masked. Thus, in the field, soil methanotrophic activity must operate far below its optimum due to environmental constraints. Such constraints may, in part, explain

the prolonged recovery of methanotrophic activity after soil disturbances (Ojima et al., 1993).

The high rates of net CH<sub>4</sub> oxidation in the H<sub>b</sub> and A<sub>t</sub> layers, when compared to the H<sub>t</sub> layer, suggest that net CH<sub>4</sub> consumption in the Perridge soil is mainly driven by lower soil layers (i.e. below the H<sub>t</sub> layer). This corresponds with the findings of many workers who report distinct subsurface maxima for potential CH<sub>4</sub> oxidation in temperate and boreal forest soils. The greatest activity has been found either in the uppermost mineral layers (Crill, 1991; Koschorreck and Conrad, 1993; Saari et al., 1997), or, as in our study, in the lower organic and the upper mineral layers (Adamsen and King, 1993; Czepiel et al., 1995; Kruse et al., 1996; Amaral and Knowles, 1997; Saari et al., 1997).

Our study showed that a deciduous litter layer did not exhibit net methanotrophic activity. This finding is supported by Saari et al. (1997) who observed no CH<sub>4</sub> uptake by coniferous litters and even net CH<sub>4</sub> production.

The depth of maximum CH<sub>4</sub> oxidation potential differed when potentials were calculated on a mass or area basis (Table 2). The higher by mass potential of the H<sub>b</sub> layer suggests that it has a larger, or more active, methanotrophic community per gram of soil than the A<sub>t</sub> layer. However, the A<sub>t</sub> layer had higher potential oxidising activity by area (or volume). Roslev et al. (1997), using isotopically labelled CH<sub>4</sub> as a biomarker, demonstrated that the depth of maximum CH<sub>4</sub> assimilation within intact soil cores corresponded to the depth of the maximum CH<sub>4</sub> oxidation potential by volume. This suggests that the A<sub>t</sub> layer in the Perridge soil would dominate CH<sub>4</sub> oxidation in intact soil profiles and questions the ecological relevance of determining depth profiles of CH<sub>4</sub> oxidation potential using only sieved soil (Saari et al., 1997).

#### 4.2. Kinetics of CH<sub>4</sub> oxidation

All fresh soil layers exhibited Michaelis–Menten saturation curves, typical of high-affinity methanotrophic activity (Bender and Conrad, 1992). Indeed, the values of  $K_t$  and  $Th$  observed by us were within or just below the range reported by Conrad (1996) for soils where the atmosphere was the largest source of CH<sub>4</sub> ( $K_t$ : 20–200 nM;  $Th$ : 30–700 pM  $\equiv$  21–480 nl l<sup>-1</sup> under the conditions of our study).

The characteristics of the functional methanotrophic community changed with depth. The highest  $V_{max}$  and  $K_t$  values were measured in the H<sub>b</sub> layer, as were the highest methanotrophic potentials on a mass basis. The highest  $a_A^\circ$  was also observed in the H<sub>b</sub> layer, suggesting that the methanotrophs in this layer were the most efficient scavengers of CH<sub>4</sub> (Button, 1991). Only  $Th$  values were roughly equivalent between the three layers, indicating that the methanotrophs within each layer were all capable of oxidising CH<sub>4</sub> at extremely low partial pressures.

#### 4.3. Kinetics of CH<sub>4</sub> oxidation for CH<sub>4</sub>-enriched soil

The observed kinetic values and the monophasic form of

the kinetics means that low-affinity methanotrophic activity was not present in the Perridge soil. Bender and Conrad (1992), after determining similar kinetic profiles for the A<sub>b</sub> horizons from a number of sites, proposed that such soils were exclusively exposed to CH<sub>4</sub> mixing ratios below atmospheric values. Bender and Conrad (1992) only observed biphasic kinetics, the second phase approximating to low-affinity kinetics, when soils had been exposed to concentrations of CH<sub>4</sub> significantly higher than atmospheric. In contrast, low-affinity CH<sub>4</sub> oxidising activity was not induced when our soil was challenged with 20% CH<sub>4</sub> in air. Our observation opens to question whether the detection of only high-affinity kinetics within a soil can be used to reliably predict that high concentrations of CH<sub>4</sub> do not develop within that soil and, thus, that CH<sub>4</sub> production is insignificant.

The  $V_{max}$  and  $K_t$  of the high-affinity activity markedly increased after incubation of soils under a 20% CH<sub>4</sub> headspace, as observed by Bender and Conrad (1992), but the  $a_A^\circ$  remained unchanged. This suggests that the methanotrophic community increased in size during exposure to elevated CH<sub>4</sub> but that the methanotroph community did not undergo a functional shift. Despite the increase in  $V_{max}$  and  $K_t$  observed in our study, the atmospheric CH<sub>4</sub> oxidising potential for the CH<sub>4</sub>-enriched soils did not increase. This suggests that CH<sub>4</sub> uptake was still limited by factors other than the ‘true potential’ of the soil methanotrophic community for CH<sub>4</sub> oxidation. Thus, methods commonly used to assess oxidation potentials of soils do not provide optimal conditions for CH<sub>4</sub> uptake by soil methanotrophs. Physical conditions, such as soil moisture, may still influence CH<sub>4</sub> supply to an extent that efficiently masks differences in potential methanotrophic activity. This should be considered when oxidation potentials are reported.

#### 4.4. Waterlogging soil cores

The immediate drop in net CH<sub>4</sub> oxidation rates of soil cores on waterlogging suggests that the inhibition was largely a physical process. This is consistent with the hypothesis that the regulation of CH<sub>4</sub> uptake by soil water status is mediated by diffusional limitation of CH<sub>4</sub> to the methanotrophs, rather than due to stimulation of methanogenic activity.

#### 4.5. Conclusion

Net CH<sub>4</sub> flux in the Perridge soil was dominated by CH<sub>4</sub> oxidation, methanogenesis played a minor role. The data support the contention that rate of soil CH<sub>4</sub> diffusion to the site of CH<sub>4</sub> oxidation explains the strong regulation of net CH<sub>4</sub> oxidation by soil water potential at this site (Bradford et al., 2001). The full spatial heterogeneity in potential CH<sub>4</sub> oxidation rates for specific soil layers was not exhibited in intact core CH<sub>4</sub> uptake rates. Based on this observation we suggest that environmental constraints regulate net CH<sub>4</sub> uptake so efficiently as to mask markedly different potential

uptake activities in intact soils. The H<sub>b</sub> layer contained the most active methanotrophic community and was best able to oxidise CH<sub>4</sub> at low substrate concentrations. However, the A<sub>t</sub> layer soil exhibited the greatest oxidation potential for an equivalent area and on the basis of the work by Roslev et al. (1997), the A<sub>t</sub> layer dominated *in situ* CH<sub>4</sub> oxidation. Our study demonstrates that by using a suite of approaches, that include potential activity, depth profile and kinetic studies, important insights into the CH<sub>4</sub> dynamics of a soil can be obtained. If these approaches are conducted in isolation care must be taken in the interpretation of the data. The insights generated from using a suite of approaches can be used to more confidently predict how and why soil CH<sub>4</sub> flux may respond to future environmental change, such as altered soil moisture.

### Acknowledgements

The authors thank the UK Natural Environment Research Council for funding and Sir Henry Studholme for use of Perridge Forest.

### References

- Adamsen, A.P.S., King, G.M., 1993. Methane consumption in temperate and subarctic forest soils: rates, vertical zonation, and responses to water and nitrogen. *Applied and Environmental Microbiology* 59, 485–490.
- Amaral, J.A., Knowles, R., 1997. Localization of methane consumption and nitrification activities in some boreal forest soils and the stability of methane consumption on storage and disturbance. *Journal of Geophysical Research—Atmospheres* 102, 29255–29260.
- Bender, M., Conrad, R., 1992. Kinetics of CH<sub>4</sub> oxidation in oxic soils exposed to ambient air or high CH<sub>4</sub> mixing ratios. *FEMS Microbiology Ecology* 101, 261–270.
- Bender, M., Conrad, R., 1995. Effect of CH<sub>4</sub> concentrations and soil conditions on the induction of CH<sub>4</sub> oxidation activity. *Soil Biology & Biochemistry* 27, 1517–1527.
- Bradford, M.A., Wookey, P.A., Ineson, P., Lappin-Scott, H.M., 2001. Controlling factors and effects of chronic nitrogen and sulphur deposition on methane oxidation in a temperate forest soil. *Soil Biology & Biochemistry* 33, 93–102.
- Button, D.K., 1991. Biochemical basis for whole-cell uptake kinetics: specific affinity, oligotrophic capacity, and the meaning of the Michaelis constant. *Applied and Environmental Microbiology* 57, 2033–2038.
- Castro, M.S., Melillo, J.M., Steudler, P.A., Chapman, J.W., 1994. Soil moisture as a predictor of methane uptake by temperate forest soils. *Canadian Journal of Forest Research* 24, 1805–1810.
- Conrad, R., 1996. Soil microorganisms as controllers of atmospheric trace gases (H<sub>2</sub>, CO, CH<sub>4</sub>, OCS, N<sub>2</sub>O and NO). *Microbiological Reviews* 60, 609–640.
- Crill, P.M., 1991. Seasonal patterns of methane uptake and carbon dioxide release by a temperate woodland soil. *Global Biogeochemical Cycles* 5, 319–334.
- Czepiel, P.M., Crill, P.M., Harriss, R.C., 1995. Environmental factors influencing the variability of methane oxidation in temperate zone soils. *Journal of Geophysical Research—Atmospheres* 100, 9359–9364.
- Findlay, D.C., Colborne, G.J.N., Cope, D.W., Harrod, T.R., Hogan, D.V., Staines, S.J., 1984. Soils and their use in south west England. Soil Survey of England and Wales. Bulletin No. 14. Harpenden, Herts, UK.
- Gulledge, J., Schimel, J.P., 1998. Low-concentration kinetics of atmospheric CH<sub>4</sub> oxidation in soil and mechanism of NH<sub>4</sub><sup>+</sup> inhibition. *Applied and Environmental Microbiology* 64, 4291–4298.
- Hall, G.H., Simon, B.M., Pickup, R.W., 1996. CH<sub>4</sub> production in blanket peat bog: A procedure for sampling, sectioning and incubating samples whilst maintaining anaerobic conditions. *Soil Biology & Biochemistry* 28, 9–15.
- Ineson, P., Coward, P.A., Hartwig, U.A., 1998. Soil gas fluxes of N<sub>2</sub>O, CH<sub>4</sub> and CO<sub>2</sub> beneath *Lolium perenne* under elevated CO<sub>2</sub>: the Swiss free air carbon dioxide enrichment experiment. *Plant and Soil* 198, 89–95.
- IPCC, 1995. In: Houghton, J.T., Meira Filho, L.G., Callander, B.A., Harris, N., Kattenberg, A., Maskell, K. (Eds.). *The Science of Climate Change. Contribution of Working Group 1 to the Second Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge.
- Koschorreck, M., Conrad, R., 1993. Oxidation of atmospheric methane in soil: Measurements in the field, in soil cores and in soil samples. *Global Biogeochemical Cycles* 7, 109–121.
- Kruse, C.W., Moldrup, P., Iversen, N., 1996. Modelling diffusion and reaction in soils—II: Atmospheric methane diffusion and consumption in a forest soil. *Soil Science* 161, 355–365.
- Mancinelli, R.L., 1995. The regulation of methane oxidation in soil. *Annual Reviews of Microbiology* 49, 581–605.
- Moore, T.R., Dalva, M., 1997. Methane and carbon dioxide exchange potentials of peat soils in aerobic and anaerobic laboratory incubations. *Soil Biology & Biochemistry* 29, 1157–1164.
- Ojima, D.S., Valentine, D.W., Mosier, A.R., Parton, W.J., Schimel, D.S., 1993. Effect of land use change on methane oxidation in temperate forest and grassland soils. *Chemosphere* 26, 675–685.
- Prinn, R.G., 1994. Global atmospheric–biospheric chemistry. In: Prinn, R.G. (Ed.). *Global Atmospheric–Biospheric Chemistry*. Plenum Press, New York, pp. 1–18.
- Reeburgh, W.S., Whalen, S.C., Alperin, M.J., 1993. The role of methylo-trophy in the global methane budget. In: Murrell, J.C., Kelly, D.P. (Eds.). *Microbial Growth on C1 Compounds*. Intercept Ltd, UK, pp. 1–14.
- Roslev, P., Iversen, N., Henriksen, K., 1997. Oxidation and assimilation of atmospheric methane by soil methane oxidizers. *Applied and Environmental Microbiology* 63, 874–880.
- Saari, A., Martikainen, P.J., Ferm, A., Ruuskanen, J., de Boer, W., Troelstra, S.R., Laanbroek, H.J., 1997. Methane oxidation in soil profiles of Dutch and Finnish coniferous forests with different soil texture and atmospheric nitrogen deposition. *Soil Biology & Biochemistry* 29, 1625–1632.
- SAS Institute, 1988. *SAS/STAT™ User's Guide Release 6.03 Edition*. SAS Institute Inc, Cary, NC, USA.
- Schut, F., de Vries, E.J., Gottschal, J.C., Robertson, B.R., Harder, W., Prins, R.A., Button, D.K., 1993. Isolation of typical marine bacteria by dilution culture: Growth, maintenance, and characteristics of isolates under laboratory conditions. *Applied and Environmental Microbiology* 59, 2150–2160.
- Steudler, P.A., Bowden, R.D., Melillo, J.M., Aber, J.D., 1989. Influence of nitrogen fertilisation on methane uptake in temperate forest soils. *Nature* 341, 314–316.
- Whalen, S.C., Reeburgh, W.S., Sandbeck, K.A., 1990. Rapid methane-oxidation in a landfill cover soil. *Applied and Environmental Microbiology* 56, 3405–3411.
- Yamamoto, S., Alcauskas, J.B., Crozier, T.E., 1976. Solubility of methane in distilled water and seawater. *Journal of Chemical and Engineering Data* 21, 78–80.